

(19) World Intellectual Property Organization International Bureau





(43) International Publication Date 21 February 2002 (21.02.2002)

PCT

(10) International Publication Number WO 02/14358 A2

(51) International Patent Classification⁷: C07K 14/00

(21) International Application Number: PCT/US01/21124

(22) International Filing Date: 30 July 2001 (30.07.2001)

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data:

60/224,642 60/241,779 11 August 2000 (11.08.2000) US 19 October 2000 (19.10.2000) US

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- (81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC,

LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.

(84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Declarations under Rule 4.17:

- as to applicant's entitlement to apply for and be granted a patent (Rule 4.17(ii)) for the following designations AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG)
- as to the applicant's entitlement to claim the priority of the earlier application (Rule 4.17(iii)) for all designations
- as to the applicant's entitlement to claim the priority of the earlier application (Rule 4.17(iii)) for all designations

Published:

 without international search report and to be republished upon receipt of that report

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

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(54) Title: NOVEL SECRETED PROTEINS AND THEIR USES

(57) Abstract: The present invention provides nucleic acid sequences encoding novel human proteins. These novel nucleic acids are useful for constructing the claimed DNA vectors and host cells of the invention and for preparing the claimed nucleic acids, recombinant proteins and antibodies that are useful in the claimed methods and medical uses.

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NOVEL SECRETED PROTEINS AND THEIR USES

FIELD OF THE INVENTION

This application claims priority of Provisional Applications Serial No. 60/224,642 filed August 11, 2000, and Serial No. 60/241,779 filed October 19, 2000.

The present invention relates to the identification and isolation of novel DNA, therapeutic and drug discovery uses, and the recombinant production of novel secreted polypeptides, designated herein as LP105, LP061, LP224, LP240, LP239(a), LP243(a), LP243(b), LP253, LP218, LP251(a), LP252, LP239(b), LP223(a), LP255(a), LP244, LP186, LP251(b), LP255(b), and LP223(b). The present invention also relates to vectors, host cells, and antibodies directed to these polypeptides.

BACKGROUND OF THE INVENTION

Extracellular proteins play an important role in the formation, differentiation and maintenance of multi-20 cellular organisms. The fate of many individual cells, e.g., proliferation, migration, differentiation, or interaction with other cells, is typically governed by information received from other cells and/or the immediate environment. This information is often transmitted by 25 secreted polypeptides (for instance, mitogenic factors, survival factors, cytotoxic factors, differentiation factors, neuropeptides, and hormones) which are, in turn, received and interpreted by diverse cell receptors or membrane-bound proteins. These secreted polypeptides or 30 signaling molecules normally pass through the cellular secretory pathway to reach their site of action in the extracellular environment.

Secreted proteins have various industrial applications, including pharmaceuticals, diagnostics, biosensors and bioreactors. Most protein drugs available at present such as thrombolytic agents, interferons, interleukins, erythropoietins, colony stimulating factors, and various other cytokines are secretory proteins. Their receptors, which are membrane proteins, also have potential as therapeutic or diagnostic agents.

The present invention describes the cloning and characterization of novel proteins, termed LP105, LP061, LP224, LP240, LP239(a), LP243(a), LP243(b), LP253, LP218, LP251(a), LP252, LP239(b), LP223(a), LP255(a), LP244, LP186, LP251(b), LP255(b), and LP223(b), as well as active variants and/or fragments thereof.

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SUMMARY OF THE INVENTION

The present invention provides isolated LP105, LP061, LP224, LP240, LP239(a), LP243(a), LP243(b), LP253, LP218, LP251(a), LP252, LP239(b), LP223(a), LP255(a), LP244, LP186, LP251(b), LP255(b), and LP223(b) polypeptide encoding nucleic acids and the polypeptides encoded thereby, including fragments and/or specified variants thereof. Contemplated by the present invention are LP probes, primers, recombinant vectors, host cells, transgenic animals, chimeric antibodies and constructs, LP polypeptide antibodies, as well as methods of making and using them diagnostically and therapeutically as described and enabled herein.

The present invention includes isolated nucleic acid molecules comprising polynucleotides that encode LP105, LP061, LP224, LP240, LP239(a), LP243(a), LP243(b), LP253, LP218, LP251(a), LP252, LP239(b), LP223(a), LP255(a), LP244,

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LP186, LP251(b), LP255(b), and LP223(b) polypeptides as defined herein, as well as fragments and/or specified variants thereof, or isolated nucleic acid molecules that are complementary to polynucleotides that encode such LP polypeptides, or fragments and/or specified variants thereof as defined herein.

A polypeptide of the present invention includes an isolated LP polypeptide comprising at least one fragment, domain, or specified variant of at least 90-100% of the contiguous amino acids of at least one portion of SEQ ID NO:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, or 38.

The present invention also provides an isolated LP polypeptide as described herein, wherein the polypeptide further comprises at least one specified substitution, insertion, or deletion corresponding to portions or specific residues of SEQ ID NO:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, or 38.

The present invention also provides an isolated nucleic acid probe, primer, or fragment, as described herein, wherein the nucleic acid comprises a polynucleotide of at least 10 nucleotides, corresponding or complementary to at least 10 nucleotides of SEQ ID NO:1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, or 37.

The present invention also provides compositions, including pharmaceutical compositions, comprising an LP polypeptide, an LP polypeptide-encoding polynucleotide, an LP polynucleotide, and/or an LP polypeptide antibody, wherein the composition has a measurable effect on an activity associated with a particular LP polypeptide as disclosed herein. A method of treatment or prophylaxis based on an LP polypeptide associated activity as disclosed

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herein can be effected by administration of one or more of the polypeptides, nucleic acids, antibodies, vectors, host cells, transgenic cells, and/or compositions described herein to a mammal in need of such treatment or 5 prophylactic. Accordingly, the present invention also includes methods for the prophylaxis or treatment of a patho-physiological condition in which at least one cell type involved in said condition is sensitive or responsive to an LP polypeptide, LP polypeptide-encoding 10 polynucleotide, LP nucleic acid, LP polypeptide antibody, host cell, transgenic cell, and/or composition of the present invention.

The present invention also provides an article of manufacture comprising a container, holding a composition effective for treating a condition disclosed herein, and a label.

The present invention also provides a method for identifying compounds that bind an LP polypeptide, comprising:

- a) admixing at least one isolated LP polypeptide as described herein with a test compound or composition; and
- b) detecting at least one binding interaction between the polypeptide and the compound or composition, optionally further comprising detecting a change in biological activity, such as a reduction or increase.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

Applicants have identified cDNA clones comprising polynucleotides that encode novel polypeptides or novel variants of known polypeptides:

1) LP105

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LP105 polypeptides comprising the amino acid sequence of the open reading frame encoded by the polynucleotide sequence as shown in SEQ ID NO:1 are contemplated as one embodiment of the present invention. Specifically, polypeptides of the present invention comprise the amino acid sequence as shown in SEQ ID NO:2, as well as fragments, variants, and derivatives thereof. Accordingly, LP105 polynucleotides encoding the LP105 polypeptides of the present invention are also contemplated by the present invention. LP105 polynucleotides are predominantly expressed in prostate and kidney tissues.

The LP105 polypeptide as shown in SEQ ID NO:2 shares sequence similarity with a recently reported human lefty 15 protein (WO 99/09198). Lefty proteins are members of the TGF-beta family and two LEFTY genes, LEFTY A and B, have been localized by FISH to 1q42, a region syntenic to the location to which the mouse Lefty genes have been mapped at 1H5 [Kosaki, et al., Am. J. Hum. Genet. 64(3):712-21 (1999); 20 Meno, et al., Genes Cells 2(8):513-24 (1997)]. LEFTY A is identical to EBAF [Kothapalli, et al., J. Clin. Invest. 99(10):2342-50 (1997)]. Analysis of 126 human cases of L-R axis malformation showed a single nonsense and a single missense mutation in the LEFTY A gene. Both mutations lay in 25 the cysteine-knot region of the LEFTY A protein and the phenotype of affected individuals was very similar to that typically seen in Lefty-1 -/- mice with L-R axis malformations. Furthermore, the LP105 polypeptide as shown in SEQ ID NO:2 shares sequence similarity with human bone morphogenic protein 18 (WO 99/29718). Compositions 30 comprising LP105 polypeptides, polynucleotides, and/or antibodies are useful for the treatment of defects in or

wounds to tissues including, but not limited to, epidermis, nerve, muscle, cardiac muscle, and organs including, but not limited to liver, lung, epithelium, brain, spleen, cardiac, pancreas and kidney. Furthermore, compositions comprising LP105 polypeptides, polynucleotides, and/or antibodies can 5 be useful for modulating sexual development, pituitary hormone production, hematopoiesis, wound healing, tissue repair, and the formation of bone and cartilage. Compositions comprising LP105 polypeptides, polynucleotides, and/or antibodies can also be used to treat such conditions 10 as cancer including, but not limited to prostate and kidney cancer, interstitial lung disease, infectious diseases, autoimmune diseases, arthritis, leukemia, lymphomas, immunosuppression, immunity, humoral immunity, inflammatory bowel disease, myelosuppression, periodontal disease, 15 osteoarthritis, osteoporosis, and other abnormalities of bone, cartilage, muscle, tendon, ligament, meniscus, and/or other connective tissues as well as dysfunctional growth and differentiation patterns of cells.

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2) LP061

In another embodiment, polypeptides comprising the amino acid sequence of the open reading frame encoded by the polynucleotide sequence as shown in SEQ ID NO:3 are contemplated by the present invention. Specifically, polypeptides of the present invention comprise the amino acid sequence as shown in SEQ ID NO:4, as well as fragments, variants, and derivatives thereof. Accordingly, LP061 polynucleotides encoding LP061 polypeptides of the present invention are also contemplated by the present invention. LP061 polynucleotides are predominantly expressed in diseased thyroid. The present invention includes a human

nucleotide sequence (Incyte clone 2719035H1) as shown in SEQ ID NO:3 which appears to be a shortened splice-variant of the published human fukutin nucleotide sequence (GenBank AB008226).

5 Fukuyama-type congenital muscular dystrophy (FCMD), one of the most common autosomal recessive disorders in Japan (incidence is 0.7 to 1.2 per 10,000 births), is characterized by congenital muscular dystrophy, in conjunction with brain malformation (micropolygria). The 10 FCMD gene was mapped to a region of less than 100 kilobases which included the marker locus D9S2107 on human chromosome 9q31. The mutation responsible for FCMD is a retrotransposal insertion of tandemly repeated sequences within this candidate-gene in all FCMD chromosomes carrying the founder 15 haplotype (87%). The inserted sequence is about 3 kilobases long and is located in the 3' untranslated region of a gene encoding a new 461 amino acid protein. This novel gene, termed fukutin, is expressed in heart, brain, skeletal muscle, pancreas and lymphoblasts in normal individuals, but 20 not in FCMD patients who carry the insertion. Two independent point mutations confirm that mutation of this gene is responsible for FCMD. The predicted fukutin protein contains an amino-terminal signal sequence and one glycosylation site, which together with results from 25 transfection experiments suggests that fukutin is a secreted protein. Abnormalities in basal lamina in FCMD muscle and brain have been seen by electron microscopy [Nakano, et al., Acta Neuropathol. 91(3):313-21 (1996); Ishii, et al., Neuromuscul. Disord. 7(3):191-7 (1997)]. Therefore it has 30 been suggested that secreted fukutin may be associated with the extracellular matrix surrounding expressing cells where it forms a complex with other extracellular components to

stabilize a microenvironment supportive for normal cellular/tissue function. In muscle, fukutin complexes might stabilize the function of muscle fibers/sarcomeres; in brain, fukutin may assist the migration of neuronal precursors during cortical development [Fukuyama, et al., Brain Dev. 3(1):1-29 (1981).

Accordingly, compositions comprising LP061 polypeptides, polynucleotides, and/or antibodies are useful for diagnosis, treatment and intervention of diseases, disorders, and/or conditions including, but not limited to, 10 brain malformation (micropolygria), Alzheimer's Disease, Parkinson's Disease, Huntington's Disease, Tourette Syndrome, ALS, muscular pathologies including, but not limited to, muscular dystrophies including, but not limited to, congenital muscular dystrophies such as fukuyama-type 15 congenital muscular dystrophy, meningitis, encephalitis, demyelinating diseases, peripheral neuropathies, neoplasia, trauma, spinal cord injuries, ischemia and infarction, aneurysms, hemorrhages, schizophrenia, mania, dementia, 20 paranoia, obsessive compulsive disorder, panic disorder learning disabilities, psychoses, autism, and altered behaviors, including disorders in feeding, sleep patterns, balance, and perception.

25 **3) LP224**

In another embodiment, polypeptides comprising the amino acid sequence of the open reading frame encoded by the polynucleotide sequence as shown in SEQ ID NO:5 are contemplated by the present invention. Specifically, polypeptides of the present invention comprise the amino acid sequence as shown in SEQ ID NO:6, as well as fragments, variants, and derivatives thereof. Accordingly, LP224

polynucleotides encoding LP224 polypeptides of the present invention are also contemplated by the present invention.

The gene encoding the LP224 polypeptide has been localized to chromosome 4p16 (GenBank hit g7022852) and the LP224 polypeptide as shown in SEQ ID NO:6 shares sequence 5 similarity with a protein encoded by the leucine-rich, glioma inactivated 1 tumor suppressor gene. LGI1 has the highest homology with a number of transmembrane and extracellular proteins which function as receptors and adhesion proteins. LGI1 is predominantly expressed in 10 neural tissues, especially in brain; its expression is reduced in low grade brain tumors and it is significantly reduced or absent in malignant gliomas. Its localization to the 10q24 region, and rearrangements or inactivation in malignant brain tumors, suggest that LGI1 is a candidate 15 tumor suppressor gene involved in progression of glial tumors [Chernova, et al., Oncogene 17(22):2873-81 (1998)]. Accordingly, compositions comprising LP224 polypeptides, polynucleotides, and/or antibodies are useful for diagnosis, treatment and intervention of bipolar affective disorder 20 [Ewald, et al., Mol. Psychiatry 3(5):442-8 (1998)], hearing defects [Van Camp, et al., J. Med. Genet. 36(7):532-6, (1999)], cherubism [Mangion, et al., Am. J. Hum. Genet. 65(1):151-7 (1999)], Wolf-Hirschhorn (WH) syndrome [Zollino, et al., Am. J. Med. Genet. 82(5):371-5 (1999); Ann. Genet. 25 41(2):73-6 (1998)], Ellis-van Creveld syndrome [EVC; Polymeropoulos, et al., Genomics 35(1):1-5 (1996)], autosomal dominant postaxial polydactyly, nail dystrophy, and dental abnormalities [Howard, et al., Am. J. Hum. Genet. 61(6):1405-12 (1997)], Pitt-Rogers-Danks (PRD) syndrome and 30 overgrowth syndrome [Partington, et al., J. Med. Genet. 34(9):719-28 (1997)], bladder cancer and malignant brain

tumors [Bell, et al., Genes Chromosomes Cancer 17(2):108-17 (1996)].

4) LP240

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In another embodiment, polypeptides comprising the 5 amino acid sequence of the open reading frame encoded by the polynucleotide sequence as shown in SEQ ID NO:7 are contemplated by the present invention. Specifically, polypeptides of the present invention comprise the amino acid sequence as shown in SEQ ID NO:8, as well as fragments, 10 variants, and derivatives thereof. Accordingly, LP240 polynucleotides encoding LP240 polypeptides of the present invention are also contemplated by the present invention. The gene encoding the LP240 polypeptide as shown in SEQ ID NO:8 has been localized to chromosome 19. 15

LP240 polynucleotides, polypeptides, and antibodies corresponding to LP240 are useful for diagnosis, treatment and intervention of diseases, disorders, and/or conditions including, but not limited to, stroke, Alzheimer's disease, Parkinson's disease, Huntington's disease, neurodegenerative disorders, brain cancer, cardiovascular disease, atherosclerosis, myocardial infarction, inflammation, and rheumatoid arthritis.

5) LP239(a) and LP239(b) 25

In another embodiment, polypeptides comprising the amino acid sequences of the open reading frames encoded by the polynucleotide sequences as shown in SEQ ID NO:9 and SEQ ID NO:23 are contemplated by the present invention. Specifically, LP239(a) and LP239(b) polypeptides of the present invention comprise the amino acid sequences as shown in SEQ ID NO:10 and SEQ ID NO:24, respectively, as

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well as fragments, variants, and derivatives thereof.

Accordingly, LP239(a) and LP239(b) polynucleotides

comprising polynucleotides as identified in SEQ ID NO:9 and

SEQ ID NO:23, respectively, are also contemplated by the

present invention.

LP239 polypeptide-encoding polynucleotide sequences are primarily expressed in digestive, hemic, immune, and nervous system tissues. The LP239(a) and LP239(b) polypeptides as shown in SEQ ID NO:10 and SEQ ID NO:24 share sequence similarity with a thyroxine-binding globulin [see AF204929; Mori, et al., Endocr. J. 46(4):613-9 (1999)]. LP239 polypeptides therefore may serve as hormone binding proteins in serum, protease inhibitors, or hormones when administered to patients suffering from a deficiency of endogenous LP239 polypeptides. LP239 polynucleotides, polypeptides, and antibodies corresponding to LP239 are useful for diagnosis, treatment and intervention of diseases, disorders, and/or conditions including, but not limited to, hypothyroidism, anemia, sepsis, gram negative bacteremia, allergic responses, allergic autoimmune diseases, type 1 diabetes, Th1-dependent insulitis, inflammation, multiple sclerosis, rheumatoid arthritis, inflammatory bowel disease, liver failure, ARDS, cancers, leukemia, and immunodeficiency.

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6) LP243(a) and LP243(b)

In another embodiment, polypeptides comprising the amino acid sequences of the open reading frames encoded by the polynucleotide sequences as shown in SEQ ID NO:11 and SEQ ID NO:13 are contemplated by the present invention. Specifically, LP243(a) and LP243(b) polypeptides of the present invention comprise the amino acid sequences as shown

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in SEQ ID NO:12 and SEQ ID NO:14, respectively, as well as fragments, variants, and derivatives thereof. Accordingly, LP243(a) and LP243(b) polynucleotides comprising polynucleotides as identified in SEQ ID NO:11 and SEQ ID NO:13 are also contemplated by the present invention.

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The gene encoding the disclosed LP243 polypeptides have been localized to chromosome 19p13.3 (GenBank hit g2894631) and is mainly expressed in nervous system and digestive system. The LP243 polypeptides as shown in SEQ ID NO:12 and SEQ ID NO:14 share sequence similarity with the amino acid 10 sequence of protein PRO227 disclosed in WO 99/14328, the amino acid sequence of the human Tango-79 protein disclosed in WO 99/06427, the glioma amplified on chromosome 1 (GAC1) protein (AF030435), and the Rattus norvegicus (AF133730) 15 SLIT protein. GAC1 is a member of the leucine-rich repeat superfamily on chromosome band 1q32.1. GAC1 is amplified and overexpressed in malignant gliomas [Almeida, et al., Oncogene 16(23):2997 (1998)]. In Drosophila, at least, SLIT proteins are believed to be involved in axon pathway 20 development. The two predicted proteins LP243(a) and LP243(b) as shown in SEQ ID NO:12 and 14, respectively, have different N-terminals but are identical starting from amino acid 27 through amino acid 557 as shown in SEQ ID NO:12. The present invention provides isolated LP243(a) and LP243(b) polypeptides as described herein, wherein the 25 polypeptide has at least one activity, such as, but not limited to, inducing cellular proliferation, tumorigenesis, synapse formation, neurotransmission, learning, cognition, homeostasis, neuronal outgrowth, differentiation or survival, or tissue regeneration including but not limited 30 to neural tissue regeneration. An LP243 polynucleotide,

polypeptide, and/or antibody can thus be screened for a corresponding activity according to known methods.

The present invention also provides a composition comprising an isolated LP243 nucleic acid, polypeptide, 5 and/or antibody as described herein and a carrier or diluent. The carrier or diluent can optionally be pharmaceutically acceptable, according to known methods. LP243 polynucleotides are expressed in nervous tissues and the polypeptides encoded thereby appear to play a role in 10 tumorigenesis, neurodegenerative diseases, behavioral disorders, and/or inflammatory conditions. Accordingly, compositions comprising LP243 polypeptides, polynucleotides, and/or antibodies are useful for diagnosis, treatment and intervention of diseases, disorders, and/or conditions 15 including, but not limited to, cancer, including, but not limited to sporadic ovarian tumors [Wang, et al., Br. J. Cancer 80(1-2):70-2 (1999)], Peutz-Jeghers syndrome [Nakagawa, et al., Hum. Genet. 102(2):203-6 (1998); Olschwang, et al., J. Med. Genet. 35(1):42-4 (1998)], 20 adenoma malignum of the uterine cervix [Lee, et al., Cancer Res. 15;58(6):1140-3 (1998)], pancreatic carcinomas [Hoglund, et al., Genes Chromosomès Cancer 21(1):8-16 (1998)], multiple myeloma and plasma cell leukemia [Taniwaki, et al., Blood 84(7):2283-90 (1994)], Alzheimer's 25 Disease, Parkinson's Disease, Huntington's Disease, Tourette Syndrome, meningitis, encephalitis, demyelinating diseases, peripheral neuropathies, neoplasia, trauma, congenital malformations, spinal cord injuries, ischemia and infarction, aneurysms, hemorrhages, schizophrenia, mania, 30 dementia, paranoia, obsessive compulsive disorder, panic

disorder, learning disabilities, ALS, psychoses, autism, and

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altered behaviors, including disorders in feeding, sleep patterns, balance, and perception.

7) LP253

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Cytokines are secreted regulatory peptides that mediate 5 a wide range of biological activities by binding to specific cell surface receptors on target cells. Cytokine actions include control of cell proliferation and differentiation, regulation of hematopoiesis, and immune inflammatory responses. Cytokines are also major orchestrators of host 10 defense processes and, as such, are involved in responses to exogenous as well as endogenous insults and in repair or restoration of tissue integrity. In order for a cytokine to exert its effect on cells, it is now accepted by those skilled in the art that the molecule must interact with 15 molecules, located on cell membranes, referred to as receptors. Patents which exemplify disclosures of interleukin receptors include Honjo, et al., U.S. Patent 4,816,565; Urdal, et al., U.S. Patent 4,578,335; Dower, et al., U.S. Patent 5,180,812; and Taniguchi, et al., U.S. 20 Patent 5,198,359; and WO 2000/15759, the disclosures of which are incorporated by reference.

In another embodiment of the present invention, we provide polypeptides comprising the amino acid sequence of the open reading frame encoded by the polynucleotide sequence as shown in SEQ ID NO:15. Specifically, LP253 polypeptides of the present invention comprise the amino acid sequence as shown in SEQ ID NO:16, as well as fragments, variants, and derivatives thereof. Accordingly, LP253 polynucleotides comprising polynucleotides as identified in SEQ ID NO:15 are also contemplated by the present invention. LP253 encoding polynucleotide sequences WO 02/14358 PCT/US01/21124

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are primarily expressed in digestive, hemic, immune, and nervous system tissues. The LP253 polypeptide as shown in SEQ ID NO:16 shares structural similarity with other interleukin receptors.

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The effects of IL-1 in vivo can be regulated via the administration of a soluble form of its receptor [Fanslow, et al., Science 248:739-41 (1990)]. Systemic administration of a soluble, extracellular portion of the receptor for IL-1 (soluble IL-1R) had profound inhibitory effects on the development of in vivo alloreactivity. Survival of heterotopic heart allografts was prolonged from twelve days in controls to seventeen days in mice treated with soluble IL-1R. Lymph node hyperplasia in response to localized injection of allogeneic cells was completely blocked by soluble IL-1R treatment.

The availability of the purified LP253 polypeptide, in soluble form, presents therapeutic possibilities as well. The results that Fanslow report demonstrate the ability of a soluble cytokine receptor to modulate biological activity upon exogeneous administration in vivo, presumably by acting as a neutralizing agent for the endogeneously produced, corresponding ligand, and provides evidence of the therapeutic potential of soluble cytokine receptors in a variety of clinical disorders.

Administration of a soluble form of LP253 would interfere with the effect of its endogenous ligand on the cells, since the ligand would not bind to the endogenous membrane bound LP253 as freely. Hence, an aspect of the present invention is the treatment of pathological conditions caused by excess expression and/or activity of LP253 polypeptides by adding an amount of soluble LP253 polypeptides sufficient to inhibit binding of a cytokine to

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the aforementioned cells. This methodology can also be modified, and the soluble receptor can also be used as a screening agent for pharmaceuticals. Briefly, a pharmaceutical which works as an LP253 antagonist can do so by blocking the binding of endogenous ligand to the LP253. Prior to determining whether a material would be effective in vivo, one may use the purified LP253 polypeptide in connection with a potential pharmaceutical to determine if there is binding. If there is in fact binding, further testing may be indicated.

Expression of recombinant polypeptides in high levels and its use as an antigen allows production of additional neutralizing monoclonal and polyclonal antibodies. Such neutralizing antibodies can be used in *in vivo* model settings to elucidate the role that LP253 and its ligand play in normal as well as pathologic immune responses (i.e., disease states that are aggravated by activated T- and NK-cells like auto-immune diseases, graft versus host disease and rheumatoid arthritis). Thus, purified LP253 polypeptides, polynucleotides, and/or antibodies compositions will be useful in diagnostic assays for LP253 and its ligand, and also in raising antibodies to LP253 for use in diagnosis or therapy.

LP253 polypeptides, polynucleotides, and/or antibodies can be administered, for example, for the purpose of suppressing immune responses in a human. A variety of diseases or conditions are caused by an immune response to alloantigen, including allograft rejection and graft-versus-host reaction. In alloantigen-induced immune responses, soluble LP253 polypeptides may suppress lymphoproliferation and inflammation which result upon activation of T cells. Soluble LP253 polypeptides may therefore be used to

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effectively suppress alloantigen-induced immune responses in the clinical treatment of, for example, rejection of allografts (such as skin, kidney, and heart transplants), and graft-versus-host reactions in patients who have received bone marrow transplants.

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LP253 polypeptides, polynucleotides, and/or antibodies may also be used in clinical treatment of autoimmune dysfunctions, such a rheumatoid arthritis, diabetes and multiple sclerosis, which are dependent upon the activation of T cells against antigens not recognized as being indigenous to the host. LP253 polypeptides, polynucleotides, and/or antibodies may also be useful in treatment of septic shock in which interferon gamma produced in response to various interleukins plays a central role in causing morbidity and mortality [Doherty, et al., J. Immunol. In addition, compositions comprising 149:1666 (1992)]. soluble LP253 polypeptides may be used directly in therapy to bind or scavenge endogenous LP253 ligands, thereby providing a means for regulating the immune or inflammatory activities. In its use to prevent or reverse pathologic immune responses, soluble LP253 polypeptides can be combined with other cytokine antagonists such as antibodies to the other known interleukin receptors, soluble interleukin receptors, soluble TNF (tumor necrosis factor) receptors, and/or interleukin receptor antagonists, and the like.

The dose ranges for the administration of the LP253 polypeptides, polynucleotides, and/or antibodies and fragments thereof may be determined by those of ordinary skill in the art without undue experimentation. In general, appropriate dosages are those which are large enough to produce the desired effect, for example, blocking the binding of endogenous ligands of LP253 to endogenous LP253.

8) LP218

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In another embodiment, polypeptides comprising the amino acid sequence of the open reading frame encoded by the polynucleotide sequence as shown in SEQ ID NO:17 are contemplated by the present invention. Specifically, LP218 polypeptides of the present invention comprise the amino acid sequence as shown in SEQ ID NO:18, as well as fragments, variants, and derivatives thereof. Accordingly, LP218 polynucleotides comprising polynucleotides as identified in SEQ ID NO:17 are also contemplated by the present invention.

The gene encoding the LP218 polypeptide as shown in SEQ ID NO:18 has been localized to chromosome 22q11 and shares 15 sequence similarity with acetyl LDL receptor. LP218 polynucleotides are expressed in hemic, immune, reproductive, and urinary tract tissues. Accordingly, compositions comprising LP218 polypeptides, polynucleotides, and/or antibodies are useful for diagnosis, treatment and intervention of schizophrenia [Li, Mol. Psychiatry 5(1):77-20 84 (2000)], hypocalcemia [Garabedian, et al., Genet Couns. 10(4):389-94 (1999)], rhabdoid tumor [Zhou, et al., Gene 241(1):133-41 (2000)], DiGeorge/velo-cardio-facial syndrome [Amati, et al., Eur. J. Hum. Genet. 7(8):903-9 (1999)], congenital heart disease [Borgmann, et al., Eur. J. Pediatr. 25 158(12):958-63 (1999)], and abdominal lymphatic dysplasia [Mansir, et al., Genet. Couns. 10(1):67-70 (1999)], oesophageal atresia [Digilio, et al., J. Med. Genet. 36(2):137-9 (1999)].

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9) LP251(a) and LP251(b)

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In another embodiment, polypeptides comprising the amino acid sequence of the open reading frames encoded by the polynucleotide sequences as shown in SEQ ID NO:19 and SEQ ID NO:33 are contemplated by the present invention. Specifically, LP251(a) and LP251(b) polypeptides of the present invention comprise the amino acid sequence as shown in SEQ ID NO:20 and SEQ ID NO:34, respectively, as well as fragments, variants, and derivatives thereof. Accordingly, LP251(a) and LP251(b) polynucleotides comprising polynucleotides as identified in SEQ ID NO:19 or SEQ ID NO:33 are also contemplated by the present invention.

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The LP251 polypeptides as shown in SEQ ID NO:20 and SEQ ID NO:34 share sequence similarity with aqualysin I precursor, a subtilisin-type serine protease which is secreted into the culture medium by Thermus aquaticus YT-1, an extremely thermophilic gram-negative bacterium [Terada, et al., J. Biol. Chem. 265(12):6576-81 (1990)].

The gene encoding the disclosed polypeptides has been localized to chromosome 1p33-p34.3. Accordingly, 20 compositions comprising LP251 polypeptides, polynucleotides, and/or antibodies are useful for diagnosis, treatment and intervention of breast cancer [Emi, et al., Genes Chromosomes Cancer 26(2):134-41 (1999)], myelodysplastic syndromes including but not limited to thrombocytemia and abnormal megakaryopoiesis [Jondeau, et al., Leukemia 25 10(11):1692-5 (1996)], Schnyder's crystalline corneal dystrophy [Shearman, et al., Hum. Mol. Genet. 5(10):1667-72 1996), tumorigenesis including, but not limited to, colorectal cancer [Praml, et al., Oncogene 11(7):1357-62 30 (1995)], Schwartz-Jampel syndrome [Nicole, Hum. Mol. Genet. 4(9):1633-6 (1995), and autosomal dominant hypercholesterolemia [Varret, et al., Am. J. Hum. Genet. 64(5):1378-87 (1999)].

Furthermore, LP251 polynucleotides are mainly expressed in skin and liver. Accordingly, compositions comprising LP251 polypeptides, polynucleotides, and/or antibodies are generally useful for the treatment of defects in or wounds to tissues including, but not limited to skin and liver.

10) LP252

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In another embodiment, polypeptides comprising the amino acid sequence of the open reading frame encoded by the polynucleotide sequence as shown in SEQ ID NO:21 are contemplated by the present invention. Specifically, LP252 polypeptides of the present invention comprise the amino acid sequence as shown in SEQ ID NO:22, as well as fragments, variants, and derivatives thereof. Accordingly, LP252 polynucleotides comprising polynucleotides as identified in SEQ ID NO:21 are also contemplated by the present invention.

The LP252 polypeptide as shown in SEQ ID NO:22 shares 20 sequence similarity with thyroid hormone-induced protein B precursor. Furthermore, the gene encoding the disclosed polypeptide has been localized to chromosome 9p12-13. Accordingly, compositions comprising LP252 polypeptides, polynucleotides, and/or antibodies are useful for diagnosis, 25 treatment and intervention of ectrodactyly, ectodermal dysplasia and cleft lip/palate syndrome [Fukushima, et al., Clin. Genet. 44(1):50 (1993); Hasegawa, et al., Clin. Genet. 40(3):202-6 (1991)]; and cancer, including, but not limited to, kidney cancer and carcinoma of the respiratory tract 30 [Schraml, et al., J. Pathol. 190(4):457-61 (2000); Higashi, et al., Genes Chromosomes Cancer 3(1):21-3 (1991)].

11) LP223(a) and (b)

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In another embodiment, polypeptides comprising the amino acid sequences of the open reading frames encoded by 5 the polynucleotide sequences as shown in SEQ ID NO:25 and SEQ ID NO:37 are contemplated by the present invention. Specifically, LP223(a) and LP223(b) polypeptides of the present invention comprise the amino acid sequences as shown in SEQ ID NO:26 and SEQ ID NO:38, respectively, as well as 10 fragments, variants, and derivatives thereof. Accordingly, LP223(a) and LP223(b) polynucleotides comprising polynucleotides as identified in SEQ ID NO:25 and SEQ ID NO:37 are also contemplated by the present invention. LP223(a) and LP223(b) polypeptides as shown in SEQ ID NO:26 15 and SEQ ID NO:38 share sequence similarity with a human secreted protein sequence disclosed in WO 2000/04140.

The present invention also provides isolated LP223 polypeptides as described herein, wherein the polypeptides have at least one activity, such as, but not limited to, inducing cellular proliferation, synapse formation, neurotransmission, learning, cognition, homeostasis, neuronal outgrowth, differentiation or survival, or tissue regeneration including but not limited to neural tissue regeneration. An LP223 polynucleotide, polypeptide, and/or antibody can thus be screened for a corresponding activity according to known methods.

The present invention also provides a composition comprising an isolated LP223 nucleic acid, polypeptide, and/or antibody as described herein and a carrier or diluent. The carrier or diluent can optionally be pharmaceutically acceptable, according to known methods. LP223 polynucleotides and polypeptides are expressed in

nervous tissues and appear to play a role in neurodegenerative diseases, behavioral disorders, and/or inflammatory conditions. Accordingly, compositions comprising LP223 polypeptides, polynucleotides, and/or antibodies are useful for diagnosis, treatment and intervention of diseases, disorders, and/or conditions including, but not limited to, Alzheimer's Disease, Parkinson's Disease, Huntington's Disease, Tourette Syndrome, meningitis, encephalitis, demyelinating diseases, peripheral neuropathies, neoplasia, trauma, congenital 10 malformations, spinal cord injuries, ischemia and infarction, aneurysms, hemorrhages, schizophrenia, mania, dementia, paranoia, obsessive compulsive disorder, panic disorder learning disabilities, ALS, psychoses, autism, and altered behaviors, including disorders in feeding, sleep 15 patterns, balance, and perception. In addition, the gene or gene product may also play a role in the treatment and/or detection of developmental disorders associated with the developing embryo, sexually-linked disorders, or disorders 20 of the cardiovascular system.

12) LP255(a) and LP255(b)

In another embodiment, polypeptides comprising the amino acid sequences of the open reading frames encoded by the polynucleotide sequences as shown in SEQ ID NO:27 and SEQ ID NO:35 are contemplated by the present invention. Specifically, LP255(a) and LP255(b) polypeptides of the present invention comprise the amino acid sequences as shown in SEQ ID NO:28 and SEQ ID NO:36, respectively, as well as fragments, variants, and derivatives thereof. Accordingly, LP255(a) and LP255(b) polynucleotides comprising polynucleotides as identified in SEQ ID NO:27 and SEQ ID

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NO:35 are also contemplated by the present invention. The LP255(a) and LP255(b) polypeptides as shown in SEQ ID NO:28 and SEQ ID NO:36 share sequence similarity with a human secreted protein sequence disclosed in WO 99/14328 as PRO332.

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The present invention also provides isolated LP255 polypeptides as described herein, wherein the polypeptides have at least one activity, such as, but not limited to, promoting cell growth. An LP255 polynucleotide, polypeptide, and/or antibody can thus be screened for a corresponding activity according to known methods.

The present invention also provides a composition comprising an isolated LP255 nucleic acid, polypeptide, and/or antibody as described herein and a carrier or . 15 diluent. The carrier or diluent can optionally be pharmaceutically acceptable, according to known methods. LP255 polynucleotides and polypeptides are expressed in nervous tissues and appear to play a role in neurological Accordingly, compositions comprising LP255 disorders. 20 polypeptides, polynucleotides, and/or antibodies are useful for diagnosis, treatment and intervention of diseases, disorders, and/or conditions arising from Alzheimer's Disease, Parkinson's Disease, Huntington's Disease, Tourette Syndrome, meningitis, encephalitis, demyelinating diseases, 25 peripheral neuropathies, neoplasia, trauma, congenital malformations, spinal cord injuries, ischemia and infarction, aneurysms, hemorrhages, schizophrenia, mania, dementia, paranoia, obsessive compulsive disorder, panic disorder learning disabilities, ALS, psychoses, autism, and altered behaviors, including disorders in feeding, sleep patterns, balance, and perception.

13) LP244

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In another embodiment, polypeptides comprising the amino acid sequence of the open reading frame encoded by the polynucleotide sequence as shown in SEQ ID NO:29 are contemplated by the present invention. Specifically, LP244 polypeptides of the present invention comprise the amino acid sequence as shown in SEQ ID NO:30, as well as fragments, variants, and derivatives thereof. Accordingly, LP244 polynucleotides comprising polynucleotides as identified in SEQ ID NO:29 are also contemplated by the present invention.

The gene encoding the LP244 polypeptide has been localized to chromosome 19 and is predominantly expressed in embryonic structures. The LP244 polypeptide as shown in SEQ ID NO:30 appears to be a novel shortened splice-variant of 15 the human Epstein-Barr virus induced gene 3 (EBI3; GenBank accession no. NP_005746) wherein the first 67 amino acids of the LP244 polypeptide are identical to those of EBI3. EBI3 is reported to be a hematopoietin receptor family member related to the p40 subunit of interleukin-12 and to 20 the ciliary neurotrophic factor receptor, whose expression is induced in B lymphocytes by Epstein-Barr virus (EBV) infection [Devergne, et al., J. Virology 70(2):1143-53 (1996)]. The gene encodes a 34-kDa glycoprotein which lacks a membrane-anchoring motif and is secreted. Despite the 25 absence of a membrane-anchoring motif and of cysteines likely to mediate covalent linkage to an integral membrane protein, EBI3 is also present on the plasma membrane of EBVtransformed B lymphocytes and of transfected cells. Most newly synthesized EBI3 is retained in the endoplasmic reticulum in an endoglycosidase H-sensitive form associated with the molecular chaperone calnexin and with a novel 60-

kDa protein. EBI3 is expressed in vivo by scattered cells in interfollicular zones of tonsil tissue, by cells associated with sinusoids in perifollicular areas of spleen tissue, and at very high levels by placental syncytiotrophoblasts. EBI3 expression in vitro is induced in EBV-5 negative cell lines by expression of the EBV latent infection membrane protein-1 and in peripheral blood mononuclear cells by pokeweed mitogen stimulation. maps to chromosome 19p13.2/3, near genes encoding the 10 erythropoietin receptor and the cytokine receptor-associated kinase, Tyk2. EBI3 synthesis by trophoblasts and by EBVtransformed cells and similarities to interleukin-12 p40 are compatible with a role for EBI3 in regulating cell-mediated immune responses. LP244 therefore may function as a 15 modulator of EBI3 activity. Administration of a soluble form of LP244 would interfere with the effect of its endogenous ligand on the cells, since the ligand would not bind to the endogenous membrane bound LP244 as freely. Hence, an aspect of the present invention is the treatment 20 of pathological conditions caused by excessive expression of LP244 polypeptides by adding an amount of soluble LP244 polypeptides sufficient to inhibit binding of a cytokine to the aforementioned cells. This methodology can also be modified, and the soluble receptor can also be used as a 25 screening agent for pharmaceuticals. Briefly, a pharmaceutical which works as an LP244 antagonist can do so by blocking the binding of endogenous ligand to the LP244. Prior to determining whether a material would be effective in vivo, one may use the purified LP244 polypeptide in 30 connection with a potential pharmaceutical to determine if there is binding. If there is in fact binding, further testing may be indicated.

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Expression of recombinant polypeptides in high levels and its use as an antigen allows production of additional neutralizing monoclonal and polyclonal antibodies. Such neutralizing antibodies can be used in in vivo model settings to elucidate the role that LP244 and its ligand play in normal as well as pathologic immune responses (i.e., disease states that are aggravated by activated T- and NKcells like autoimmune diseases, graft versus host disease, and rheumatoid arthritis). Thus, purified LP244 polypeptides, polynucleotides, and/or antibodies compositions will 10 be useful in diagnostic assays for LP244 and its ligand, and also in raising antibodies to LP244 for use in diagnosis or therapy. More specifically, compositions comprising LP244 polypeptides, polynucleotides, and/or antibodies are useful for diagnosis, treatment and intervention of diseases, disorders, and/or conditions including, but not limited to, infectious diseases, hypothyroidism, anemia, sepsis, gram negative bacteremia, allergic responses, allergic autoimmune diseases, type 1 diabetes, Th1-dependent insulitis, inflammation, multiple sclerosis, rheumatoid arthritis, inflammatory bowel disease, liver failure, ARDS, cancers, leukemia, and immuno-deficiency, arthritis, leukemia, lymphomas, immuno-suppression, immunity, humoral immunity, myelosuppression, periodontal disease, and osteoarthritis.

LP244 polypeptides, polynucleotides, and/or antibodies can be administered, for example, for the purpose of suppressing immune responses in a human. A variety of diseases or conditions are caused by an immune response to alloantigen, including allograft rejection and graft-versus-In alloantigen-induced immune responses, host reaction. soluble LP244 polypeptides may suppress lymphoproliferation and inflammation which result upon activation of T cells.

Soluble LP244 polypeptides may therefore be used to effectively suppress alloantigen-induced immune responses in the clinical treatment of, for example, rejection of allografts (such as skin, kidney, and heart transplants), and graft-versus-host reactions in patients who have received bone marrow transplants.

LP244 polypeptides, polynucleotides, and/or antibodies may also be used in clinical treatment of autoimmune dysfunctions, such a rheumatoid arthritis, diabetes and multiple sclerosis, which are dependent upon the activation 10 of T cells against antigens not recognized as being indigenous to the host. LP244 polypeptides, polynucleotides, and/or antibodies may also be useful in treatment of septic shock in which interferon gamma produced in response to various interleukins plays a central role in causing morbidity and mortality [Doherty, et al., J. Immunol. 149:1666 (1992)]. In addition, compositions comprising soluble LP244 compositions may be used directly in therapy to bind or scavenge endogenous LP244 ligands, thereby providing a means for regulating the immune or inflammatory 20 In its use to prevent or reverse pathologic activities. immune responses, soluble LP244 polypeptides can be combined with other cytokine antagonists such as antibodies to the other known interleukin receptors, soluble interleukin receptors, soluble TNF (tumor necrosis factor) receptors, 25 and/or interleukin receptor antagonists, and the like.

14) LP186

In another embodiment, polypeptides comprising the amino acid sequence of the open reading frame encoded by the polynucleotide sequence as shown in SEQ ID NO:31 are contemplated by the present invention. Specifically, LP186

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polypeptides of the present invention comprise the amino acid sequence as shown in SEQ ID NO:32, as well as fragments, variants, and derivatives thereof. Accordingly, LP186 polynucleotides comprising polynucleotides as identified in SEQ ID NO:31 are also contemplated by the present invention.

The LP186 polypeptide as shown in SEQ ID NO:32 shares sequence similarity with the human delta homologue, DLL3 [Bulman, et al., Nature Genetics 24(4):438-41 (2000)]. Mutations in the human delta homologue, DLL3, cause axial 10 skeletal defects in spondylocostal dysostosis. Two of the mutations predict truncations within conserved extracellular domains. The third is a missense mutation in a highly conserved glycine residue of the fifth epidermal growth factor (EGF) repeat, which has revealed an important 15 functional role for this domain. Spondylocostal dysostosis (SD) is a group of vertebral malsegmentation syndromes with reduced stature resulting from axial skeletal defects. is characterized by multiple hemivertebrae, rib fusions and deletions with a non-progressive kyphoscoliosis. D113 is 20 mutated in the X-ray-induced mouse mutant pudgy (pu), causing a variety of vertebrocostal defects similar to SD phenotypes [Kusumi, et al., Nature Genetics 19(3):274-8 (1998)]. These mutations highlight the critical role of the 25 Notch signalling pathway and its components in patterning the mammalian axial.

LP186 polypeptide-encoding polynucleotide sequences are primarily expressed in the nervous system. Thus, polynucleotides, polypeptides, and antibodies corresponding to this gene are useful for diagnosis, treatment and intervention of diseases, disorders, and conditions of the nervous system. Thus, the present sequence represents a

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polypeptide which suppresses proliferation and differentiation of undifferentiated cells such as neurons and blood cells. The polypeptide may be used for the prevention and control of disorders involving undifferentiated cells, such as leukaemia and malignant tumours, and improvement of blood formation, e.g., after immunosuppression.

The polynucleotides and polypeptides of the present invention are designated herein as "LP polynucleotides" or "LP polypeptide-encoding polynucleotides" and "LP polypeptides." When immediately followed by a numerical designation (i.e., LP105), the term "LP" refers to a specific group of molecules as defined herein. A complete designation wherein the term "LP" is immediately followed by a numerical designation and a molecule type (i.e., LP105 polypeptide) refers to a specific type of molecule within the designated group of molecules as designated herein.

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The terms "LP polypeptide-encoding polynucleotides" or "LP polynucleotides" and "LP polypeptides" wherein the term "LP" is followed by an actual numerical designation as used herein encompass novel polynucleotides and polypeptides, respectively, which are further defined herein. The LP molecules described herein may be isolated from a variety of sources including, but not limited to human tissue types, or prepared by recombinant or synthetic methods.

One aspect of the present invention provides an isolated nucleic acid molecule comprising a polynucleotide which encodes an LP105, LP061, LP224, LP240, LP239(a), LP243(a), LP243(b), LP253, LP218, LP251(a), LP252, LP239(b), LP223(a), LP255(a), LP244, LP186, LP251(b), LP255(b), or LP223(b) polypeptide as defined herein. In a preferred embodiment of the present invention, the isolated

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nucleic acid comprises 1) a polynucleotide encoding an LP105, LP061, LP224, LP240, LP239(a), LP243(a), LP243(b), LP253, LP218, LP251(a), LP252, LP239(b), LP223(a), LP255(a), LP244, LP186, LP251(b), LP255(b), or LP223(b)

5 polypeptide having an amino acid sequence as shown in SEQ ID NO:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, or 38, respectively; 2) a polynucleotide complementary to such encoding nucleic acid sequences, and which remain stably bound to them under at least moderate, and optionally, high stringency conditions; or 3) any fragment and/or variant of 1) or 2).

The term "LP polypeptide" specifically encompasses truncated or secreted forms of an LP polypeptide, (e.g., soluble forms containing, for instance, an extracellular domain sequence), variant forms (e.g., alternatively spliced forms) and allelic variants of an LP polypeptide.

In one embodiment of the invention, the native sequence LP polypeptide is a full-length or mature LP polypeptide comprising amino acids as shown in SEQ ID NO:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, or 38. The predicted signal peptides are indicated in the sequence listing of the present application by negative integers below the amino acids disclosed for a particular polypeptide. Also, while the LP polypeptides disclosed herein are shown to begin with a methionine residue designated as amino acid position 1, it is conceivable and possible that another methionine residue located either upstream or downstream from amino acid position 1 may be employed as the starting amino acid residue.

A "portion" of an LP polypeptide sequence is at least about 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80,

85, 90, 95, or 100 contiguous amino acid residues in length.

"LP polypeptide variant" is intended to refer to an "active" LP polypeptide, wherein activity is as defined 5 herein, having at least about 90% amino acid sequence identity with an LP polypeptide having a deduced amino acid sequences as shown above. Such LP polypeptide variants include, for instance, LP polypeptides, wherein one or more amino acid residues are added, substituted or deleted, at 10 the N- or C-terminus or within the sequences shown. Ordinarily, an LP polypeptide variant will have at least about 90% amino acid sequence identity, preferably at least about 91% sequence identity, yet more preferably at least about 92% sequence identity, yet more preferably at least 15 about 93% sequence identity, yet more preferably at least about 94% sequence identity, yet more preferably at least about 95% sequence identity, yet more preferably at least about 96% sequence identity, yet more preferably at least about 97% sequence identity, yet more preferably at least about 98% sequence identity, yet more preferably at least 20 about 99% amino acid sequence identity with the amino acid sequence described, with or without the signal peptide.

"Percent (%) amino acid sequence identity" with respect to the LP amino acid sequences identified herein is defined as the percentage of amino acid residues in a candidate sequence that are identical with the amino acid residues in an LP polypeptide sequence, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity, and not considering any conservative substitutions as part of the sequence identity. Alignment for purposes of determining percent amino acid sequence identity can be achieved in various

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ways that are within the skill in the art, for instance, using publicly available computer software such as ALIGN, ALIGN-2, Megalign (DNASTAR) or BLAST (e.g., Blast, Blast-2, WU-Blast-2) software. Those skilled in the art can determine appropriate parameters for measuring alignment, including any algorithms needed to achieve maximal alignment over the full length of the sequences being compared. For example, the percent identity values used herein are generated using WU-BLAST-2 [Altschul, et al., Methods in Enzymology 266:460-80 (1996)]. Most of the WU-10 BLAST-2 search parameters are set to the default values. Those not set to default values, i.e., the adjustable parameters, are set with the following values: overlap span = 1; overlap fraction = 0.125; word threshold (T) = 11; and scoring matrix = BLOSUM 62. For purposes 15 herein, a percent amino acid sequence identity value is determined by dividing (a) the number of matching identical amino acid residues between the amino acid sequence of the LP polypeptide of interest and the comparison amino acid sequence of interest (i.e., the sequence against which the 20 LP polypeptide of interest is being compared) as determined by WU-BLAST-2, by (b) the total number of amino acid residues of the LP polypeptide of interest, respectively.

variant," or "LP variant nucleic acid sequence" are intended to refer to an nucleic acid molecule as defined below having at least about 75% nucleic acid sequence identity with the polynucleotide sequence as shown in SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, or 37. Ordinarily, an LP polynucleotide variant will have at least about 75% nucleic acid sequence identity, more preferably at least about 80% nucleic acid

sequence identity, yet more preferably at least about 81% nucleic acid sequence identity, yet more preferably at least about 82% nucleic acid sequence identity, yet more preferably at least about 83% nucleic acid sequence identity, yet more preferably at least about 84% nucleic acid sequence identity, yet more preferably at least about 85% nucleic acid sequence identity, yet more preferably at least about 86% nucleic acid sequence identity, yet more preferably at least about 87% nucleic acid sequence identity, yet more preferably at least about 88% nucleic 10 acid sequence identity, yet more preferably at least about 89% nucleic acid sequence identity, yet more preferably at least about 90% nucleic acid sequence identity, yet more preferably at least about 91% nucleic acid sequence identity, yet more preferably at least about 92% nucleic 15 acid sequence identity, yet more preferably at least about 93% nucleic acid sequence identity, yet more preferably at least about 94% nucleic acid sequence identity, yet more preferably at least about 95% nucleic acid sequence identity, yet more preferably at least about 96% nucleic 20 acid sequence identity, yet more preferably at least about 97% nucleic acid sequence identity, yet more preferably at least about 98% nucleic acid sequence identity, yet more preferably at least about 99% nucleic acid sequence identity with the nucleic acid sequences shown above. 25 Variants specifically exclude or do not encompass the native nucleotide sequence, as well as those prior art sequences that share 100% identity with the nucleotide sequences of the invention.

"Percent (%) nucleic acid sequence identity" with respect to the LP polynucleotide sequences identified herein is defined as the percentage of nucleotides in a

candidate sequence that are identical with the nucleotides in the LP polynucleotide sequence after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity. Alignment for purposes of determining percent nucleic acid sequence identity can be achieved in various ways that are within the skill in the art, for instance, using publicly available computer software such as ALIGN, Align-2, Megaligm (DNASTAR), or BLAST (e.g., Blast, Blast-2) Those skilled in the art can determine 10 appropriate parameters for measuring alignment, including any algorithms needed to achieve maximal alignment over the full length of the sequences being compared. For purposes herein, however, % nucleic acid identity values are generated using the WU-BLAST-2 (BlastN module) program 15 [Altschul, et al., Methods in Enzymology 266:460-80 (1996)]. Most of the WU-BLAST-2 search parameters are set to the default values. Those not set default values, i.e., the adjustable parameters, are set with the following values: overlap span = 1; overlap fraction = 0.125; word 20 threshold (T) = 11; and scoring matrix = BLOSUM62. For purposes herein, a percent nucleic acid sequence identity value is determined by dividing (a) the number of matching identical nucleotides between the nucleic acid sequence of the LP polypeptide-encoding nucleic acid molecule of 25 interest and the comparison nucleic acid molecule of interest (i.e., the sequence against which the LP polypeptide-encoding nucleic acid molecule of interest is being compared) as determined by WU-BLAST-2, by (b) the total number of nucleotides of the LP polypeptide-encoding nucleic acid molecule of interest.

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In other embodiments, the LP variant polypeptides are encoded by nucleic acid molecules which are capable of hybridizing, preferably under stringent hybridization and wash conditions, to nucleotide sequences encoding the fulllength LP polypeptide as shown in SEQ ID NO:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, or 38. This scope of variant polynucleotides specifically excludes those sequences that are known as of the filing and/or priority dates of the present application.

10 The term "mature protein" or "mature polypeptide" as used herein refers to the form(s) of the protein produced by expression in a mammalian cell. It is generally hypothesized that once export of a growing protein chain across the rough endoplasmic reticulum has been initiated, 15 proteins secreted by mammalian cells have a signal peptide (SP) sequence which is cleaved from the complete polypeptide to produce a "mature" form of the protein. Oftentimes, cleavage of a secreted protein is not uniform and may result in more than one species of mature protein. 20 The cleavage site of a secreted protein is determined by the primary amino acid sequence of the complete protein and generally cannot be predicted with complete accuracy. Methods for predicting whether a protein has an SP sequence, as well as the cleavage point for that sequence, 25 are available. A cleavage point may exist within the Nterminal domain between amino acid 10 and amino acid 35. More specifically the cleavage point is likely to exist after amino acid 15 but before amino acid 30, more likely after amino acid 27. As one of ordinary skill would . 30 appreciate, however, cleavage sites sometimes vary from organism to organism and cannot be predicted with absolute certainty. Optimally, cleavage sites for a secreted

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protein are determined experimentally by amino-terminal sequencing of the one or more species of mature proteins found within a purified preparation of the protein.

The term "positives," in the context of sequence comparison performed as described above, includes residues in the sequences compared that are not identical but have similar properties (e.g., as a result of conservative substitutions). The percent identity value of positives is determined by the fraction of residues scoring a positive value in the BLOSUM 62 matrix. This value is determined by dividing (a) the number of amino acid residues scoring a positive value in the BLOSUM62 matrix of WU-BLAST-2 between the LP polypeptide amino acid sequence of interest and the comparison amino acid sequence (i.e., the amino acid sequence against which the LP polypeptide sequence is being compared) as determined by WU-BLAST-2, by (b) the total number of amino acid residues of the LP polypeptide of interest.

"Isolated," when used to describe the various polypeptides disclosed herein, means a polypeptide that has 20 been identified and separated and/or recovered from a component of its natural environment. Preferably, the isolated polypeptide is free of association with all components with which it is naturally associated. Contaminant components of its natural environment are 25 materials that would typically interfere with diagnostic or therapeutic uses for the polypeptide, and may include enzymes, hormones, and other proteinaceous or nonproteinaceous solutes. In preferred embodiments, the polypeptide will be purified (1) to a degree sufficient to obtain at least 15 residues of N-terminal or internal amino acid sequence by use of a spinning cup sequenator, or (2)

to homogeneity by SDS-PAGE under non-reducing or reducing conditions using Coomassie blue or, preferably, silver Isolated polypeptide includes polypeptide in situ within recombinant cells, since at least one component of the LP polypeptide natural environment will not be present. Ordinarily, however, isolated polypeptide will be prepared by at least one purification step.

An "isolated LP polypeptide-encoding nucleic acid" or "isolated LP nucleic acid" is a nucleic acid molecule that is identified and separated from at least one contaminant nucleic acid molecule with which it is ordinarily associated in the natural source of the nucleic acid. an isolated nucleic acid molecule is other than in the form or setting in which it is found in nature. Isolated nucleic acid molecules therefore are distinguished from the 15 nucleic acid molecule as it exists in natural cells. However, an isolated LP polypeptide-encoding nucleic acid molecule includes LP polypeptide-encoding nucleic acid molecules contained in cells that ordinarily express LP polypeptide where, for example, the nucleic acid molecule 20 is in a chromosomal location different from that of natural cells.

Nucleic acid is "operably linked" when it is placed into a functional relationship with another nucleic acid 25 sequence. For example, DNA for a presequence or secretory leader is operably linked to DNA for a polypeptide if it is expressed as a preprotein that participates in the secretion of the polypeptide; a promoter or enhancer is operably linked to a coding sequence if it affects the transcription of the sequence; or a ribosome binding site 30 is operably linked to a coding sequence if it is positioned so as to facilitate translation. Generally, "operably

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linked" means that the DNA sequences being linked are contiguous, and, in the case of a secretory leader, contiguous and in reading phase. However, enhancers do not have to be contiguous. Linking is accomplished by ligation at convenient restriction sites. If such sites do not exist, the synthetic oligonucleotide adapters or linkers are used in accordance with conventional practice.

The term "amino acid" is used herein in its broadest sense, and includes naturally occurring amino acids as well as non-naturally occurring amino acids, including amino acid analogs and derivatives. The latter includes molecules containing an amino acid moiety. One skilled in the art will recognize, in view of this broad definition, that reference herein to an amino acid includes, for example, naturally occurring proteogenic L-amino acids; Damino acids; chemically modified amino acids such as amino acid analogs and derivatives; naturally-occurring nonproteogenic amino acids such as norleucine, beta-alanine, ornithine, etc.; and chemically synthesized compounds having properties known in the art to be characteristic of amino acids. As used herein, the term "proteogenic" indicates that the amino acid can be incorporated into a peptide, polypeptide, or protein in a cell through a metabolic pathway.

The incorporation of non-natural amino acids, including synthetic non-native amino acids, substituted amino acids, or one or more D-amino acids into the LP peptides, polypeptides, or proteins of the present invention ("D-LP polypeptides") is advantageous in a number of different ways. D-amino acid-containing peptides, polypeptides, or proteins exhibit increased stability in vitro or in vivo compared to L-amino acid-containing

counterparts. Thus, the construction of peptides, polypeptides, or proteins incorporating D-amino acids can be particularly useful when greater intracellular stability is desired or required. More specifically, D-peptides, polypeptides, or proteins are resistant to endogenous peptidases and proteases, thereby providing improved bioavailability of the molecule and prolonged lifetimes in vivo when such properties are desirable. When it is desirable to allow the peptide, polypeptide, or protein to remain active for only a short period of time, the use of L-amino acids therein will permit endogenous peptidases, proteases, etc., in a cell to digest the molecule in vivo, thereby limiting the cell's exposure to the molecule. Additionally, D-peptides, polypeptides, or proteins cannot be processed efficiently for major histocompatibility complex class II-restricted presentation to T helper cells, and are therefore less likely to induce humoral immune responses in the whole organism.

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In addition to using D-amino acids, those of ordinary 20 skill in the art are aware that modifications in the amino acid sequence of a peptide, polypeptide, or protein can result in equivalent, or possibly improved, second generation peptides, polypeptides, or proteins, that display equivalent or superior functional characteristics 25 when compared to the original amino acid sequences. Alterations in the LP peptides, polypeptides, or proteins of the present invention can include one or more amino acid insertions, deletions, substitutions, truncations, fusions, shuffling of subunit sequences, and the like, either from natural mutations or human manipulation, provided that the sequences produced by such modifications have substantially the same (or improved or reduced, as may be desirable)

activity(ies) as the naturally-occurring counterpart sequences disclosed herein.

One factor that can be considered in making such changes is the hydropathic index of amino acids. importance of the hydropathic amino acid index in conferring interactive biological function on a protein has been discussed by Kyte and Doolittle [J. Mol. Biol. 157:105-32 (1982)]. It is accepted that the relative hydropathic character of amino acids contributes to the secondary structure of the resultant protein. This, in 10 turn, affects the interaction of the protein with molecules such as enzymes, substrates, receptors, ligands, DNA, antibodies, antigens, etc. Based on its hydrophobicity and charge characteristics, each amino acid has been assigned a hydropathic index as follows: isoleucine (+4.5); valine 15 (+4.2); leucine (+3.8); phenylalanine (+2.8); cysteine/cystine (+2.5); methionine (+1.9); alanine (+1.8); glycine (-0.4); threonine (-0.7); serine (-0.8); tryptophan (-0.9); tyrosine (-1.3); proline (-1.6); histidine (-3.2); glutamate/glutamine/aspartate/asparagine (-3.5); lysine 20 (-3.9); and arginine (-4.5).

As is known in the art, certain amino acids in a peptide, polypeptide, or protein can be substituted for other amino acids having a similar hydropathic index or score and produce a resultant peptide, polypeptide, or protein having similar biological activity, i.e., which still retains biological functionality. In making such changes, it is preferable that amino acids having hydropathic indices within ±2 are substituted for one another. More preferred substitutions are those wherein the amino acids have hydropathic indices within ±1. Most

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preferred substitutions are those wherein the amino acids have hydropathic indices within ± 0.5 .

Like amino acids can also be substituted on the basis of hydrophilicity. U.S. Patent 4,554,101 discloses that the greatest local average hydrophilicity of a protein, as 5 governed by the hydrophilicity of its adjacent amino acids, correlates with a biological property of the protein. following hydrophilicity values have been assigned to amino acids: arginine/lysine (+3.0); aspartate/glutamate (+3.0±1); serine (+0.3); asparagine/glutamine (+0.2); glycine (0); 10 threonine (-0.4); proline (-0.5±1); alanine/histidine (-0.5); cysteine (-1.0); methionine (-1.3); valine (-1.5); leucine/isoleucine (-1.8); tyrosine (-2.3); phenylalanine (-2.5); and tryptophan (-3.4). Thus, one amino acid in a peptide, polypeptide, or protein can be substituted by 15 another amino acid having a similar hydrophilicity score and still produce a resultant peptide, polypeptide, or protein having similar biological activity, i.e., still retaining correct biological function. In making such changes, amino acids having hydropathic indices within ± 2 20 are preferably substituted for one another, those within ± 1 are more preferred, and those within ± 0.5 are most preferred.

As outlined above, amino acid substitutions in the LP

25 polypeptides of the present invention can be based on the
relative similarity of the amino acid side-chain
substituents, for example, their hydrophobicity, hydrophilicity, charge, size, etc. Exemplary substitutions that
take various of the foregoing characteristics into

30 consideration in order to produce conservative amino acid
changes resulting in silent changes within the present

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peptides, polypeptides, or proteins can be selected from other members of the class to which the naturally occurring amino acid belongs. Amino acids can be divided into the following four groups: (1) acidic amino acids; (2) basic amino acids; (3) neutral polar amino acids; and (4) neutral non-polar amino acids. Representative amino acids within these various groups include, but are not limited to: (1) acidic (negatively charged) amino acids such as aspartic acid and glutamic acid; (2) basic (positively charged) amino acids such as arginine, histidine, and lysine; (3) neutral polar amino acids such as glycine, serine, threonine, cysteine, cystine, tyrosine, asparagine, and glutamine; and (4) neutral non-polar amino acids such as alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan, and methionine.

It should be noted that changes which are not expected to be advantageous can also be useful if these result in the production of functional sequences. Since small peptides polypeptides, and some proteins can be easily produced by conventional solid phase synthetic techniques, the present invention includes peptides, polypeptides, or proteins such as those discussed herein, containing the amino acid modifications discussed above, alone or in various combinations. To the extent that such modifications can be made while substantially retaining the activity of the peptide, polypeptide, or protein, they are included within the scope of the present invention. The utility of such modified peptides, polypeptides, or proteins can be determined without undue experimentation by, for example, the methods described herein.

While biologically functional equivalents of the present LP polypeptides can have any number of conservative

or non-conservative amino acid changes that do not significantly affect their activity(ies), or that increase or decrease activity as desired, 40, 30, 20, 10, 5, or 3 changes, such as 1 to 30 changes or any range or value therein, may be preferred. In particular, ten or fewer amino acid changes may be preferred. More preferably, seven or fewer amino acid changes may be preferred; most preferably, five or fewer amino acid changes may be The encoding nucleotide sequences (gene, plasmid preferred. 10 DNA, cDNA, synthetic DNA, or mRNA, for example) will thus have corresponding base substitutions, permitting them to code on expression for the biologically functional equivalent forms of the LP polypeptides. In any case, the LP peptides, polypeptides, or proteins exhibit the same or 15 similar biological or immunological activity(ies) as that (those) of the LP polypeptides specifically disclosed herein, or increased or reduced activity, if desired. activity(ies) of the variant LP polypeptides can be determined by the methods described herein. Variant LP 20 polypeptides biologically functionally equivalent to those specifically disclosed herein have activity(ies) differing from those of the presently disclosed molecules by about ±50% or less, preferably by about ±40% or less, more preferably by about ±30% or less, more preferably by about 25 ±20% or less, and even more preferably by about ±10% or less, when assayed by the methods disclosed herein.

Amino acids in an LP polypeptide of the present invention that are essential for activity can be identified by methods known in the art, such as site-directed

30 mutagenesis or alanine-scanning mutagenesis [Cunningham and Wells, Science 244:1081-5 (1989)]. The latter procedure introduces single alanine mutations at every residue in the

molecule. The resulting mutant molecules are then tested, for biological activity. Sites that are critical for ligand-protein binding can also be identified by structural analysis such as crystallization, nuclear magnetic resonance, or photoaffinity labeling [Smith, et al., J. Mol. Biol. 224:899-904 (1992); de Vos, et al., Science 255:306-12 (1992)].

"Stringency" of hybridization reactions is readily determinable by one of ordinary skill in the art, and generally is an empirical calculation dependent upon probe length, washing temperature, and salt concentration. general, longer nucleic acid probes required higher temperatures for proper annealing, while shorter nucleic acid probes need lower temperatures. Hybridization generally depends on the ability of denatured DNA to reanneal when complementary strands are present in an environment below their melting temperature. The higher the degree of desired homology between the probe and hybridizable sequence, the higher the relative temperature that can be used. As a result, it follows that higher relative temperatures would tend to make the reactions more stringent, while lower temperatures less so. For additional details and explanation of stringency of hybridization reactions, see Ausubel, et al., Current Protocols in Molecular Biology, Wiley Interscience Publishers (1995).

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"Stringent conditions" or "high stringency conditions," as defined herein, may be identified by those that (1) employ low ionic strength and high temperature for washing, for example, 15 mM sodium chloride/1.5 mM sodium citrate/0.1% sodium dodecyl sulfate at 50 degrees C;

(2) employ during hybridization a denaturing agent, such as formamide, for example, 50% (v/v) formamide with 0.1%

bovine serum albumin/0.1% ficoll/0.1% polyvinylpyrrolidone/50 mM sodium phosphate buffer at pH 6.5 with 750 mM
sodium chloride/75 mM sodium citrate at 42 degrees C; or
(3) employ 50% formamide, 5X SSC (750 mM sodium chloride,
5 75 mM sodium citrate), 50 mM sodium phosphate (pH 6.8),
0.1% sodium pyrophosphate, 5X Denhardt's solution,
sonicated salmon sperm DNA (50 μg/mL), 0.1% SDS, and 10%
dextran sulfate at 42 degrees C with washes at 42 degrees C
in 0.2X SSC (30 mM sodium chloride/3 mM sodium citrate) and
10 50% formamide at 55 degrees C, followed by a highstringency wash consisting of 0.1X SSC containing EDTA at
55 degrees C.

"Moderately stringent conditions" may be identified as described by Sambrook, et al. [Molecular Cloning: A Laboratory Manual, New York: Cold Spring Harbor Press, 15 (1989)], and include the use of washing solution and hybridization conditions (e.g., temperature, ionic strength and %SDS) less stringent than those described above. example of moderately stringent conditions is overnight 20 incubation at 37 degrees C in a solution comprising: 20% formamide, 5X SSC (750 mM sodium chloride, 75 mM sodium citrate), 50 mM sodium phosphate at pH 7.6, 5X Denhardt's solution, 10% dextran sulfate, and 20 mg/mL denatured sheared salmon sperm DNA, followed by washing the filters in 1X SSC at about 37 to 50 degrees C. The skilled artisan 25 will recognize how to adjust the temperature, ionic strength, etc., as necessary to accommodate factors such as probe length and the like.

The term "epitope tagged" where used herein refers to

30 a chimeric polypeptide comprising an LP polypeptide, or

domain sequence thereof, fused to a "tag polypeptide." The

tag polypeptide has enough residues to provide an epitope

against which an antibody may be made, or which can be identified by some other agent, yet is short enough such that it does not interfere with the activity of the LP polypeptide. The tag polypeptide preferably is also fairly unique so that the antibody does not substantially cross-react with other epitopes. Suitable tag polypeptides generally have at least six amino acid residues and usually between about eight to about fifty amino acid residues (preferably, between about ten to about twenty residues).

As used herein, the term "immunoadhesin," sometimes referred to as an Fc fusion, designates antibody-like molecules that combine the binding specificity of a heterologous protein (an "adhesin") with the effector functions of immunoglobulin constant domains.

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Structurally, the immunoadhesins comprise a fusion of an amino acid sequence with the desired binding specificity which is other than the antigen recognition and binding site of an antibody (i.e., is "heterologous"), and an immunoglobulin constant domain sequence. The adhesin part of an immunoadhesin molecule typically is a contiguous amino acid sequence comprising at least the binding site of a receptor or a ligand. The immunoglobulin constant domain sequence in the immunoadhesin may be obtained from any immunoglobulin, such as IgG-1, IgG-2, IgG-3 or IgG-4 subtypes, IgA (including IgA-1 and IgA-2), IgE, IgD or IgM.

"Active" or "activity" for the purposes herein refers to form(s) of LP polypeptide which retain all or a portion of the biologic and/or immunologic activities of native or naturally-occurring LP polypeptide. Elaborating further, "biological" activity refers to a biological function (either inhibitory or stimulatory) caused by a native or naturally-occurring LP polypeptide other than the ability

to induce the production of an antibody against an antigenic epitope possessed by a native or naturally-occurring LP polypeptide. An "immunological" activity refers only to the ability to induce the production of an antibody against an antigenic epitope possessed by a native or naturally-occurring LP polypeptide.

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The term "antagonist" is used in the broadest sense and includes any molecule that partially or fully blocks, inhibits, or neutralizes a biological activity of a native LP polypeptide disclosed herein. In a similar manner, the term "agonist" is used in the broadest sense and includes any molecule that mimics a biological activity of a native LP polypeptide disclosed herein. Suitable agonist or antagonist molecules specifically include agonist or antagonist antibodies or antibody fragments, fragments or amino acid sequence variants of native LP polypeptides, peptides, ribozymes, anti-sense nucleic acids, small organic molecules, etc. Methods for identifying agonists or antagonists of an LP polypeptide may comprise contacting an LP polypeptide with a candidate agonist or antagonist molecule and measuring a detectable change in one or more biological activities normally associated with the LP polypeptide.

"Antibodies" (Abs) and "immunoglobulins" (Igs) are
glycoproteins having the same structural characteristics.
While antibodies exhibit binding specificity to a specific
antigen, immunoglobulins include both antibodies and other
antibody-like molecules that lack antigen specificity.
Polypeptides of the latter kind are, for example, produced
at low levels by the lymph system and at increased levels
by myelomas. The term "antibody" is used in the broadest
sense and specifically covers, without limitation, intact

monoclonal antibodies, polyclonal antibodies, multispecific antibodies (e.g., bispecific antibodies) formed from at least two intact antibodies, and antibody fragments so long as they exhibit the desired biological activity.

The terms "treating," "treatment," and "therapy" as used herein refer to curative therapy, prophylactic therapy, and preventive therapy. An example of "preventive therapy" is the prevention or lessened targeted pathological condition or disorder. Those in need of treatment include those already with the disorder as well as those prone to have the disorder or those in whom the disorder is to be prevented.

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"Chronic" administration refers to administration of the agent(s) in a continuous mode as opposed to an acute mode, so as to maintain the initial therapeutic effect (activity) for an extended period of time. "Intermittent" administration is treatment that is not consecutively done without interruption but, rather, is cyclic in nature.

Administration "in combination with" one or more further therapeutic agents includes simultaneous (concurrent) and consecutive administration in any order.

A "therapeutically-effective amount" is the minimal amount of active agent (e.g., an LP polypeptide, antagonist or agonist thereof) which is necessary to impart therapeutic benefit to a mammal. For example, a "therapeutically-effective amount" to a mammal suffering or prone to suffering or to prevent it from suffering is such an amount which induces, ameliorates, or otherwise causes an improvement in the pathological symptoms, disease progression, physiological conditions associated with or resistance to succumbing to the aforedescribed disorder.

"Carriers" as used herein include pharmaceuticallyacceptable carriers, excipients, or stabilizers which are nontoxic to the cell or mammal being exposed thereto at the dosages and concentrations employed. Often the 5 physiologically-acceptable carrier is an aqueous pH buffered solution. Examples of physiologically acceptable carriers include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid; low molecule weight (less than about 10 residues) 10 polypeptides; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, arginine or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, 15 mannose, or dextrins; chelating agents such as EDTA; sugar alcohols such as mannitol or sorbitol; salt-forming counterions such as sodium; and/or nonionic surfactants such as TWEEN®, polyethylene glycol (PEG), and PLURONIC®.

"Antibody fragments" comprise a portion of an intact
20 antibody, preferably the antigen binding or variable region
of the intact antibody. Examples of antibody fragments
include Fab, Fab', F(ab')₂ and Fv fragments; diabodies;
linear antibodies [Zapata, et al., Protein Engin. 8
(10):1057-62 (1995)]; single-chain antibody molecules; and
25 multispecific antibodies formed from antibody fragments.

"Fv" is the minimum antibody fragment which contains a complete antigen-recognition and binding site. This region consists of a dimer of one heavy- and one light-chain variable domain in tight, non-covalent association. It is in this configuration that the three CDRs of each variable domain interact to define an antigen-binding site on the surface of the V_HV_L dimer. Collectively, the six CDRs

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confer antigen-binding specificity to the antibody. However, even a single variable domain (or half of an Fv comprising only three CDR specific for an antigen) has the ability to recognize and bind antigen, although at a lower affinity than the entire binding site.

"Single-chain Fv" or "sFv" antibody fragments comprise the V_H and V_L domains of antibody, wherein these domains are present in a single polypeptide chain. Preferably, the Fv polypeptide further comprises a polypeptide linker between the V_H and V_L domain, which enables the sFv to form the desired structure for antigen binding. For a review of sFv, see Pluckthun, The Pharmacology of Monoclonal Antibodies, vol. 113, Rosenburg and Moore, eds., Springer-Verlag, New York, pp. 269-315 (1994).

15 The term "diabodies" refers to small antibody fragments with two antigen-binding sites, which fragments comprise a heavy-chain variable domain (V_H) connected to a light-chain variable domain (V_L) in the same polypeptide chain (V_H-V_L). By using a linker that is too short to allow pairing between the two domains on the same chain, the domains are forced to pair with the complementary domains of another chain and create two antigen-binding sites.

Diabodies are described more fully in, for example, EP 404 097; WO 93/11161; and Hollinger, et al., Proc. Natl.

25 Acad. Sci. USA 90:6444-8 (1993).

An "isolated" antibody is one which has been identified and separated and/or recovered from a component of its natural environment. Contaminant components of its natural environment are materials which would interfere with diagnostic or therapeutic uses for the antibody, and may include enzymes, hormones, and other proteinaceous or non-proteinaceous solutes. In preferred embodiments, the

antibody will be purified (1) to greater than 95% by weight of antibody as determined by the Lowry method, and most preferably more than 99% by weight, (2) to a degree sufficient to obtain at least 15 residues of N-terminal or internal amino acid sequence by use of a spinning cup sequenator, or (3) to homogeneity by SDS-PAGE under reducing or nonreducing conditions using Coomassie blue, or preferably, silver stain. Isolated antibody includes the antibody in situ within recombinant cells since at least one component of the antibody's natural environment will not be present. Ordinarily, however, isolated antibody will be prepared by at least one purification step.

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An "LP polypeptide antibody" or "LP antibody" refers to an antibody as defined herein that recognizes and binds at least one epitope of an LP polypeptide of the present invention. The term "LP polypeptide antibody" or "LP antibody" wherein the term "LP" is followed by a numerical designation refers to an antibody that recognizes and binds to at least one epitope of that particular LP polypeptide as disclosed herein.

A "liposome" is a small vesicle composed of various types of lipids, phospholipids and/or surfactant which is useful for delivery of a drug (such as an LP polypeptide or antibody thereto) to a mammal. The components of the liposome are commonly arranged in a bilayer formation, similar to the lipid arrangement of biological membranes.

A "small molecule" is defined herein to have a molecular weight below about 500 daltons.

The term "modulate" means to affect (e.g., either upregulate, downregulate or otherwise control) the level of a signaling pathway. Cellular processes under the control of signal transduction include, but are not limited to,

transcription of specific genes, normal cellular functions, such as metabolism, proliferation, differentiation, adhesion, apoptosis and survival, as well as abnormal processes, such as transformation, blocking of differentiation and metastasis.

An LP polynucleotide can be composed of any polyribonucleotide or polydeoxyribonucleotide, which may be unmodified RNA or DNA or modified RNA or DNA. For example, the LP polynucleotides can be composed of single- and double-stranded DNA, DNA that is a mixture of single- and 10 double-stranded regions, single- and double-stranded RNA, and RNA that is mixture of single- and double-stranded regions, hybrid molecules comprising DNA and RNA that may be single-stranded or, more typically, double-stranded or a 15 mixture of single- and double-stranded regions. addition, LP polynucleotides can be composed of triplestranded regions comprising RNA or DNA or both RNA and DNA. LP polynucleotides may also contain one or more modified bases or DNA or RNA backbones modified for stability or for other reasons. "Modified" bases include, for example, 20 tritylated bases and unusual bases such as inosine. A variety of modifications can be made to DNA and RNA; thus, "polynucleotide" embraces chemically, enzymatically, or metabolically modified forms.

LP polypeptides can be composed of amino acids joined to each other by peptide bonds or modified peptide bonds, i.e., peptide isosteres, and may contain amino acids other than the gene-encoded amino acids. The LP polypeptides may be modified by either natural processes, such as post-translational processing, or by chemical modification techniques which are well known in the art. Such modifications are well described in basic texts and in more

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detailed monographs, as well as in a voluminous research literature. Modifications can occur anywhere in the LP polypeptides, including the peptide backbone, the amino acid side-chains and the amino or carboxyl termini. It will be appreciated that the same type of modification may be present in the same or varying degrees at several sites in a given LP polypeptide. Also, a given LP polypeptide may contain many types of modifications. LP polypeptides may be branched, for example, as a result of ubiquitination, and 10 they may be cyclic, with or without branching. Cyclic, branched, and branched cyclic LP polypeptides may result from post-translation natural processes or may be made by synthetic methods. Modifications include acetylation, acylation, ADP-ribosylation, amidation, covalent attachment 15 of flavin, covalent attachment of a heme moiety, covalent attachment of a nucleotide or nucleotide derivative, covalent attachment of a lipid or lipid derivative, covalent attachment of phosphotidylinositol, cross-linking, cyclization, disulfide bond formation, demethylation, 20 formation of covalent cross-links, formation of cysteine, formation of pyroglutamate, formylation, gammacarboxylation, glycosylation, GPI anchor formation, hydroxylation, iodination, methylation, myristoylation, oxidation, pegylation, proteolytic processing, 25 phosphorylation, prenylation, racemization, selenoylation, sulfation, transfer-RNA mediated addition of amino acids to proteins such as arginylation, and ubiquitination. See, for

proteins such as arginylation, and ubiquitination. See, for instance, Creighton, Proteins - Structure and Molecular Properties, 2nd Ed., W. H. Freeman and Company, New York (1993); Johnson, Post-translational Covalent Modification of Proteins, Academic Press, New York, pp. 1-12 (1983);

Seifter, et al., Meth. Enzymol. 182:626-46 (1990); Rattan, et al., Ann. NY Acad. Sci. 663:48-62 (1992).

Variations in the full-length sequence LP polypeptide or in various domains of the LP polypeptide described herein can be made, for example, using any of the techniques and guidelines for conservative and nonconservative mutations set forth, for instance, in U.S. Patent 5,364,934. Variations may be a substitution, deletion or insertion of one or more codons encoding LP polypeptide that results in a change in the amino acid 10 sequence of the LP polypeptide as compared with the native sequence LP polypeptide or an LP polypeptide as disclosed Optionally the variation is by substitution of at least one amino acid with any other amino acid in one or more of the domains of the LP polypeptide. Guidance in 15 determining which amino acid residue may be inserted, substituted or deleted without adversely affecting the desired activity may be found by comparing the sequence of the LP polypeptide with that of homologous known protein molecules and minimizing the number of amino acid sequence 20 changes made in regions of high homology. Amino acid substitutions can be the result of replacing one amino acid with another amino acid having similar structural and/or chemical properties, such as the replacement of a leucine with a serine, i.e., conservative amino acid replacements. 25 Insertions or deletions may optionally be in the range of one to five amino acids. The variation allowed may be determined by systematically making insertions, deletions or substitutions of amino acids in the sequence and testing the resulting variants for activity (such as in any of the 30 in vitro assays described herein) for activity exhibited by the full-length or mature polypeptide sequence.

LP polypeptide fragments are also provided herein. Such fragments may be truncated at the N-terminus or C-terminus, or may lack internal residues, for example, when compared with a full length or native protein.

Certain fragments contemplated by the present invention may lack amino acid residues that are not essential for a desired biological activity of the LP polypeptide.

LP polypeptide fragments may be prepared by any of a number of conventional techniques. Desired peptide 10 fragments may be chemically synthesized. An alternative approach involves generating LP fragments by enzymatic digestion, e.g., by treating the protein with an enzyme known to cleave proteins at sites defined by particular amino acid residues, or by digesting the DNA with suitable 15 restriction enzymes and isolating the desired fragment. Yet another suitable technique involves isolating and amplifying a DNA fragment encoding a desired polypeptide fragment by polymerase chain reaction (PCR). Oligonucleotides that define the desired termini of the DNA fragment 20 are employed at the 5' and 3' primers in the PCR. Preferably, LP polypeptide fragments share at least one biological and/or immunological activity with at least one of the LP polypeptides as shown in SEQ ID NO:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, or 25 38.

Covalent modifications of LP polypeptides are included within the scope of this invention. One type of covalent modification includes reacting targeted amino acid residues of an LP polypeptide with an organic derivatizing agent that is capable of reacting with selected side chains or the N- or C-terminal residues of an LP polypeptide.

Derivatization with bifunctional agents is useful, for

instance, for crosslinking LP polypeptide to a waterinsoluble support matrix or surface for use in the method
for purifying anti-LP polypeptide antibodies, and viceversa. Commonly used crosslinking agents include, e.g.,

1,1-bis(diazoacetyl)-2-phenylethane, glutaraldehyde,
N-hydroxysuccinimide esters, for example, esters with
4-azidosalicylic acid, homobifunctional imidoesters,
including disuccinimidyl esters such as 3,3'-dithiobis(succinimidylproprionate), bifunctional maleimides such as
bis-N-maleimido-1,8-octane and agents such as methyl-3-[(p-azidophenyl)dithiolproprioimidate.

Other modifications include deamidation of glutaminyl and asparaginyl residues to the corresponding glutamyl and aspartyl residues, respectively, hydroxylation of proline and lysine, phosphorylation of hydroxyl groups of seryl or threonyl residues, methylation of the alpha-amino groups of lysine, arginine, and histidine side chains [Creighton, Proteins: Structure and Molecular Properties, W.H. Freeman & Co., San Francisco, pp. 79-86 (1983)], acetylation of the N-terminal amine, and amidation of any C-terminal carboxyl group.

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Another type of covalent modification of the LP polypeptides included within the scope of this invention comprises altering the native glycosylation pattern of the polypeptide. "Altering the native glycosylation pattern" is intended for purposes herein to mean deleting one or more carbohydrate moieties found in native sequence LP polypeptide and/or adding one or more glycosylation sites that are not present in the native sequences of LP polypeptides. Additionally, the phrase includes qualitative changes in the glycosylation of the native

proteins, involving a change in the nature and proportions of the various carbohydrate moieties present.

Addition of glycosylation sites to LP polypeptides may be accomplished by altering the amino acid sequence

5 thereof. The alteration may be made, for example, by the addition of, or substitution by, one or more serine or threonine residues to the native sequences of LP polypeptides (for O-linked glycosylation sites). The LP amino acid sequences may optionally be altered through

10 changes at the DNA level, particularly by mutating the DNA encoding the LP polypeptides at preselected bases such that codons are generated that will translate into the desired amino acids.

Another means of increasing the number of carbohydrate moieties on the LP polypeptides is by chemical or enzymatic coupling of glycosides to the polypeptide. Such methods are described in the art, e.g., in WO 87/05330, and in Aplin and Wriston, CRC Crit. Rev. Biochem., pp. 259-306 (1981).

20 Removal of carbohydrate moieties present on the LP polypeptide may be accomplished chemically or enzymatically or by mutational substitution of codons encoding for amino acid residues that serve as targets for glycosylation. Chemical deglycosylation techniques are known in the art 25 and described, for instance, by Sojar, et al., Arch. Biochem. Biophys. 259:52-7 (1987), and by Edge, et al., Anal. Biochem. 118:131-7 (1981). Enzymatic cleavage of carbohydrate moieties on polypeptides can be achieved by the use of a variety of endo- and exo-glycosidases as described by Thotakura, et al., Meth. Enzymol. 138:350-9 (1987).

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Another type of covalent modification of LP comprises linking any one of the LP polypeptides to one of a variety of non-proteinaceous polymers (e.g., polyethylene glycol, polypropylene glycol, or polyoxyalkylenes) in the manner set forth in U.S. Patent Nos. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192, or 4,179,337.

LP polypeptides of the present invention may also be modified in a way to form chimeric molecules comprising an LP polypeptide fused to another heterologous polypeptide or amino acid sequence. In one embodiment, such a chimeric molecule comprises a fusion of an LP polypeptide with a tag polypeptide which provides an epitope to which an anti-tag antibody can selectively bind. The epitope tag is generally placed at the amino- or carboxyl- terminus of LP105, LP061, LP224, LP240, LP239(a), LP243(a), LP243(b), LP253, LP218, LP251(a), LP252, LP239(b), LP223(a), LP255(a), LP244, LP186, LP251(b), LP255(b), or LP223(b) polypeptide. The presence of such epitope-tagged forms of an LP polypeptide can be detected using an antibody against the tag polypeptide. Also, provision of the epitope tag enables an LP polypeptide to be readily purified by affinity purification using an anti-tag antibody or another type of affinity matrix that binds to the epitope tag.

In an alternative embodiment, the chimeric molecule

25 may comprise a fusion of an LP polypeptide with an

immunoglobulin or a particular region of an immunoglobulin.

For a bivalent form of the chimeric molecule, such a fusion

could be to the Fc region of an IgG molecule. The Ig

fusions preferably include the substitution of a soluble

30 transmembrane domain deleted or inactivated form of an LP

polypeptide in place of at least one variable region within

an Ig molecule. In a particularly preferred embodiment,

the immunoglobulin fusion includes the hinge, CH2 and CH3 or the hinge, CH1, CH2 and CH3 regions of an IgG1 molecule. For the production of immunoglobulin fusions, see also U.S. Patent 5,428,130.

In yet a further embodiment, the LP polypeptides of 5 the present invention may also be modified in a way to form a chimeric molecule comprising an LP polypeptide fused to a leucine zipper. Various leucine zipper polypeptides have been described in the art. See, e.g., Landschulz, et al., Science 240(4860):1759-64 (1988); WO 94/10308; Hoppe, et 10 al., FEBS Letters 344(2-3):191-5 (1994); Abel, et al., Nature 341(6237):24-5 (1989). It is believed that use of a leucine zipper fused to an LP polypeptide may be desirable to assist in dimerizing or trimerizing soluble LP 15 polypeptide in solution. Those skilled in the art will appreciate that the zipper may be fused at either the N- or C-terminal end of an LP polypeptide.

The description below relates primarily to production of LP polypeptides by culturing cells transformed or transfected with a vector containing an LP polypeptide-20 encoding nucleic acid. It is, of course, contemplated that alternative methods, which are well known in the art, may be employed to prepare LP polypeptides. For instance, the LP polypeptide sequence, or portions thereof, may be 25 produced by direct peptide synthesis using solid-phase techniques [see, e.g., Stewart, et al., Solid-Phase Peptide Synthesis, W.H. Freeman & Co., San Francisco, CA (1969); Merrifield, J. Am. Chem. Soc. 85:2149-2154 (1963)]. In vitro protein synthesis may be performed using manual techniques or by automation. Automated synthesis may be 30 accomplished, for instance, using an Applied Biosystems Peptide Synthesizer (Foster City, CA) using manufacturer's

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instructions. Various portions of an LP polypeptide may be chemically synthesized separately and combined using chemical or enzymatic methods to produce a full-length LP polypeptide.

DNA encoding an LP polypeptide may be obtained from a 5 cDNA library prepared from tissue believed to possess the LP polypeptide-encoding mRNA and to express it at a detectable level. Libraries can be screened with probes (such as antibodies to an LP polypeptide or 10 oligonucleotides of at least about 20 to 80 bases) designed to identify the gene of interest or the protein encoded by Screening the cDNA or genomic library with the selected probe may be conducted using standard procedures, such as described in Sambrook, et al., Molecular Cloning: A 15 Laboratory Manual, Cold Spring Harbor Laboratory Press, NY (1989). An alternative means to isolate the gene encoding an LP polypeptide is to use PCR methodology [Sambrook, et al., supra; Dieffenbach, et al., PCR Primer: A Laboratory Manual, Cold Spring Harbor Laboratory Press, NY (1995)].

Nucleic acids having protein coding sequence may be obtained by screening selected cDNA or genomic libraries using the deduced amino acid sequence disclosed herein for the first time and, if necessary, using conventional primer extension procedures as described in Sambrook, et al., supra, to detect precursors and processing intermediates of mRNA that may not have been reverse-transcribed into cDNA.

Host cells are transfected or transformed with expression or cloning vectors described herein for LP polypeptide production and cultured in conventional nutrient media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences. The culture conditions,

such as media, temperature, pH and the like, can be selected by the skilled artisan without undue experimentation. In general, principles, protocols, and practical techniques for maximizing the productivity of cell cultures can be found in Mammalian Cell Biotechnology: 5 A Practical Approach, Butler, ed. (IRL Press, 1991) and Sambrook, et al., supra. Methods of transfection are known to the ordinarily skilled artisan, for example, calcium phosphate and electroporation. General aspects of mammalian cell host system transformations have been 10 described in U.S. Patent 4,399,216. Transformations into yeast are typically carried out according to the method of van Solingen, et al., J Bact. 130(2):946-7 (1977) and Hsiao, et al., Proc. Natl. Acad. Sci. USA 76(8):3829-33 (1979). However, other methods for introducing DNA into 15 cells, such as by nuclear microinjection, electroporation, bacterial protoplast fusion with intact cells, or polycations, e.g., polybrene or polyornithine, may also be used. For various techniques for transforming mammalian cells, see Keown, et al., Methods in Enzymology 185:527-37 20 (1990) and Mansour, et al., Nature 336(6197):348-52 (1988). Suitable host cells for cloning or expressing the nucleic acid (e.g., DNA) in the vectors herein include prokaryote, yeast, or higher eukaryote cells. Suitable 25

nucleic acid (e.g., DNA) in the vectors herein include prokaryote, yeast, or higher eukaryote cells. Suitable

25 prokaryotes include but are not limited to eubacteria, such as Gram-negative or Gram-positive organisms, for example, Enterobacteriacea such as E. coli. Various E. coli strains are publicly available, such as E. coli K12 strain MM294 (ATCC 31,446); E. coli strain X1776 (ATCC 31,537); E. coli strain W3110 (ATCC 27,325) and K5 772 (ATCC 53,635). Other suitable prokaryotic host cells include Enterobacteriaceae such as Escherichia, e.g., E. coli,

Enterobacter, Erwinia, Klebsiella, Proteus, Salmonella, e.g., Salmonella typhimurium, Serratia, e.g., Serratia marcescans, and Shigella, as well as Bacilli such as B. subtilis and B. licheniformis (e.g., B. licheniformis 41P disclosed in DD 266,710, published 12 April 1989), 5 Pseudomonas such as P. aeruginosa, and Streptomyces. examples are illustrative rather than limiting. Strain W3110 is one particularly preferred host or parent host because it is a common host strain for recombinant DNA product fermentations. Preferably, the host cell secretes 10 minimal amounts of proteolytic enzymes. For example, strain W3 110 may be modified to effect a genetic mutation in a gene encoding proteins endogenous to the host, with examples of such hosts including E. coli W3110 strain 1A2, which has the complete genotype tonAD; E. coli W3110 strain 15 9E4, which has the complete genotype tonAD ptr3; E. coli W3110 strain 27C7 (ATCC 55,244), which has the complete genotype tonAD ptr3 phoADE15 D(argF-lac)169 ompTD degP41kan ; E. coli W3110 strain 37D6, which has the complete genotype tonAD ptr3 phoADE15 D(argF-lac)169 ompTD 20 degP41kanR rbs7D ilvG; E. coli W3110 strain 40B4, which is strain 37D6 with a non-kanamycin resistant degP deletion mutation; and an E. coli strain having mutant periplasmic protease as disclosed in U.S. Patent 4,946,783 issued 7 August 1990. Alternatively, in vivo methods of cloning, 25 e.g., PCR or other nucleic acid polymerase reactions, are suitable.

In addition to prokaryotes, eukaryotic microbes such as filamentous fungi or yeast are suitable cloning or expression hosts for LP vectors. Saccharomyces cerevisiae is a commonly used lower eukaryotic host microorganism.

Others include Schizosaccharomyces pombe [Beach and Nurse,

Nature 290:140-3 (1981); EP 139,383 published 2 May 1995]; Muyveromyces hosts [U.S. Patent 4,943,529 Fleer, et al., Bio/Technology 9(10):968-75 (1991)] such as, e.g., K lactis (MW98-8C, CBS683, CBS4574) [de Louvencourt, et al., J. Bacteriol. 154(2):737-42 (1983)]; K. fiagilis (ATCC 5 12,424), K. bulgaricus (ATCC 16,045), K wickeramii (ATCC 24,178), K waltii (ATCC 56,500), K. drosophilarum (ATCC 36.906) [Van den Berg, et al., Bio/Technology 8(2):135-9 (1990)]; K. thermotolerans, and K. marxianus; yarrowia 10 (EP 402,226); Pichia pastoris (EP 183,070) [Sreekrishna, et al., J. Basic Microbiol. 28(4):265-78 (1988)]; Candida; Trichoderma reesia (EP 244,234); Neurospora crassa [Case, et al., Proc. Natl. Acad Sci. USA 76(10):5259-63 (1979)]; Schwanniomyces such as Schwanniomyces occidentulis 15 (EP 394,538 published 31 October 1990); and filamentous fungi such as, e.g., Neurospora, Penicillium, Tolypocladium (WO 91/00357 published 10 January 1991), and Aspergillus hosts such as A. nidulans [Ballance, et al., Biochem. Biophys. Res. Comm. 112(1):284-9 (1983); Tilburn, et al., 20 Gene 26(2-3):205-21 (1983); Yelton, et al., Proc. Natl. Acad. Sci. USA 81(5):1470-4 (1984)] and A. niger [Kelly and Hynes, EMBO J. 4(2):475-9 (1985)]. Methylotropic yeasts are selected from the genera consisting of Hansenula, Candida, Kloeckera, Pichia, Saccharomyces, Torulopsis, and 25 Rhodotoruia. A list of specific species that are exemplary of this class of yeast may be found in Antony, The

Suitable host cells for the expression of glycosylated LP polypeptides are derived from multicellular organisms. Examples of invertebrate cells include insect cells such as Drosophila S2 and Spodoptera Sp, Spodoptera high5 as well as plant cells. Examples of useful mammalian host cell

Biochemistry of Methylotrophs 269 (1982).

lines include Chinese hamster ovary (CHO) and COS cells.

More specific examples include monkey kidney CV1 line
transformed by SV40 (COS-7, ATCC CRL 1651); human embryonic
kidney line [293 or 293 cells subcloned for growth in

5 suspension culture, Graham, et al., J. Gen Virol.,
36(1):59-74 (1977)]; Chinese hamster ovary cells/-DHFR
[CHO, Urlaub and Chasin, Proc. Natl. Acad. Sci. USA,
77(7):4216-20 (1980)]; mouse sertoli cells [TM4, Mather,
Biol. Reprod. 23(1):243-52 (1980)]; human lung cells (W138,
10 ATCC CCL 75); human liver cells (Hep G2, HB 8065); and
mouse mammary tumor (MMT 060562, ATCC CCL51). The
selection of the appropriate host cell is deemed to be
within the skill in the art.

LP polypeptides may be produced recombinantly not only 15 directly, but also as a fusion polypeptide with a heterologous polypeptide, which may be a signal sequence or other polypeptide having a specific cleavage site at the N-terminus of the mature protein or polypeptide. general, the signal sequence may be a component of the vector, or it may be a part of the LP polypeptide-encoding 20 DNA that is inserted into the vector. The signal sequence may be a prokaryotic signal sequence selected, for example, from the group of the alkaline phosphatase, penicillinase, lpp, or heat-stable enterotoxin II leaders. For yeast secretion the signal sequence may be, e.g., the yeast 25 invertase leader, alpha factor leader (including Saccharomyces and Kluyveromyces cc-factor leaders, the latter described in U.S. Patent 5,010,182), or acid phosphatase leader, the C. albicans glucoamylase leader (EP 362,179), or the signal described in WO 90/13646. 30 mammalian cell expression, mammalian signal sequences may be used to direct secretion of the protein, such as signal

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sequences from secreted polypeptides of the same or related species as well as viral secretory leaders.

Both expression and cloning vectors contain a nucleic acid sequence that enables the vector to replicate in one or more selected host cells. Such sequences are well known for a variety of bacteria, yeast, and viruses. The origin of replication from the plasmid pBR322 is suitable for most Gram-negative bacteria, the 2μ plasmid origin is suitable for yeast, and various viral origins (SV40, polyoma, adenovirus, VSV or BPV) are useful for cloning vectors in mammalian cells.

Expression and cloning vectors will typically contain a selection gene, also termed a selectable marker. Typical selection genes encode proteins that (a) confer resistance to antibiotics or other toxins, e.g., ampicillin, neomycin, methotrexate, or tetracycline, (b) complement auxotrophic deficiencies, or (c) supply critical nutrients not available from complex media, e.g., the gene encoding D-alanine racemase for Bacilli.

20 An example of suitable selectable markers for mammalian cells are those that enable the identification of cells competent to take up the LP polypeptide-encoding nucleic acid, such as DHFR or thymidine kinase. appropriate host cell when wild-type DHFR is employed is the CHO cell line deficient in DHFR activity, prepared and 25 propagated as described Urlaub and Chasin, Proc. Natl. Acad. Sci. USA, 77(7):4216-20 (1980). A suitable selection gene for use in yeast is the trpl gene present in the yeast plasmid YRp7 [Stinchcomb, et al., Nature 282(5734):39-43 (1979); Kingsman, et al., Gene 7(2):141-52 (1979); 30 Tschumper, et al., Gene 10(2):157-66 (1980)]. The trpl gene provides a selection marker for a mutant strain of

yeast lacking the ability to grow in tryptophan, for example, ATCC No. 44076 or PEPC1 [Jones, Genetics 85:23-33 (1977)].

Expression and cloning vectors usually contain a 5 promoter operably linked to the LP polypeptide-encoding nucleic acid sequence to direct mRNA synthesis. Promoters recognized by a variety of potential host cells are well Promoters suitable for use with prokaryotic hosts known. include the P-lactamase and lactose promoter systems [Chang, et al., Nature 275(5681):617-24 (1978); Goeddel, et 10 al., Nature 281(5732):544-8 (1979)], alkaline phosphatase, a tryptophan (up) promoter system [Goeddel, Nucleic Acids Res. 8(18):4057-74 (1980); EP 36,776 published 30 September 1981], and hybrid promoters such as the tat promoter [deBoer, et al., Proc. Natl. Acad. Sci. USA 80(1):21-5 15 (1983)]. Promoters for use in bacterial systems also will contain a Shine-Dalgarno (S.D.) sequence operably linked to the DNA encoding LP polypeptide.

Examples of suitable promoting sequences for use with

yeast hosts include the promoters for 3-phosphoglycerate
kinase [Hitzeman, et al., J. Biol. Chem. 255(24):12073-80

(1980)] or other glycolytic enzymes [Hess, et al., J. Adv.
Enzyme Reg. 7:149 (1968); Holland, Biochemistry

17(23):4900-7 (1978)], such as enolase, glyceraldehyde-3
phosphate dehydrogenase, hexokinase, pyruvate
decarboxylase, phosphofructokinase, glucose-6-phosphate
isomerase, 3-phosphoglycerate mutase, pyruvate kinase,
triosephosphate isomerase, phosphoglucose isomerase, and
glucokinase.

Other yeast promoters, which are inducible promoters having the additional advantage of transcription controlled by growth conditions, are the promoter regions for alcohol

dehydrogenase 2, isocytochrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose 5 utilization. Suitable vectors and promoters for use in yeast expression are further described in EP 73,657. transcription from vectors in mammalian host cells is controlled, for example, by promoters obtained from the genomes of viruses such as polyoma virus, fowlpox virus, adenovirus (such as Adenovirus 2), bovine papilloma virus, 10 avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis-B virus and Simian Virus 40 (SV40), from heterologous mammalian promoters, e.g., the actin promoter or an immunoglobulin promoter, and from heat-shock promoters, provided such promoters are compatible with the 15 host cell systems.

Transcription of a polynucleotide encoding an LP polypeptide by higher eukaryotes may be increased by inserting an enhancer sequence into the vector. Enhancers are cis-acting elements of DNA, usually about from 10 to 20 300 bp, that act on a promoter to increase its transcription. Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin, alphaketoprotein, and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples 25 include the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. enhancer may be spliced into the vector at a position 5' or 3' to the LP polypeptide coding sequence but is preferably located at a site 5' from the promoter.

Expression vectors used in eukaryotic host cells (yeast, fungi, insect, plant, animal, human, or nucleated cells from other multicellular organisms) will also contain sequences necessary for the termination of transcription and for stabilizing the mRNA. Such sequences are commonly available from the 5' and occasionally 3' untranslated regions of eukaryotic or viral DNAs or cDNAs. These regions contain nucleotide segments transcribed as polyadenylated fragments in the untranslated portion of the mRNA encoding LP.

Gene amplification and/or expression may be measured in a sample directly, for example, by conventional Southern blotting, Northern blotting to quantitate the transcription of mRNA [Thomas, Proc. Natl. Acad. Sci. USA 77(9):5201-5 15 (1980)], dot blotting (DNA analysis), or in situ hybridization, using an appropriately labeled probe, based on the sequences provided herein. Alternatively, antibodies may be employed that can recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. The antibodies in 20 turn may be labeled and the assay may be carried out where the duplex is bound to a surface, so that upon the formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Gene expression, alternatively, may be measured by immunological methods, such as immunohistochemical staining of cells or tissue sections and assay of cell culture or body fluids, to quantitate directly the expression of gene product. Antibodies useful for immunohistochemical staining and/or assay of sample fluids may be either monoclonal or polyclonal and may be prepared in any mammal. Conveniently, the antibodies may be prepared against a

native sequence provided herein or against exogenous sequence fused to an LP polypeptide-encoding DNA and encoding a specific antibody epitope.

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Various forms of an LP polypeptide may be recovered from culture medium or from host cell lysates. If membrane-bound, it can be released from the membrane using a suitable detergent solution (e.g., Triton X-100TM) or by enzymatic cleavage. Cells employed in expression of an LP polypeptide can be disrupted by various physical or chemical means, such as freeze-thaw cycling, sonication, mechanical disruption, or cell lysing agents.

It may be desireable to purify LP polypeptides from recombinant cell proteins or polypeptides. The following procedures are exemplary of suitable purification procedures: by fractionation on an ion-exchange column; ethanol precipitation; reversed-phase HPLC; chromatography on silica or on a cation-exchange resin such as DEAE; chromatofocusing; SDS-PAGE; ammonium sulfate precipitation; gel filtration using, for example, Sephadex® G-75; protein A Sepharose® columns to remove contaminants such as IgG; and metal chelating columns to bind epitope-tagged forms of an LP polypeptide. Various methods of protein purification may be employed and such methods are known in the art and described, for example, in Deutscher, Methods in Enzymology 182:83-9 (1990) and Scopes, Protein Purification: Principles and Practice, Springer-Verlag, NY (1982). purification step(s) selected will depend, for example, on the nature of the production process used and the particular LP polypeptide produced.

Nucleotide sequences (or their complement) encoding LP polypeptides have various applications in the art of molecular biology, including uses as hybridization probes,

in chromosome and gene mapping and in the generation of anti-sense RNA and DNA. LP polypeptide-encoding nucleic acids will also be useful for the preparation of LP polypeptides by the recombinant techniques described herein.

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The full-length LP polypeptide-encoding nucleotide sequence (e.g., SEQ ID NO:1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, or 37), or portions thereof, may be useful as hybridization probes for probing a cDNA or genomic library to isolate the full-length LP polypeptide-encoding cDNA or genomic sequences including promoters, enhancer elements and introns of native sequence LP polypeptide-encoding DNA or to isolate still other genes (for instance, those encoding naturally-occurring variants of LP polypeptides or the same from other species) which have a desired sequence identity to the LP polypeptideencoding nucleotide sequence disclosed in SEQ ID NO:1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, or 37. Hybridization techniques are well known in the art and some of which are described in further detail in the Examples below.

Other useful fragments of the LP polypeptide-encoding nucleic acids include anti-sense or sense oligonucleotides comprising a single-stranded nucleic acid sequence (either RNA or DNA) capable of binding to target LP polypeptide-encoding mRNA (sense) of LP polypeptide-encoding DNA (anti-sense) sequences. Anti-sense or sense oligonucleotides, according to the present invention, comprise a fragment of the coding region of LP polypeptide-encoding DNA. Such a fragment generally comprises at least about 14 nucleotides, preferably from about 14 to 30 nucleotides. The ability to derive an anti-sense or a sense oligonucleotide, based upon

a cDNA sequence encoding a given protein is described in, for example, Stein and Cohen, Cancer Res. 48(10):2659-68 (1988) and van der Krol, et al., Bio/Techniques 6(10):958-76 (1988).

Binding of anti-sense or sense oligonucleotides to target nucleic acid sequences results in the formation of duplexes that block transcription or translation of the target sequence by one of several means, including enhanced degradation of the duplexes, premature termination of transcription or translation, or by other means. The anti-10 sense oligonucleotides thus may be used to block expression of LP mRNA and therefore any LP polypeptide encoded thereby. Anti-sense or sense oligonucleotides further comprise oligonucleotides having modified sugarphosphodiester backbones (or other sugar linkages, such as 15 those described in WO 91/06629) and wherein such sugar linkages are resistant to endogenous nucleases. oligonucleotides with resistant sugar linkages are stable in vivo (i.e., capable of resisting enzymatic degradation) but retain sequence specificity to be able to bind to 20 target nucleotide sequences.

Other examples of sense or anti-sense oligonucleotides include those oligonucleotides which are covalently linked to organic moieties, such as those described in WO 90/10448, and other moieties that increase affinity of 25 the oligonucleotide for a target nucleic acid sequence, such poly-L-lysine. Further still, intercalating agents, such as ellipticine, and alkylating agents or metal complexes may be attached to sense or anti-sense oligonucleotides to modify binding specificities of the 30 anti-sense or sense oligonucleotide for the target nucleotide sequence.

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Anti-sense or sense oligonucleotides may be introduced into a cell containing the target nucleic acid sequence by any gene transfer method, including, for example, calcium phosphate-mediated DNA transfection, electroporation, or by using gene transfer vectors such as Epstein-Barr virus. In a preferred procedure, an anti-sense or sense oligonucleotide is inserted into a suitable retroviral vector. A cell containing the target nucleic acid sequence is contacted with the recombinant retroviral vector, either *in vivo* or ex vivo. Suitable retroviral vectors include, but are not limited to, those derived from the murine retrovirus M-MSV, N2 (a retrovirus derived from M-MuLV), or the double copy vectors designated CDTSA, CTSB and DCTSC (see WO 90/13641).

Alternatively, a sense or an anti-sense oligonucleotide may be introduced into a cell containing the target nucleic acid sequence by formation of an oligonucleotidelipid complex, as described in WO 90/10448. The sense or anti-sense oligonucleotide-lipid complex is preferably dissociated within the cell by an endogenous lipase.

When the amino acid sequence for an LP polypeptide encodes a protein which binds to another protein (for example, where the LP polypeptide functions as a receptor), the LP polypeptide can be used in assays to identify the other proteins or molecules involved in the binding interaction. By such methods, inhibitors of the receptor/ligand binding interaction can be identified. Proteins involved in such binding interactions can also be used to screen for peptide or small molecule inhibitors or agonists of the binding interaction. Also, the receptor LP polypeptide can be used to isolate correlative ligand(s). Screening assays can be designed to find lead compounds that mimic the biological activity of the LP polypeptides

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disclosed herein or a receptor for such LP polypeptides. Typical screening assays will include assays amenable to high-throughput screening of chemical libraries, making them particularly suitable for identifying small molecule drug candidates. Small molecules contemplated include synthetic organic or inorganic compounds. The assays can be performed in a variety of formats, including protein-protein binding assays, biochemical screening assays, immunoassays and cell based assays, which are well characterized in the art.

Nucleic acids which encode an LP polypeptide of the present invention or any of its modified forms can also be used to generate either transgenic animals or "knockout" animals which, in turn, are useful in the development and screening of therapeutically useful reagents. Methods for generating transgenic animals, particularly animals such as mice or rats, have become conventional in the art and are described, for example, in U.S. Patents 4,736,866 and 4,870,009. Typically, particular cells would be targeted for an LP transgene incorporation with tissue-specific Transgenic animals that include a copy of a enhancers. transgene introduced into the germ line of the animal at an embryonic stage can be used to examine the effect of increased expression of DNA encoding an LP polypeptide. Such animals can be used as tester animals for reagents thought to confer protection from, for example, pathological conditions associated with its overexpression. In accordance with this facet of the invention, an animal is treated with the reagent and a reduced incidence of the pathological condition, compared to untreated animals. bearing the transgene, would indicate a potential therapeutic intervention for the pathological condition.

Alternatively, non-human homologs of LP polynucleotides can be used to construct a "knockout" animal which has a defective or altered gene encoding a particular LP polypeptide as a result of homologous recombination between the endogenous gene encoding the LP polypeptide and the altered genomic DNA introduced into an embryonic cell of the animal. For example, cDNA encoding an LP polypeptide can be used to clone genomic DNA encoding that LP polypeptide in accordance with established techniques. A portion of the genomic DNA encoding an LP 10 polypeptide can be deleted or replaced with another gene, such as a gene encoding a selectable marker which can be used to monitor integration. Typically, several kilobases of unaltered flanking DNA (both at the 5' and 3' ends) are included in the vector [see, e.g., Thomas and Capecchi, 15 Cell 51(3):503-12 (1987) for a description of homologous recombination vectors]. The vector is introduced into an embryonic stem cell line (e.g., by electroporation), and cells in which the introduced DNA has homologously recombined with the endogenous DNA are selected [see, e.g., 20 Li, et al., Cell 69(6):915-26 (1992)]. The selected cells are then injected into a blastocyst of an animal (e.g., a mouse or rat) to form aggregation chimeras [see, e.g., Bradley, Teratocarcinomas and Embryonic Stem Cells: A Practical Approach, E. J. Robertson, ed., pp. 113-152 (IRL, 25 Oxford, 1987)]. A chimeric embryo can then be implanted into a suitable pseudopregnant female foster animal and the embryo brought to term to create a "knockout" animal. Progeny harboring the homologously recombined DNA in their germ cells can be identified by standard techniques and 30 used to breed animals in which all cells of the animal contain the homologously recombined DNA. Knockout animals

can be characterized, for instance, for their ability to defend against certain pathological conditions and for their development of pathological conditions due to absence of the native LP polypeptide.

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Transgenic non-human mammals are useful as an animal models in both basic research and drug development endeavors. Transgenic animals expressing at least one LP polypeptide or nucleic acid can be used to test compounds or other treatment modalities which may prevent, suppress, or cure a pathology or disease associated with at least one of the above mentioned activities. Such transgenic animals can also serve as a model for the testing of diagnostic methods for those same diseases. Furthermore, tissues derived from such transgenic non-human mammals are useful as a source of cells for cell culture in efforts to develop in vitro bioassays to identify compounds that modulate LP polypeptide activity or LP polypeptide dependent signaling. Accordingly, another aspect of the present invention contemplates a method of identifying compounds efficacious in the treatment of at least one previously described disease or pathology associated with an LP polypeptide associated activity. A non-limiting example of such a method comprises:

- a) generating a transgenic non-human animal which expresses an LP polypeptide of the present invention and which is, as compared to a wild-type animal, pathologically distinct in some detectable or measurable manner from wild-type version of said non-human mammal;
 - b) exposing said transgenic animal to a compound, and;
- c) determining the progression of the pathology in the treated transgenic animal, wherein an arrest, delay, or reversal in disease progression in transgenic animal treated

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with said compound as compared to the progression of the pathology in an untreated control animals is indicative that the compound is useful for the treatment of said pathology.

Another embodiment of the present invention provides a method of identifying compounds capable of inhibiting LP polypeptide activity in vivo and/or in vitro wherein said method comprises:

- a) administering an experimental compound to an LP polypeptide expressing transgenic non-human animal, or tissues derived therefrom, exhibiting one or more physiological or pathological conditions attributable to the expression of an LP transgene; and
- b) observing or assaying said animal and/or animal tissues to detect changes in said physiological or pathological condition or conditions.

Another embodiment of the invention provides a method for identifying compounds capable of overcoming deficiencies in LP polypeptide activity in vivo or in vitro wherein said method comprises:

- a) administering an experimental compound to an LP polypeptide expressing transgenic non-human animal, or tissues derived therefrom, exhibiting one or more physiological or pathological conditions attributable to the disruption of the endogenous LP polypeptide-encoding gene; and
 - b) observing or assaying said animal and/or animal tissues to detect changes in said physiological or pathological condition or conditions.

Various means for determining a compound's ability to

30 modulate the activity of an LP polypeptide in the body of
the transgenic animal are consistent with the invention.

Observing the reversal of a pathological condition in the LP

polypeptide expressing transgenic animal after administering a compound is one such means. Another more preferred means is to assay for markers of LP activity in the blood of a transgenic animal before and after administering an experimental compound to the animal. The level of skill of an artisan in the relevant arts readily provides the practitioner with numerous methods for assaying physiological changes related to therapeutic modulation of LP activity.

"Gene therapy" includes both conventional gene 10 therapy, where a lasting effect is achieved by a single treatment, and the administration of gene therapeutic agents, which involves the one time or repeated administration of a therapeutically effective DNA or mRNA. Anti-sense RNAs and DNAs can be used as therapeutic agents 15 for blocking the expression of certain genes in vivo. has already been shown that short anti-sense oligonucleotides can be imported into cells where they act as inhibitors, despite their low intracellular concentrations caused by their restricted uptake by the cell membrane 20 [Zamecnik, et al., Proc. Natl. Acad Sci. USA 83(12):4143-6 (1986)]. The oligonucleotides can be modified to enhance their uptake, e.g., by substituting their negatively charged phosphodiester groups with uncharged groups.

There are a variety of techniques available for introducing nucleic acids into viable cells. The techniques vary depending upon whether the nucleic acid is transferred into cultured cell in vitro or in vivo in the cells of the intended host. Techniques suitable for the transfer of nucleic acid into mammalian cells in vitro include the use of liposomes, electroporation, microinjection, cell fusion, DEAE-dextran, the calcium phosphate

precipitation method, etc. The currently preferred in vivo gene transfer techniques include transfection with viral (typically, retroviral) vectors and viral coat proteinliposome mediated transfection [Dzau, et al., Trends in 5 Biotechnology 11(5):205-10 (1993)]. In some situations it is desirable to provide the nucleic acid source with an agent that targets the target cells, such as an antibody specific for a cell surface membrane protein or the target cell, a ligand for a receptor on the target cells, etc. 10 Where liposomes are employed, proteins which bind to a cell surface membrane protein associated with endocytosis may by used for targeting and/or to facilitate uptake, e.g., capsid proteins or fragments thereof trophic for a particular cell type, antibodies for proteins which undergo 15 internalization in cycling, proteins that target intracellular localization and enhance intracellular halflife. The technique of receptor-mediated endocytosis is described, for example, by Wu, et al., J. Biol. Chem. 262(10):4429-32 (1987); and Wagner, et al., Proc. Natl. Acad. Sci. USA 87(9):3410-4 (1990). For a review of gene 20 marking and gene therapy protocols, see Anderson, Science 256(5058):808-13 (1992).

The nucleic acid molecules encoding LP polypeptides or fragments thereof described herein are useful for chromosome identification. In this regard, there exists an ongoing need to idenfity new chromosome markers, since relatively few chromosome marking reagents, based upon actual sequence data, are presently available. Each LP polypeptide-encoding nucleic acid molecule of the present invention can be used as a chromosome marker. An LP polypeptide-encoding nucleic acid or fragments thereof can

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also be used for chromosomal localization of the gene encoding that LP polypeptide.

The present invention further provides anti-LP polypeptide antibodies. Exemplary antibodies include polyclonal, monoclonal, humanized, bispecific, and heteroconjugate antibodies.

The anti-LP polypeptide antibodies of the present invention may comprise polyclonal antibodies. Methods of preparing polyclonal antibodies are known to the skilled 10 artisan. Polyclonal antibodies can be raised in a mammal, for example, by one or more injections of an immunizing agent and, if desired, an adjuvant. Typically, the immunizing agent and/or adjuvant will be injected in the mammal by multiple subcutaneous or intraperitoneal 15 The immunizing agent may include the LP injections. polypeptide or a fusion protein thereof. It may be useful to conjugate the immunizing agent to a protein known to be immunogenic in the mammal being immunized. Examples of such immunogenic proteins include, but are not limited to, keyhole limpet hemocyanin, serum albumin, bovine 20 thyroglobulin, and soybean trypsin inhibitor. Examples of adjuvants which may be employed include Freund's complete adjuvant and MPL-TDM adjuvant (monophosphoryl Lipid A, synthetic trehalose dicorynomycolate). The immunization 25 protocol may be selected by one skilled in the art without undue experimentation.

The anti-LP polypeptide antibodies may, alternatively, be monoclonal antibodies. Monoclonal antibodies may be prepared using hybridoma methods, such as those described by Kohler and Milstein, *Nature* 256(5517):495-7 (1975). In a hybridoma method, a mouse, hamster, or other appropriate host animal is typically immunized with an immunizing agent

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to elicit lymphocytes that produce or are capable of producing antibodies that will specifically bind to the immunizing agent. Alternatively, the lymphocytes may be immunized in vitro.

The immunizing agent will typically include an LP polypeptide or a fusion protein thereof. Generally, either peripheral blood lymphocytes ("PBLs") are used, if cells of human origin are desired, or spleen cells or lymph node cells are used, if non-human mammalian sources are desired. The lymphocytes are then fused with an immortalized cell line using a suitable fusing agent, such as polyethylene glycol, to form a hybridoma cell [Goding, Monoclonal Antibodies: Principles and Practice, Academic Press, pp. 59-103 (1986)]. Immortalized cell lines are usually transformed mammalian cells, particularly myeloma cells of rodent, bovine and human origin. Usually, rat or mouse myeloma cell lines are employed. The hybridoma cells may be cultured in a suitable culture medium that preferably contains one or more substances that inhibit the growth or survival of the unfused, immortalized cells. For example, if the parental cells lack the enzyme hypoxanthine guanine phosphoribosyl transferase (HGPRT or HPRT), the culture medium for the hybridomas typically will include hypoxanthine, aminopterin, and thymidine ("HAT medium"), which prevents the growth of HGPRT-deficient cells.

Preferred immortalized cell lines are those that fuse efficiently, support stable high level expression of antibody by the selected antibody-producing cells, and are sensitive to a medium such as HAT medium. More preferred immortalized cell lines are murine myeloma lines, which can be obtained, for instance, from the Salk Institute Cell Distribution Center, San Diego, CA, and the American Type

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Culture Collection (ATCC), Rockville, MD. Human myeloma and mouse-human heteromyeloma cell lines also have been described for the production of human monoclonal antibodies [Kozbor, J. Immunol. 133(6):3001-5 (1984); Brodeur, et al., Monoclonal Antibody Production Techniques and Applications, Marcel Dekker, Inc., NY, pp. 51-63 (1987)].

The culture medium in which the hybridoma cells are cultured can then be assayed for the presence of monoclonal antibodies directed against an LP polypeptide. Preferably, the binding specificity of monoclonal antibodies produced by the hybridoma cells is determined by immunoprecipitation or by an *in vitro* binding assay, such as radioimmunoassay (RIA) or enzyme-linked immunoabsorbent assay (ELISA). Such techniques and assays are known in the art. The binding affinity of the monoclonal antibody can, for example, be determined by the Scatchard analysis of Munson and Rodbard, Anal. Biochem. 107(1):220-39 (1980).

After the desired hybridoma cells are identified, the clones may be subcloned by limiting dilution procedures and grown by standard methods [Goding, Monoclonal Antibodies: Principles and Practice, Academic Press, pp. 59-103 (1986)]. Suitable culture media for this purpose include, for example, Dulbecco's Modified Eagle's Medium and RPMI-1640 medium. Alternatively, the hybridoma cells may be grown in vivo as ascites in a mammal.

The monoclonal antibodies may also be made by recombinant DNA methods, such as those described in U.S. Patent 4,816,567. DNA encoding the monoclonal antibodies of the invention can be readily isolated and sequenced using conventional procedures (e.g., by using oligonucleotide probes that are capable of binding specifically to genes encoding the heavy and light chains

of murine antibodies). The hybridoma cells of the invention serve as a preferred source of such DNA. Once isolated, the DNA may be placed into expression vectors, which are then transfected into host cells such as simian COS cells, Chinese hamster ovary (CHO) cells, or myeloma cells that do not otherwise produce immunoglobulin protein, to obtain the synthesis of monoclonal antibodies in the recombinant host cells. The DNA also may be modified, for example, by substituting the coding sequence for human heavy and light chain constant domains in place of the 10 homologous murine sequences [U.S. Patent 4,816,567; Morrison, et al., Proc. Natl. Acad. Sci. USA 81(21):6851-5 (1984)] or by covalently joining to the immunoglobulin coding sequence all or part of the coding sequence for a non-immunoglobulin polypeptide. Such a non-immunoglobulin 15 polypeptide can be substituted for the constant domains of an antibody of the invention or can be substituted for the variable domains of one antigen-combining site of an antibody of the invention to create a chimeric bivalent 20 antibody.

The antibodies may be monovalent antibodies. Methods for preparing monovalent antibodies are well known in the art. For example, one method involves recombinant expression of immunoglobulin light chain and modified heavy chain. The heavy chain is truncated generally at any point in the Fc region so as to prevent heavy chain crosslinking. Alternatively, the relevant cysteine residues are substituted with another amino acid residue or are deleted so as to prevent crosslinking.

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30 In vitro methods are also suitable for preparing monovalent antibodies. Digestion of antibodies to produce

fragments thereof, particularly Fab fragments, can be accomplished using routine techniques known in the art.

The anti-LP polypeptide antibodies of the invention may further comprise humanized antibodies or human 5 antibodies. Humanized forms of non-human (e.g., murine) antibodies are chimeric immunoglobulins, immunoglobulin chains or fragments thereof (such as Fv, Fab, Fab', F(ab')2 or other antigen-binding subsequences of antibodies) which contain minimal sequence derived from non-human 10 immunoglobulin. Humanized antibodies include human immunoglobulins (recipient antibody) in which residues from a complementary-determining region (CDR) of the recipient are replaced by residues from a CDR of a non-human species (donor antibody) such as mouse, rat or rabbit having the desired specificity, affinity and capacity. In some instances, Fv framework residues of the human immunoglobulin are replaced by corresponding non-human residues. Humanized antibodies may also comprise residues which are found neither in the recipient antibody nor in the imported 20 CDR or framework sequences. In general, the humanized antibody will comprise substantially all of at least one, and typically two, variable domains, in which all or substantially all of the CDR regions correspond to those of a non-human immunoglobulin, and all or substantially all of 25 the FR regions are those of a human immunoglobulin consensus sequence. The humanized antibody optimally also will comprise at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin [Jones, et al., Nature 321(6069):522-5 30 (1986); Riechmann, et al., Nature 332(6162):323-7 (1988); and Presta, Curr. Op. Struct. Biol. 2:593-6 (1992)].

Methods for humanizing non-human antibodies are well known in the art. Generally, a humanized antibody has one or more amino acid residues introduced into it from a source which is non-human. These non-human amino acid residues are often referred to as "import" residues, which 5 are typically taken from an "import" variable domain. Humanization can be essentially performed following the method of Winter and co-workers [Jones, et al., Nature 321(6069):522-5 (1986); Riechmann, et al., Nature 332(6162):323-7 (1988); Verhoeyen, et al., Science 10 239(4847):1534-6 (1988)], by substituting rodent CDRs or CDR sequences for the corresponding sequences of a human antibody. Accordingly, such "humanized" antibodies are chimeric antibodies (U.S. Patent 4,816,567) wherein substantially less than an intact human variable domain has 15 been substituted by the corresponding sequence from a nonhuman species. In practice, humanized antibodies are typically human antibodies in which some CDR residues and possibly some FR residues are substituted by residues from analogous sites in rodent antibodies. 20

Human antibodies can also be produced using various techniques known in the art, including phage display libraries [Hoogenboom and Winter, J. Mol. Biol. 227(2):381-8 (1992); Marks, et al., J. Mol. Biol. 222(3):581-97 (1991)]. The techniques of Cole, et al., and Boerner, et al., are also available for the preparation of human monoclonal antibodies (Cole, et al., Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, p. 77 (1985), and Boerner, et al., J. Immunol. 147(1):86-95 (1991)].

30 Similarly, human antibodies can be made by introducing human immunoglobulin loci into transgenic animals, e.g., mice in which the endogenous immunoglobulin genes have been

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partially or complete inactivated. Upon challenge, human antibody production is observed, which closely resembles that seen in humans in all respects, including gene rearrangement, assembly and antibody repertoire. This approach is described, for example, in U.S. Patents 5 5,545,807; 5,545,806; 5,569,825; 5,625,126; 5,633,425; 5,661,016, and in the following scientific publications: Marks, et al., Biotechnology 10(7):779-83 (1992); Lonberg, et al., Nature 368(6474):856-9 (1994); Morrison, Nature 368(6474):812-3 (1994); Fishwild, et al., Nature 10 Biotechnology 14(7):845-51 (1996); Neuberger, Nature Biotechnology 14(7):826 (1996); Lonberg and Huszar, Int. Rev. Immunol. 13(1):65-93 (1995).

Bispecific antibodies are monoclonal, preferably human or humanized, antibodies that have binding specificities 15 for at least two different antigens. In the present case, one of the binding specificities is for an LP polypeptide, the other one is for any other antigen, and preferably for a cell-surface protein or receptor or receptor subunit. Methods for making bispecific antibodies are known in the 20 art. Antibodies with more than two valencies are contemplated. For example, trispecific antibodies can be prepared [Tutt, et al., J Immunol. 147(1):60-9 (1991)].

Heteroconjugate antibodies are also within the scope of the present invention. Heteroconjugate antibodies are 25 composed of two covalently joined antibodies. Such antibodies have, for example, been proposed to target immune system cells to unwanted cells [U.S. Patent 4,676,980], and for treatment of HIV infection [WO 91/00360; WO 92/20373]. It is contemplated that the 30 antibodies may be prepared in vitro using known methods in synthetic protein chemistry, including those involving

crosslinking agents. For example, immunotoxins may be constructed using a disulfide exchange reaction or by forming a thioether bond. Examples of suitable reagents for this purpose include iminothiolate and methyl-4-mercaptobutyrimidate and those disclosed, for example, in U.S. Patent 4,676,980.

The invention also pertains to immunoconjugates comprising an antibody conjugated to a cytotoxic agent such as a chemotherapeutic agent, toxin (e.g., an enzymatically active toxin of bacterial, fungal, plant or animal origin, or fragments thereof, or a small molecule toxin), or a radioactive isotope (i.e., a radioconjugate).

Conjugates of the antibody and cytotoxic agent are made using a variety of bifunctional protein coupling agents such as N-succinimidy1-3-(2-pyridyldithiol) 15 propionate (SPDP), iminothiolane (IT), bifunctional derivatives of imidoesters (such as dimethyl adipimidate HCL), active esters (such as disuccinimidyl suberate), aldehydes (such as glutaraldehyde), bis-azido compounds, bis-diazonium derivatives (such as bis-2-diazoniumbenzoyl)-20 ethylenediamine) diisocyanates (such as tolylene-2,6diisocyanate), and bioactive fluorine compounds. example, a ricin immunotoxin can be prepared as described in Vitetta, et al., Science 238(4830):1098-104 (1987). Carbon-14-labeled 1-isothiocyanatobenzyl-3-methyldiethylene 25 triaminepentaacetic acid (MX-DTPA) is an exemplary chelating agent for conjugation of radionucleotide to the antibody.

In another embodiment, the antibody may be conjugated to a "receptor" (such as streptavidin) for utilization in tumor pretargeting wherein the antibody-receptor conjugate is administered to the patient, followed by removal of

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unbound conjugate from the circulation using a clearing agent, and then administration of a "ligand" (e.g., avidin) which is conjugated to a cytotoxic agent (e.g., a radionucleotide).

The antibodies disclosed herein may also be formulated as immunoliposomes. Liposomes containing the antibody are prepared by methods known in the art, such as described in Eppstein, et al., Proc. Natl. Acad. Sci. USA 82:3688-92 (1985); Hwang, et al., Proc. Natl. Acad. Sci. USA 77(7):4030-4 (1980); and U.S. Patents 4,485,045 and 4,544,545. Liposomes with enhanced circulation time are disclosed in U.S. Patent 5,013,556.

Particularly useful liposomes can be generated by the reverse phase evaporation method with a lipid composition comprising phosphatidylcholine, cholesterol and PEG-derivatized phosphatidylethanolamine (PEG-PE). Liposomes are extruded through filters of defined pore size to yield liposomes with the desired diameter. Fab' fragments of the antibody of the present invention can be conjugated to the liposomes as described in Martin, et al., J. Biol. Chem. 257(1):286-8 (1982) via a disulfide interchange reaction. A chemotherapeutic agent (such as Doxorubicin) is optionally contained within the liposome. See Gabizon, et al., J. National Cancer Inst. 81(19):484-8 (1989).

Antibodies specifically binding an LP polypeptide identified herein, as well as other molecules identified by the screening assays disclosed hereinbefore, can be administered for the treatment of various disorders in the form of pharmaceutical compositions.

30 If an LP polypeptide is intracellular and whole antibodies are used as inhibitors, internalizing antibodies are preferred. However, lipofections or liposomes can also

be used to deliver the antibody or an antibody fragment into cells. Where antibody fragments are used, the smallest inhibitory fragment that specifically binds to the binding domain of the target protein is preferred. For example, based upon the variable-region sequences of an antibody, peptide molecules can be designed that retain the ability to bind the target protein sequence. Such peptides can be synthesized chemically and/or produced by recombinant DNA technology. See, e.g., Marasco, et al., Proc. Natl. Acad. Sci. USA 90(16):7889-93 (1993).

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The formulation herein may also contain more than one active compound as necessary for the particular indication being treated, preferably those with complementary activities that do not adversely affect each other.

Alternatively, or in addition, the composition may comprise an agent that enhances its function, such as, for example,

an agent that enhances its function, such as, for example, a cytotoxic agent, cytokines, chemotherapeutic agent, or growth-inhibitory agent. Such molecules are suitable present in combination in amounts that are effective for the purpose intended. The active ingredients may also be entrapped in microcapsules prepared, for example, by coacervation techniques or by interfacial polymerization, for example, hydroxymethylcellulose or gelatin-microcapsules and poly-(methylmethacrylate) microcapsules, respectively, in colloidal drug delivery systems (for

example, liposomes, albumin microspheres, microemulsions, nano-particles, and nanocapsules) or in macroemulsions.

Such techniques are disclosed in Remington's Pharmaceutical Sciences 16th edition (1980).

The formulations to be used for *in vivo* administration must be sterile. This is readily accomplished by filtration through sterile filtration membranes.

Sustained-release preparations may be prepared. Suitable examples of sustained-release preparations include semipermeable matrices of solid hydrophobic polymers containing the antibody, which matrices are in the form of shaped articles, e.g., films, or microcapsules. Examples 5 of sustained-release matrices include polyesters, hydrogels (for example, poly(2-hydroxyethyl-methacrylate), or poly(vinylalcohol), polylactides (U.S. Patent 3,773,919), copolymers of L-glutamic acid gamma-ethyl-L-glutamate, non-10 degradable ethylene-vinylacetate, degradable lactic acidglycolic acid copolymers such as the LUPRON DEPOTTM (injectable microspheres composed of lactic acid-glycolic acid copolymer and leuprolide acetate), and poly-D-(-)3hydroxylbutyric acid. While polymers such as ethylene-15 vinyl acetate and lactic acid-glycolic acid enable release of molecules for over 100 days, certain hydrogels release proteins for shorter time periods. When encapsulated antibodies remain in the body for a long time, they may denature or aggregate as a result of exposure to moisture 20 at 37 degrees C, resulting in a loss of biological activity and possible changes in immunogenicity. Rational strategies can be devised for stabilization depending on the mechanisms involved. For example, if the aggregation mechanism is discovered to be intermolecular S-S bond 25 formation through thiosulfide interchange, stabilization may be achieved by modifying sulfhydryl residues, lyophilizing from acidic solutions, controlling moisture content, using appropriate additives, and developing specific polymer matrix compositions.

The anti-LP polypeptide antibodies of the present invention have various utilities. For example, such antibodies may be used in diagnostic assays for LP

polypeptide expression, e.g., detecting expression in specific cells, tissues, or serum. Various diagnostic assay techniques known in the art may be used, such as competitive binding assays, direct or indirect sandwich assays and immunoprecipitation assays conducted in either 5 heterogeneous or homogeneous phases [Zola, Monoclonal Antibodies: A Manual of Techniques, CRC Press, Inc., pp. 147-158 (1987)]. The antibodies used in the assays can be labeled with a detectable moiety. The detectable moiety 10 should be capable of producing, either directly or indirectly, a detectable signal. For example, the detectable moiety may be a radioisotope, such as 3H, 14C, 32 P, 35 S, or 125 I, a fluorescent or chemiluminescent compound, such as fluorescein isothiocyanate, rhodamine, or luciferin, or an enzyme, such as alkaline phosphatase, 15 beta-galactosidase or horseradish peroxidase. Any method known in the art for conjugating the antibody to the detectable moiety may be employed, including those methods described by Hunter, et al., Nature 144:945 (1962); David, 20 et al., Biochemistry 13(5):1014-21 (1974); Pain, et al., J Immunol. Meth., 40(2):219-30 (1981); and Nygren, J. Histochem. Cytochem. 30(5):407-12 (1982).

Anti-LP polypeptide antibodies also are useful for affinity purification from recombinant cell culture or natural sources. In this process, the antibodies are immobilized on a suitable support, such a Sephadex® resin or filter paper, using methods well known in the art. The immobilized antibody is then contacted with a sample containing the LP polypeptide to be purified, and thereafter the support is washed with a suitable solvent that will remove substantially all the material in the sample except the LP polypeptide, which is bound to the

immobilized antibody. Finally, the support is washed with another suitable solvent that will release the desired polypeptide from the antibody.

This invention encompasses methods of screening compounds to identify those that mimic the activity of the 5 LP polypeptide (agonists) disclosed herein or prevent the effects of the LP polypeptide (antagonists). Screening assays for antagonist drug candidates are designed to identity compounds that bind or complex with an LP polypeptide encoded by the genes identified herein or 10 otherwise interfere with the interaction of LP polypeptides with other cellular proteins. Such screening assays will include assays amenable to high-throughput screening of chemical libraries, making them particularly suitable for 15 identifying small molecule drug candidates.

The assays can be performed in a variety of formats. In binding assays, the interaction is binding, and the complex formed can be isolated or detected in the reaction mixture. In a particular embodiment, an LP polypeptide 20 encoded by a gene identified herein or the drug candidate is immobilized on a solid phase, e.g., on a microtiter plate, by covalent or non-covalent attachments. covalent attachment generally is accomplished by coating the solid surface with a solution comprising LP polypeptide 25 and drying. Alternatively, an immobilized antibody, e.g., a monoclonal antibody, specific for the polypeptide to be immobilized can be used to anchor it to a solid surface. The assay is performed by adding the non-immobilized component, which may be labeled by a detectable label, to the immobilized component, e.g., the coated surface containing the anchored component. When the reaction is complete, the non-reacted components are removed, e.g., by

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washing, and complexes anchored on the solid surface are detected. When the originally non-immobilized component carries a detectable label, the detection of label immobilized on the surface indicates that complexing occurred. Where the originally non-immobilized component does not carry a label, complexing can be detected, for example, by using a labeled antibody specifically binding the immobilized complex.

If the candidate compound interacts with but does not bind to an LP polypeptide, its interaction with that 10 polypeptide can be assayed by methods well known for detecting protein-protein interactions. Such assays include traditional approaches, such as, e.g., crosslinking, co-immunoprecipitation, and co-purification through gradients or chromatographic columns. In addition, 15 protein-protein interactions can be monitored through gradients or chromatographic columns. In addition, protein-protein interactions can be monitored by using a yeast-based genetic system described by Fields and coworkers [Fields and Song, Nature 340(6230):245-6 (1989); 20 Chien, et al., Proc. Natl. Acad. Sci. USA 88(21):9578-82 (1991); Chevray and Nathans, Proc. Natl. Acad. Sci. USA 89(13):5789-93 (1992)]. Many transcriptional activators, such as yeast GAL4, consist of two physically discrete modular domains, one acting as the DNA-binding domain, 25 while the other functions as the transcription-activation The yeast expression system described in the foregoing publications (generally referred to as the "twohybrid system") takes advantage of this property, and employs two hybrid proteins, one in which the target protein is fused to the DNA-binding domain of GAL4, and another in which candidate activating proteins are fused to

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the activation domain. The expression of GAL1-lacZ reporter gene under control of a GAL4-activated promoter depends on reconstitution of GAL4 activity via protein-protein interaction. Colonies containing interacting polypeptides are detected with chromogenic substrate for beta-galactosidase. A complete kit (MATCHMAKERTM) for identifying protein-protein interactions between two specific proteins using the two-hybrid technique is commercially available from Clontech. This system can also be extended to map protein domains involved in specific protein interactions as well as to pinpoint amino acid residues that are crucial for these interactions.

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Compounds that interfere with the interaction of an LP polypeptide identified herein and other intra- or 15 extracellular components can be tested as follows: usually a reaction mixture is prepared containing the product of the gene and the intra- or extracellular component under conditions and for a time allowing for the interaction and binding of the two products. To test the ability of a 20 candidate compound to inhibit binding, the reaction is run in the absence and in the presence of the test compound. In addition, a placebo may be added to a third reaction mixture to serve as a positive control. The binding (complex formation) between the test compound and the 25 intra- or extracellular component present in the mixture is monitored as described hereinabove. The formation of a complex in the control reaction(s) but not in the reaction mixture containing the test compound indicates that the test compound interferes with the interaction of the test compound and its reaction partner. 30

Antagonists may be detected by combining at least one LP polypeptide and a potential antagonist with a membrane-

bound or recombinant receptor for that LP polypeptide under appropriate conditions for a competitive inhibition assay. The LP polypeptide can be labeled, such as by radioactivity, such that the number of LP polypeptide 5 molecules bound to the receptor can be used to determine the effectiveness of the potential antagonist. encoding the receptor for an LP polypeptide can be identified by numerous methods known to those of skill in the art, for example, ligand panning and FACS sorting. 10 Coligan, et al., Current Protocols in Immunology 1(2):Ch. 5 (1991). Preferably, expression cloning is employed such that polyadenylated mRNA is prepared from a cell responsive to the secreted form of a particular LP polypeptide, and a cDNA library created from this mRNA is divided into pools 15 and used to transfect COS cells or other cells that are not responsive to the secreted LP polypeptide. Transfected cells that are grown on glass slides are exposed to the labeled LP polypeptide. The LP polypeptide can be labeled by a variety of means including iodination or inclusion of 20 a recognition site for a site-specific protein kinase. Following fixation and incubation, the slides are subjected to autoradiographic analysis. Positive pools are identified and sub-pools are prepared and re-transfected using an interactive sub-pooling and re-screening process, 25 eventually yielding a single clone that encodes the putative receptor.

As an alternative approach for receptor identification, a labeled LP polypeptide can be photoaffinity-linked with cell membrane or extract preparations that express the receptor molecule. Cross-linked material is resolved by PAGE and exposed to X-ray film. The labeled complex containing the receptor can be excised, resolved into

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peptide fragments, and subjected to protein microsequencing. The amino acid sequence obtained from microsequencing would be used to design a set of degenerate oligonucleotide probes to screen a cDNA library to identify the gene encoding the putative receptor.

In another assay for antagonists, mammalian cells or a membrane preparation expressing the receptor would be incubated with a labeled LP polypeptide in the presence of the candidate compound. The ability of the compound to enhance or block this interaction could then be removed.

Alternatively, a potential antagonist may be a closely related protein, for example, a mutated form of the LP polypeptide that recognizes the receptor but imparts no effect, thereby competitively inhibiting the action of the polypeptide.

Another potential LP antagonist is an anti-sense RNA or DNA construct prepared using anti-sense technology, where, e.g., an anti-sense RNA or DNA molecule acts to block directly the translation of mRNA by hybridizing to 20 targeted mRNA and prevent its translation into protein. Anti-sense technology can be used to control gene expression through triple-helix formation or anti-sense DNA or RNA, both of which methods are based on binding of a polynucleotide to DNA or RNA. For example, the 5' coding 25 portion of the polynucleotide sequence, which encodes the mature form of an LP polypeptide can be used to design an anti-sense RNA oligonucleotide sequence of about 10 to 40 base pairs in length. A DNA oligonucleotide is designed to be complementary to a region of the gene involved in transcription [triple helix; see Lee, et al., Nucl. Acids Res 6(9):3073-91 (1979); Cooney, et al., Science 241(4864):456-9 (1988); Beal and Dervan, Science

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251(4999):1360-3 (1991)], thereby preventing transcription and production of the LP polypeptide. The anti-sense RNA oligonucleotide hybridizes to the mRNA in vivo and blocks translation of the mRNA molecules [anti-sense; see Okano, J. Neurochem. 56(2):560-7 (1991); Oligodeoxynucleotides as Anti-sense Inhibitors of Gene Expression (CRC Press: Boca Raton, FL 1988)]. The oligonucleotides described above can also be delivered to cells such that the anti-sense RNA or DNA may be expressed in vivo to inhibit production of the LP polypeptide. When anti-sense DNA is used, oligodeoxy-ribonucleotides derived from the translation-initiation site, e.g., between about -10 and +10 positions of the target gene nucleotide sequence, are preferred.

Potential antagonists include small molecules that bind to the active site, the receptor binding site, or growth factor or other relevant binding site of the LP polypeptide, thereby blocking the normal biological activity of the LP polypeptide. Examples of small molecules include, but are not limited to, small peptides or peptide-like molecules, preferably soluble peptides, and synthetic non-peptidyl organic or inorganic compounds.

Ribozymes are enzymatic RNA molecules capable of catalyzing the specific cleavage of RNA. Ribozymes act by sequence-specific hybridization to the complementary target RNA, followed by endonucleolytic cleavage. Specific ribozyme cleavage sites within a potential RNA target can be identified by known techniques. For further details, see, e.g., Rossi, Current Biology 4(5):469-71 (1994) and PCT publication No. WO 97/33551 (published September 18, 1997).

Nucleic acid molecules in triple-helix formation used to inhibit transcription should be single-stranded and

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composed of deoxynucleotides. The base composition of these oligonucleotides is designed such that it promotes triple-helix formation via Hoogsteen base-pairing rules, which generally require sizeable stretches of purines or pyrimidines on one strand of a duplex. For further details see, e.g., PCT publication No. WO 97/33551, supra.

Another use of the compounds of the invention (e.g., LP polypeptides, fragments and variants thereof and LP antibodies directed thereto) described herein is to help diagnose whether a disorder is driven to some extent by the modulation of signaling by an LP polypeptide

A diagnostic assay to determine whether a particular disorder is driven by LP polypeptide dependent signaling can be carried out using the following steps:

- a) culturing test cells or tissues expressing an LP polypeptide;
- b) administering a compound which can inhibit LP polypeptide dependent signaling; and
- c) measuring LP polypeptide mediated phenotypic effects 20 in the test cells.

The steps can be carried out using standard techniques in light of the present disclosure. Appropriate controls take into account the possible cytotoxic effect of a compound, such as treating cells not associated with a cell proliferative disorder (e.g., control cells) with a test compound and can also be used as part of the diagnostic assay. The diagnostic methods of the invention involve the screening for agents that modulate the effects of LP polypeptide-associated disorders.

The LP polypeptides or antibodies thereto as well as LP polypeptide antagonists or agonists can be employed as therapeutic agents. Such therapeutic agents are formulated

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according to known methods to prepare pharmaceutically useful compositions, whereby the LP polypeptide or antagonist or agonist thereof is combined in a mixture with a pharmaceutically acceptable carrier.

In the case of LP polypeptide antagonistic or 5 agonistic antibodies, if the LP polypeptide is intracellular and whole antibodies are used as inhibitors, internalizing antibodies are preferred. However, lipofections or liposomes can also be used to deliver the antibody, or an antibody fragment, into cells. Where 10 antibody fragments are used, the smallest inhibitory fragment which specifically binds to the binding domain of the target protein is preferred. For example, based upon the variable region sequences of an antibody, peptide molecules can be designed which retain the ability to bind 15 the target protein sequence. Such peptides can be synthesized chemically and/or produced by recombinant DNA technology [see, e.g., Marasco, et al., Proc. Natl. Acad. Sci. USA 90(16):7889-93 (1993)].

Therapeutic formulations are prepared for storage by mixing the active ingredient having the desired degree of purity with optional pharmaceutically acceptable carriers, excipients or stabilizers [Remington's Pharmaceutical Sciences 16th edition (1980)], in the form of lyophilized formulations or aqueous solutions.

The formulation herein may also contain more than one active compound as necessary for the particular indication being treated, preferably those with complementary activities that do not adversely affect each other. Such molecules are suitably present in combination in amounts that are effective for the purpose intended.

The active ingredients may also be entrapped in microcapsules prepared, for example, by coacervation techniques or by interfacial polymerization, for example, hydroxymethylcellulose or gelatin-microcapsules and polymethylmethacrylate) microcapsules, respectively, in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nano-particles and nanocapsules) or in macroemulsions. Such techniques are disclosed in Remington's Pharmaceutical Sciences, supra.

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The formulations to be used for *in vivo* administration must be sterile. This is readily accomplished by filtration through sterile filtration membranes.

Therapeutic compositions herein generally are placed into a container having a sterile access port, for example, and intravenous solution bag or vial having a stopper pierceable by a hypodermic injection needle.

Sustained-release preparations may be prepared. Suitable examples of sustained-release preparations include semipermeable matrices of solid hydrophobic polymers containing the therapeutic agent(s), which matrices are in the form of shaped articles, e.g., films, or microcapsules. Examples of sustained-release matrices include polyesters, hydrogels [for example, poly(2-hydroxyethyl-methacrylate), or poly(vinylalcohol)], polylactides, copolymers of Lglutamic acid and gamma-ethyl-L-glutamate, non-degradable ethylene-vinyl acetate, degradable lactic acid-glycolic acid copolymers such as the LUPRON DEPOT™ (injectable microspheres composed of lactic acid-glycolic acid copolymer and leuprolide acetate), and poly-D-(-)-3hydroxybutyric acid. Microencapsulation of recombinant proteins for sustained release has been successfully performed with human growth hormone (rhGH), interferon, and

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interleukin-2. Johnson, et al., Nat. Med. 2(7):795-9 (1996); Yasuda, et al., Biomed. Ther. 27:1221-3 (1993); Hora, et al., Bio/Technology 8(8):755-8 (1990); Cleland, "Design and Production of Single Immunization Vaccines Using Polylactide Polyglycolide Microsphere Systems" in Vaccine Design: The Subunit and Adjuvant Approach, Powell and Newman, Eds., Plenum Press, NY, 1995, pp. 439-462 WO 97/03692; WO 96/40072; WO 96/07399; and U.S. Patent 5,654,010.

The sustained-release formulations of these proteins may be developed using polylactic-coglycolic acid (PLGA) polymer due to its biocompatibility and wide range of biodegradable properties. The degradation products of PLGA, lactic and glycolic acids, can be cleared quickly within the human body. Moreover, the degradability of this polymer can be adjusted from months to years depending on its molecular weight and composition. See Lewis, "Controlled release of bioactive agents from lactide/glycolide polymer" in Biodegradable Polymers as

Drug Delivery Systems (Marcel Dekker; New York, 1990), M. Chasin and R. Langer (Eds.) pp. 1-41.

While polymers such as ethylene-vinyl acetate and lactic acid-glycolic acid enable release of molecules for over 100 days, certain hydrogels release proteins for shorter time periods. When encapsulated antibodies remain in the body for a long time, they may denature or aggregate as a result of exposure to moisture at 37 degrees C, resulting in a loss of biological activity and possible changes in immunogenicity.

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It is contemplated that the compounds, including, but not limited to, antibodies, small organic and inorganic molecules, peptides, anti-sense molecules, ribozymes, etc.,

of the present invention may be used to treat various conditions including those characterized by overexpression and/or activation of the disease-associated genes identified herein. The active agents of the present 5 invention (e.g., antibodies, polypeptides, nucleic acids, ribozymes, small organic or inorganic molecules) are administered to a mammal, preferably a human, in accord with known methods, such as intravenous administration as a bolus or by continuous infusion over a period of time, by 10 intramuscular, intraperitoneal, intracerebral, intracerobrospinal, subcutaneous, intra-articular, intrasynovial, intrathecal, intraoccular, intralesional, oral, topical, inhalation, pulmonary, and/or through sustained release.

Other therapeutic regimens may be combined with the administration of LP polypeptide antagonists or antagonists, anti-cancer agents, e.g., antibodies of the instant invention.

appropriate dosage of an active agent, (e.g., an antibody, polypeptide, nucleic acid, ribozyme, or small organic or inorganic molecule) will depend on the type of disease to be treated, as defined above, the severity and course of the disease, whether the agent is administered for preventive or therapeutic purposes, previous therapy, the patient's clinical history and response to the agent, and the discretion of the attending physician. The agent is suitably administered to the patient at one time or over a series of treatments.

Dosages and desired drug concentration of pharmaceutical compositions of the present invention may vary depending on the particular use envisioned. The

determination of the appropriate dosage or route of administration is well within the skill of an ordinary artisan. Animal experiments provide reliable guidance for the determination of effective does for human therapy.

Interspecies scaling of effective doses can be performed following the principles laid down by Mordenti and Chappell, "The Use of Interspecies Scaling in Toxicokinetics," in Toxicokinetics and New Drug Development, Yacobi, et al., Eds., Pergamon Press, NY, p.4246 (1989).

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When in vivo administration of a composition comprising an LP polypeptide, an LP polypeptide antibody, an LP polypeptide-encoding nucleic acid, ribozyme, or small organic or inorganic molecule is employed, normal dosage 15 amounts may vary from about 1 ng/kg up to 100 mg/kg of mammal body weight or more per day, preferably about 1 pg/kg/day up to 100 mg/kg of mammal body weight or more per day, depending upon the route of administration. Guidance as to particular dosages and methods of delivery 20 is provided in the literature; see, for example, U.S. Patents 4,657,760, 5,206,344 or 5,225,212. It is within the scope of the invention that different formulations will be effective for different treatment compounds and different disorders, that administration targeting one organ or tissue, for example, may necessitate delivery in a 25 manner different from that to another organ or tissue. Moreover, dosages may be administered by one or more separate administrations or by continuous infusion. For repeated administrations over several days or longer, depending on the condition, the treatment is sustained until a desired suppression of disease symptoms occurs. However, other dosage regimens may be useful. The progress of this therapy is easily monitored by conventional techniques and assays.

In another embodiment of the invention, an article of manufacture containing materials useful for the diagnosis or treatment of the disorders described above is provided. 5 The article of manufacture comprises a container and a Suitable containers include, for example, bottles, vials, syringes, and test tubes. The containers may be formed from a variety of materials such as glass or 10 plastic. The container holds a composition which is effective for diagnosing or treating the condition and may have a sterile access port (for example, the container may be an intravenous solution bag or a vial having a stopper pierceable by a hypodermic injection needle). The active 15 agent in the composition is typically an LP polypeptide, antagonist or agonist thereof. The label on, or associated with, the container indicates that the composition is used for diagnosing or treating the condition of choice. article of manufacture may further comprise a second 20 container comprising a pharmaceutically-acceptable buffer, such as phosphate-buffered saline, Ringer's solution and dextrose solution. It may further include other materials desirable from a commercial and user standpoint, including other buffers, diluents, filters, needles, syringes, and 25 package inserts with instructions for use.

In another embodiment of the invention, therapeutic utility of the LP polypeptide is determined by measuring phosphorylation of tyrosine residues on specific cell lines. The early cellular response of cells stimulated with the majority of proteins is protein phosphorylation of the tyrosine residues. This response includes autophosphorylation of corresponding receptors, which thereby leads

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to the activation of catalytic properties and the initiation of intracellular pathways specific to the cell type. Moreover, signaling downstream of receptors requires phosphorylation of specific kinases inside the cell and other intracellular enzymes of different origin as well as the phosphorylation of multiple adapter/scaffold, structural proteins and transcriptional factors. Therefore, diverse protein-induced cell responses can be visualized by monitoring the state of protein phosphorylation after cell stimulation.

Immunodetection is used to detect the protein phosphorylation of the stimulated cell. Several antibodies that are directed against specific phosphorylated epitopes in signaling molecules are readily available. Two specific antibodies are used: phosphospecific anti-MAPK and anti-AKT antibodies. Additionally, non-specific anti-phosphotyrosine antibodies, which recognize tyrosine-phosphorylated proteins, are used. While anti-phosphotyrosine antibodies allow detection of diverse tyrosine phosphorylated proteins without directly addressing the nature of their identity, the phosphospecific anti-MAPK and anti-AKT antibodies recognize only the corresponding proteins in their phosphorylated form.

Another assay to determine utility of LP polypeptides involves transfection of cell lines with reporter plasmids followed by cell stimulation with an LP polypeptide. Each reporter consists of a defined element, responsive to specific intracellular signaling pathways, upstream of a sequence involving a reporter protein such as luciferase. Upon stimulation of the element, reporter transcription and translation ensues, and the resulting protein levels can be detected using an assay such as a luminescence assay. The

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cell stimulation period depends on the reporter plasmid used.

For each reporter used, positive controls are designed in the form of agonist cocktails which include approximately 5 maximal stimulatory doses of several ligands known to stimulate the represented signaling pathway. Using this design, the chances of finding a positive stimulus for each cell line is maximized. The caveat, however, is that some cell line / reporter combinations will not be stimulated by 10 the specific agonist cocktail, due to lack of an appropriate ligand in the cocktail. Alternately, the lack of signal induction by an agonist cocktail may be the lack of all signaling components within a particular cell line to activate the transcriptional element. Cell line / reporter 15 combinations with no exogenous stimulus added make up the negative controls.

Another assay to determine utility of LP polypeptides involves transfection of cell lines with reporter plasmids followed by cell stimulation with an LP polypeptide. Each reporter consists of a defined element, responsive to specific intracellular signaling pathways, upstream of a sequence involving a reporter protein such as luciferase. Upon stimulation of the element, reporter transcription and translation ensues, and the resulting protein levels can be detected using an assay such as a luminescence assay. The cell stimulation period depends on the reporter plasmid used.

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For each reporter used, positive controls are designed in the form of agonist cocktails which include

30 approximately maximal stimulatory doses of several ligands known to stimulate the represented signaling pathway.

Using this design, the chances of finding a positive

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stimulus for each cell line is maximized. The caveat, however, is that some cell line/reporter combinations will not be stimulated by the specific agonist cocktail, due to lack of an appropriate ligand in the cocktail.

Alternately, the lack of signal induction by an agonist cocktail may be the lack of all signaling components within a particular cell line to activate the transcriptional element. Cell line / reporter combinations with no exogenous stimulus added make up the negative controls.

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In another assay, utility of LP polypeptide is determined by proliferation of cells. In this assay, gross changes in the number of cells remaining in a culture are monitored after exposure to an LP polypeptide for three days. The cells are incubated in an appropriate assay medium to produce a sub-optimal growth rate. For example, usually a 1:10 or 1:20 dilution of normal culture medium results in a 40 to 60% reduction in cell number compared to the undiluted culture medium. This broad growth zone is chosen so that if an LP polypeptide is a stimulator of growth, the cells have room to expand, and conversely, if the LP polypeptide is deleterious, a reduction in cell density can be detected. After a period of exposure, the assay media is replaced with media containing a detection agent such as Calcein AM, a membrane-permeant fluorescent dye, allowing quantification of the cell number.

For use in another assay, a FLAG-HIS (FLIS)-tagged version of the LP polypeptide is expressed using mammalian cells such as HEK-293EBNA, COS-7, or HEK293T. The coding region of the cDNA is amplified by PCR of a vector containing a fragment encoding the LP polypeptide. The PCR-generated fragment is cleaved with restriction enzymes and gel-purified. The fragment is then ligated into a

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mammalian expression vector containing the FLIS epitope tag fused to the C-terminus. Protein expressed by this plasmid construct includes both the FLAG tag (Asp-Tyr-Lys-Asp-Asp-Asp-Asp-Asp-Lys) and the 6X His tag at the COOH -terminus of the protein. This tag provides epitopes for commercially available tag-specific antibodies, enabling detection of the protein.

To determine expression of the LP polypeptide in tissues, a protein-binding assay is performed. The fixed tissue sample is exposed to the FLIS-tagged LP polypeptide, followed by exposure to a primary antibody and a secondary antibody containing a fluorescent dye. Tagged LP polypeptide that binds to the antigens in the tissue will fluoresce. Binding of the protein to an antigen in the tissue suggests that the protein is expressed in that tissue. Thus, protein expression can be determined by measuring which tissues fluoresce.

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Having generally described the invention, the same will be more readily understood by reference to the following examples, which are provided by way of illustration and are not intended as limiting.

EXAMPLES

Example 1

25 Expression and Purification of LP polypeptides in E. coli

The bacterial expression vector pQE60 is used for
bacterial expression in this example. (QIAGEN, Inc.,
Chatsworth, CA). pQE60 encodes ampicillin antibiotic
resistance ("Ampr") and contains a bacterial origin of
30 replication ("ori"), an IPTG inducible promoter, a ribosome
binding site ("RBS"), six codons encoding histidine residues

that allow affinity purification using nickel-nitrilo-

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triacetic acid ("Ni-NTA") affinity resin sold by QIAGEN,
Inc., and suitable single restriction enzyme cleavage sites.
These elements are arranged such that a DNA fragment
encoding a polypeptide can be inserted in such a way as to
produce that polypeptide with the six His residues (i.e., a
"6 X His tag") covalently linked to the carboxyl terminus of
that polypeptide. However, a polypeptide coding sequence
can optionally be inserted such that translation of the six
His codons is prevented and, therefore, a polypeptide is
produced with no 6 X His tag.

The nucleic acid sequence encoding the desired portion of an LP polypeptide lacking the hydrophobic leader sequence is amplified from a cDNA clone using PCR oligonucleotide primers (based on the sequences presented, e.g., in SEQ ID NO:1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, or 37) which anneal to the amino terminal encoding DNA sequences of the desired portion of the LP polypeptide-encoding nucleic acid and to sequences in the construct 3' to the cDNA coding sequence. Additional nucleotides containing restriction sites to facilitate cloning in the pQE60 vector are added to the 5' and 3' sequences, respectively.

For cloning, the 5' and 3' primers have nucleotides corresponding or complementary to a portion of the coding sequence of the LP polypeptide-encoding nucleic acid, e.g., as presented in SEQ ID NO:1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, or 37, according to known method steps. One of ordinary skill in the art would appreciate, of course, that the point in a polynucleotide sequence where the 5' primer begins can be varied to amplify a desired portion of the complete polypeptide-encoding

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polynucleotide shorter or longer than the polynucleotide which encodes the mature form of the polypeptide.

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The amplified nucleic acid fragments and the vector pQE60 are digested with appropriate restriction enzymes, and the digested DNAs are then ligated together. Insertion of the LP polypeptide-encoding DNA into the restricted pQE60 vector places the LP105, LP061, LP224, LP240, LP239(a), LP243(a), LP243(b), LP253, LP218, LP251(a), LP252, LP239(b), LP223(a), LP255(a), LP244, LP186, LP251(b), LP255(b), or LP223(b) polypeptide coding region including its associated stop codon downstream from the IPTG-inducible promoter and in-frame with an initiating AUG codon. The associated stop codon prevents translation of the six histidine codons downstream of the insertion point.

The ligation mixture is transformed into competent E. coli cells using standard procedures such as those described in Sambrook, et al., 1989; Ausubel, 1987-1998. E. coli strain M15/rep4, containing multiple copies of the plasmid pREP4, which expresses the lac repressor and confers kanamycin resistance ("Kanr"), is used in carrying out the 20 illustrative example described herein. This strain, which is only one of many that are suitable for expressing LP polypeptides, is available commercially from QIAGEN, Inc. Transformants are identified by their ability to grow on LB plates in the presence of ampicillin and kanamycin. Plasmid 25 DNA is isolated from resistant colonies and the identity of the cloned DNA confirmed by restriction analysis, PCR and DNA sequencing.

Clones containing the desired constructs are grown overnight ("O/N") in liquid culture in LB media supplemented with both ampicillin (100 μ g/mL) and kanamycin (25 μ g/mL). The O/N culture is used to inoculate a large

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culture, at a dilution of approximately 1:25 to 1:250. The cells are grown to an optical density at 600 nm ("0D600") of between 0.4 and 0.6. Isopropyl-beta-D-thiogalacto-pyranoside ("IPTG") is then added to a final concentration of 1 mM to induce transcription from the lac repressor sensitive promoter, by inactivating the lacI repressor. Cells subsequently are incubated further for three to four hours. Cells then are harvested by centrifugation.

The cells are then stirred for three to four hours at

4 degrees C in 6 M guanidine hydrochloride, pH 8. The cell
debris is removed by centrifugation, and the supernatant
containing the LP polypeptide is dialyzed against 50 mM
sodium acetate buffer, pH 6, supplemented with 200 mM
sodium chloride. Alternatively, a polypeptide can be

15 successfully refolded by dialyzing it against 500 mM sodium
chloride, 20% glycerol, 25 mM Tris hydrochloride, pH 7.4,
containing protease inhibitors.

If insoluble protein is generated, the protein is made soluble according to known method steps. After renaturation, the polypeptide is purified by ion exchange, hydrophobic interaction, and size exclusion chromatography. Alternatively, an affinity chromatography step such as an antibody column is used to obtain pure LP polypeptide. The purified polypeptide is stored at 4 degrees C or frozen at negative 40 degrees C to negative 120 degrees C.

Example 2

Cloning and Expression of LP polypeptidesin a Baculovirus Expression System

In this example, the plasmid shuttle vector pA2 GP is used to insert the cloned DNA encoding the mature LP polypeptide into a baculovirus, using a baculovirus leader

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and standard methods as described in Summers, et al., A Manual of Methods for Baculovirus Vectors and Insect Cell Culture Procedures, Texas Agricultural Experimental Station Bulletin No. 1555 (1987). This expression vector contains the strong polyhedrin promoter of the Autographa californica nuclear polyhedrosis virus (AcMNPV) followed by the secretory signal peptide (leader) of the baculovirus gp67 polypeptide and convenient restriction sites such as BamHI, Xba I, and Asp718. The polyadenylation site of the simian virus 40 ("SV40") is used for efficient polyadenylation. For easy selection of recombinant virus, the plasmid contains the beta-galactosidase gene from E. coli under control of a weak Drosophila promoter in the same orientation, followed by the polyadenylation signal of the polyhedrin gene. The inserted genes are flanked on both sides by viral sequences for cell-mediated homologous recombination with wild-type viral DNA to generate viable virus that expresses the cloned polynucleotide.

Other baculovirus vectors are used in place of the
vector above, such as pAc373, pVL941 and pAcIM1, as one
skilled in the art would readily appreciate, as long as the
construct provides appropriately located signals for
transcription, translation, secretion and the like,
including a signal peptide and an in-frame AUG as required.

Such vectors are described, for instance, in Luckow, et al.,
Virology 170:31-39.

The cDNA sequence encoding the mature LP polypeptide in a clone, lacking the AUG initiation codon and the naturally associated nucleotide binding site, is amplified using PCR oligonucleotide primers corresponding to the 5' and 3' sequences of the gene. Non-limiting examples include 5' and 3' primers having nucleotides corresponding or complementary

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to a portion of the coding sequence of an LP polypeptide-encoding polynucleotide, e.g., as presented in SEQ ID NO:1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, or 37 according to known method steps.

The amplified fragment is isolated from a 1% agarose gel using a commercially available kit (e.g., "Geneclean," BIO 101 Inc., La Jolla, CA). The fragment is then digested with the appropriate restriction enzyme and again is purified on a 1% agarose gel. This fragment is designated herein "F1."

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The plasmid is digested with the corresponding restriction enzymes and optionally, can be dephosphorylated using calf intestinal phosphatase, using routine procedures known in the art. The DNA is then isolated from a 1% agarose gel using a commercially available kit ("Geneclean" BIO 101 Inc., La Jolla, CA). This vector DNA is designated herein "V1."

Fragment F1 and the dephosphorylated plasmid V1 are ligated together with T4 DNA ligase. E. coli HB101 or other suitable E. coli hosts such as XL-1 Blue (Stratagene Cloning 20 Systems, La Jolla, CA) cells are transformed with the ligation mixture and spread on culture plates. Bacteria are identified that contain the plasmid bearing a human LP polypeptide-encoding polynucleotide using the PCR method, in which one of the primers that is used to amplify the gene 25 and the second primer is from well within the vector so that only those bacterial colonies containing an LP polypeptideencoding polynucleotide will show amplification of the DNA. . The sequence of the cloned fragment is confirmed by DNA sequencing. The resulting plasmid is designated herein as 30 pBacLP.

Five μg of the plasmid pBacLP plasmid construct is cotransfected with 1.0 μg of a commercially available linearized baculovirus DNA ("BaculoGold[®] baculovirus DNA", PharMingen, San Diego, CA), using the lipofection method described by Felgner, et al., Proc. Natl. Acad. Sci. USA 84: 7413-7 (1987). 1 μg of BaculoGold[®] virus DNA and 5 μg of the plasmid pBacLP are mixed in a sterile well of a microtiter plate containing 50 μL of serum-free Grace's medium (Life Technologies, Inc., Rockville, MD).

Afterwards, 10 μL Lipofectin plus 90 μL Grace's medium are added, mixed and incubated for fifteen minutes at room temperature. Then, the transfection mixture is added dropwise to Sf9 insect cells (ATCC CRL 1711) seeded in a 35 mm tissue culture plate with 1 mL Grace's medium without serum. The plate is rocked back and forth to mix the newly added solution. The plate is then incubated for five hours at 27 degrees C. After five hours, the transfection solution is removed from the plate and 1 mL of Grace's insect medium supplemented with 10% fetal calf serum is added. The plate is put back into an incubator and cultivation is continued at 27 degrees C for four days.

After four days, the supernatant is collected, and a plaque assay is performed. An agarose gel with "Blue Gal" (Life Technologies, Inc., Rockville, MD) is used to allow easy identification and isolation of gal-expressing clones, which produce blue-stained plaques. (A detailed description of a "plaque assay" of this type can also be found in the user's guide for insect cell culture and baculovirology distributed by Life Technologies, Inc., Rockville, MD, pp. 9-10). After appropriate incubation, blue stained plaques are picked with a micropipettor tip (e.g., Eppendorf). The agar containing the recombinant

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viruses is then resuspended in a microcentrifuge tube containing 200 μL of Grace's medium and the suspension containing the recombinant baculovirus is used to infect Sf9 cells seeded in 35 mm dishes. Four days later, the supernatants of these culture dishes are harvested and then stored at 4 degrees C.

To verify the expression of the LP polypeptide, Sf9 cells are grown in Grace's medium supplemented with 10% heat-inactivated FBS. The cells are infected with the 10 recombinant baculovirus at a multiplicity of infection ("MOI") of about two. Six hours later, the medium is removed and replaced with SF900 II medium minus methionine and cysteine (available, e.g., from Life Technologies, Inc., Rockville, MD). If radiolabeled polypeptides are desired, 42 hours later, 5 mCi of 35S-methionine and 5 mCi 15 35S-cysteine (available from Amersham, Piscataway, NJ) are added. The cells are further incubated for sixteen hours and then harvested by centrifugation. The polypeptides in the supernatant as well as the intracellular polypeptides 20 are analyzed by SDS-PAGE, followed by autoradiography (if radiolabeled). Microsequencing of the amino acid sequence of the amino terminus of purified polypeptide can be used to determine the amino terminal sequence of the mature polypeptide and, thus, the cleavage point and length of the 25 secretory signal peptide.

Example 3

Cloning and Expression of an LP Polypeptide in Mammalian Cells

A typical mammalian expression vector contains at least one promoter element, which mediates the initiation

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of transcription of mRNA, the polypeptide coding sequence, and signals required for the termination of transcription and polyadenylation of the transcript. Additional elements include enhancers, Kozak sequences and intervening sequences flanked by donor and acceptor sites for RNA splicing. Highly efficient transcription can be achieved with the early and late promoters from SV40, the long terminal repeats (LTRS) from Retroviruses, e.g., RSV, HTLVI, HIVI and the early promoter of the cytomegalovirus (CMV). However, cellular elements can also be used (e.g., 10 the human actin promoter). Suitable expression vectors for use in practicing the present invention include, for example, vectors such as pIRES1neo, pRetro-Off, pRetro-On, PLXSN, or pLNCX (Clontech Labs, Palo Alto, CA), pcDNA3.1 (+/-), pcDNA/Zeo (+/-) or pcDNA3.1/Hygro (+/-)15 (Invitrogen), PSVL and PMSG (Pharmacia, Uppsala, Sweden), pRSVcat (ATCC 37152), pSV2dhfr (ATCC 37146) and pBC12MI (ATCC 67109). Other suitable mammalian host cells include human Hela 293, H9, Jurkat cells, mouse NIH3T3, C127 cells, Cos 1, Cos 7 and CV 1, quail QC1-3 cells, mouse L cells, 20 and Chinese hamster ovary (CHO) cells.

Alternatively, the gene is expressed in stable cell lines that contain the gene integrated into a chromosome. The co-transfection with a selectable marker such as DHRF (dihydrofolate reductase), GPT neomycin, or hygromycin allows the identification and isolation of the transfected cells.

The transfected gene can also be amplified to express large amounts of the encoded polypeptide. The DHFR marker is useful to develop cell lines that carry several hundred or even several thousand copies of the gene of interest. Another useful selection marker is the enzyme glutamine

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synthase (GS) [Murphy, et al., Biochem. J. 227:277-9 (1991); Bebbington, et al., Bio/Technology 10:169-75 (1992)]. Using these markers, the mammalian cells are grown in selective medium and the cells with the highest resistance are selected. These cell lines contain the amplified gene(s) integrated into a chromosome. Chinese hamster ovary (CHO) and NSO cells are often used for the production of polypeptides.

The expression vectors pC1 and pC4 contain the strong
promoter (LTR) of the Rous Sarcoma Virus [Cullen, et al.,
Mol. Cell. Biol. 5:438-47 (1985)] plus a fragment of the
CMV-enhancer [Boshart, et al., Cell 41:521-30 (1985)].
Multiple cloning sites, e.g., with the restriction enzyme
cleavage sites BamHI, XbaI, and Asp718, facilitate the
cloning of the gene of interest. The vectors contain in
addition the 3' intron, the polyadenylation and termination
signal of the rat preproinsulin gene.

Example 3(a)

Cloning and Expression in COS Cells

The expression plasmid, pLP HA, is made by cloning a cDNA encoding LP polypeptide into the expression vector pcDNAI/Amp or pcDNAIII (which can be obtained from Invitrogen, Inc.).

The expression vector pcDNAI/amp contains: (1) an E. coli origin of replication effective for propagation in E. coli and other prokaryotic cells; (2) an ampicillin resistance gene for selection of plasmid-containing prokaryotic cells; (3) an SV40 origin of replication for propagation in eukaryotic cells; (4) a CMV promoter, a polylinker, an SV40 intron; (5) several codons encoding a hemagglutinin fragment (i.e., an "HA" tag to facilitate

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purification) or HIS tag (see, e.g., Ausubel, supra) followed by a termination codon and polyadenylation signal arranged so that a cDNA can be conveniently placed under expression control of the CMV promoter and operably linked to the SV40 intron and the polyadenylation signal by means of restriction sites in the polylinker. The HA tag corresponds to an epitope derived from the influenza hemagglutinin polypeptide described by Wilson, et al., Cell 37:767-8 (1984). The fusion of the HA tag to the target polypeptide allows easy detection and recovery of the recombinant polypeptide with an antibody that recognizes the HA epitope. pcDNAIII contains, in addition, the selectable neomycin marker.

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A DNA fragment encoding the LP polypeptide is cloned into the polylinker region of the vector so that recombinant polypeptide expression is directed by the CMV promoter. The plasmid construction strategy is as follows. The LP polypeptide-encoding cDNA of a clone is amplified using primers that contain convenient restriction sites, much as described above for construction of vectors for expression of LP polypeptides in *E. coli*. Non-limiting examples of suitable primers include those based on the coding sequences presented in SEQ ID NO:1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, and 37.

The PCR amplified DNA fragment and the vector, pcDNAI/Amp, are digested with suitable restriction enzyme(s) and then ligated. The ligation mixture is transformed into *E. coli* strain SURE (available from Stratagene Cloning Systems, La Jolla, CA), and the transformed culture is plated on ampicillin media plates which then are incubated to allow growth of ampicillin resistant colonies. Plasmid DNA is isolated from resistant

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colonies and examined by restriction analysis or other means for the presence of the LP polypeptide-encoding fragment.

For expression of recombinant LP polypeptide, COS cells are transfected with an expression vector, as described above, using DEAE-DEXTRAN, as described, for instance, in Sambrook, et al., Molecular Cloning: a Laboratory Manual, Cold Spring Laboratory Press, Cold Spring Harbor, New York (1989). Cells are incubated under conditions for expression of the LP polypeptide-encoding polynucleotide by the vector.

Expression of the LP polypeptide-HA fusion is detected by radiolabeling and immunoprecipitation, using methods described in, for example Harlow, et al., Antibodies: A Laboratory Manual, 2nd Ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York (1988). To this end, two days after transfection, the cells are labeled by incubation in media containing 35S-cysteine for eight hours. The cells and the media are collected, and the cells are washed and lysed with detergent-containing RIPA buffer: 150 mM sodium chloride, 1% NP-40, 0.1% SDS, 0.5% DOC, 50 mM TRIS, pH 7.5, as described by Wilson, et al., cited above. Proteins are precipitated from the cell lysate and from the culture media using an HA-specific monoclonal antibody. The precipitated polypeptides then are analyzed by SDS-PAGE and autoradiography. An expression product of the expected size is seen in the cell lysate, which is not seen in negative controls.

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Example 3(b)

Cloning and Expression in CHO Cells

The vector pC4 is used for the expression of the LP polypeptide. Plasmid pC4 is a derivative of the plasmid pSV2-dhfr (ATCC Accession No. 37146). The plasmid contains the mouse DHFR gene under control of the SV40 early promoter. Chinese hamster ovary cells or other cells lacking dihydrofolate activity that are transfected with these plasmids can be selected by growing the cells in a selective medium (alpha minus MEM, Life Technologies) 10 supplemented with methotrexate. The amplification of the DHFR genes in cells resistant to methotrexate (MTX) has been well documented [see, e.g., Alt, et al., J. Biol. Chem. 253:1357-70 (1978); Hamlin and Ma, Biochem. et Biophys. Acta 1097:107-43 (1990); and Page and Sydenham, 15 Biotechnology 9:64-8 (1991)]. Cells grown in increasing concentrations of MTX develop resistance to the drug by overproducing the target enzyme, DHFR, as a result of amplification of the DHFR gene. If a second gene is linked to the DHFR gene, it is usually co-amplified and over-20 expressed. It is known in the art that this approach can be used to develop cell lines carrying more than 1,000 copies of the amplified gene(s). Subsequently, when the methotrexate is withdrawn, cell lines are obtained which contain the amplified gene integrated into one or more 25 chromosome(s) of the host cell.

Plasmid pC4 contains for expressing the gene of interest the strong promoter of the long terminal repeat (LTR) of the Rous Sarcoma Virus [Cullen, et al., Mol. Cell. Biol. 5: 438-47 (1985)] plus a fragment isolated from the enhancer of the immediate early gene of human cytomegalovirus (CMV) [Boshart, et al., Cell 41: 521-30 (1985)].

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Downstream of the promoter are BamHI, XbaI, and Asp718 restriction enzyme cleavage sites that allow integration of the genes. Behind these cloning sites, the plasmid contains the 3' intron and polyadenylation site of the rat preproinsulin gene. Other high efficiency promoters can 5 also be used for the expression, e.g., the human beta-actin promoter, the SV40 early or late promoters or the long terminal repeats from other retroviruses, e.g., HIV and HTLVI. Clontech's Tet-Off and Tet-On gene expression systems and similar systems can be used to express the LP 10 polypeptide in a regulated way in mammalian cells [Gossen, and Bujard, Proc. Natl. Acad. Sci. USA 89:5547-51 (1992)]. For the polyadenylation of the mRNA other signals, e.g., from the human growth hormone or globin genes can be used 15 as well. Stable cell lines carrying a gene of interest integrated into the chromosomes can also be selected upon co-transfection with a selectable marker such as gpt, G418 or hygromycin. It is advantageous to use more than one selectable marker in the beginning, e.g., G418 plus 20 methotrexate.

The plasmid pC4 is digested with restriction enzymes and then dephosphorylated using calf intestinal phosphatase by procedures known in the art. The vector is then isolated from a 1% agarose gel.

The DNA sequence encoding the complete the LP polypeptide is amplified using PCR oligonucleotide primers corresponding to the 5' and 3' sequences of the gene. Non-limiting examples include 5' and 3' primers having nucleotides corresponding or complementary to a portion of the coding sequences of an LP polypeptide-encoding polynucleotide, e.g., as presented in SEQ ID NO:1, 3, 5, 7,

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9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, or 37, according to known method steps.

The amplified fragment is digested with suitable endonucleases and then purified again on a 1% agarose gel.

The isolated fragment and the dephosphorylated vector are then ligated with T4 DNA ligase. E. coli HB101 or XL-1 Blue cells are then transformed and bacteria are identified that contain the fragment inserted into plasmid pC4 using, for instance, restriction enzyme analysis.

Chinese hamster ovary (CHO) cells lacking an active DHFR gene are used for transfection. Five μg of the expression plasmid pC4 is cotransfected with 0.5 μg of the plasmid pSV2-neo using lipofectin. The plasmid pSV2-neo contains a dominant selectable marker, the neo gene from Tn5 encoding an enzyme that confers resistance to a group of antibiotics including G418. The cells are seeded in alpha minus MEM supplemented with 1 $\mu g/mL$ G418. After two days, the cells are trypsinized and seeded in hybridoma cloning plates (Greiner, Germany) in alpha minus MEM supplemented with 10, 25, or 50 ng/mL of methotrexate plus 1 $\mu g/mL$ G418. After about ten to fourteen days, single clones are trypsinized and then seeded in six-well petri dishes or 10 mL flasks using different concentrations of methotrexate (50 nM, 100 nM, 200 nM, 400 nM, 800 nM).

Clones growing at the highest concentrations of methotrexate are then transferred to new six-well plates containing even higher concentrations of methotrexate (1 mM, 2 mM, 5 mM, 10 mM, 20 mM). The same procedure is repeated until clones are obtained which grow at a concentration of 100 to 200 mM. Expression of the desired gene product is analyzed, for instance, by SDS-PAGE and Western blot or by reversed phase HPLC analysis.

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$rac{ ext{Example 4}}{ ext{Tissue Distribution of LP polypeptide-encoding mRNA}}$

Northern blot analysis is carried out to examine expression of LP-polypeptide mRNA in human tissues, using methods described by, among others, Sambrook, et al., cited above. A cDNA preferably probe encoding the entire LP polypeptide is labeled with ³²P using the Rediprime[™] DNA

labeling system (Amersham Life Science), according to the manufacturer's instructions. After labeling, the probe is purified using a CHROMA SPIN-100TM column (Clontech

Laboratories, Inc.) according to the manufacturer's protocol number PT1200-1. The purified and labeled probe is used to examine various human tissues for LP polypeptide mRNA.

Multiple Tissue Northern (MTN) blots containing various human tissues (H) or human immune system tissues (IM) are obtained from Clontech and are examined with the labeled probe using ExpressHybTM hybridization solution (Clontech) according to manufacturer's protocol number PT1190-1.

Following hybridization and washing, the blots are mounted, exposed to film at negative 70 degrees C overnight, and developed according to standard procedures.

Example 5

Protein Phosphorylation on Tyrosine Residues

Protein-induced cell responses are determined by monitoring tyrosine phosphorylation upon stimulation of cells by addition of LP polypeptides. This is accomplished in two steps: cell manipulation and immunodetection.

Protein phosphorylation is measured using the SK-N-MC neuroblastoma cell line (ATCC HTB-10). On day one, the cells are plated into poly-D-lysine-coated, 96 well plates

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containing cell propagation medium [DMEM:F12 (3:1), 20 mM HEPES at pH 7.5, 5% FBS, and 50 μ g/mL Gentamicin]. The cells are seeded at a concentration of 20,000 cells per well in 100 μ L medium. On day two, the propagation medium in each well is replaced with 100 μ L starvation medium containing DMEM:F12 (3:1), 20 mM HEPES at pH 7.5, 0.5% FBS, and 50 μ g/mL Gentamicin. The cells are incubated overnight.

On day three, pervanadate solution is made ten minutes before cell lysis; pervanadate is prepared by mixing 100 μ L of sodium orthovanadate (100 mM) and 3.4 μ L of hydrogen peroxide (producing 100% stock pervanadate solution). The lysis buffer is then prepared: 50 mM HEPES at pH 7.5, 150 mM sodium chloride, 10% glycerol, 1% TRITON-X100TM, 1 mM EDTA, 1 mM pervanadate, and BM protease inhibitors. The cells are stimulated by adding 10 μ L of the LP polypeptide solution to the cells, and incubating for ten minutes. Next, the medium is aspirated, and 75 μ L lysis buffer are added to each well. The cells are lysed at 4 degrees C for fifteen minutes, then 25 μ L of 4% loading buffer are added to the cell lysates. The resultant solution is mixed then heated to 95 degrees C.

Detection of tyrosine phosphorylation is accomplished by Western immunoblotting. Twenty microliters of each cell sample are loaded onto SDS-PAGE eight to sixteen percent

25 amino acid-ready gels from Bio-Rad, and the gels are run. The proteins are electrotransferred in transfer buffer

(25 mM Tris base at pH 8.3, 0.2 M glycine, 20% methanol)

from the gel to a nitrocellulose membrane using 250 mA per gel over a one hour period. The membrane is incubated for one hour at ambient conditions in blocking buffer consisting of TBST (20 mM Tris hydrochloride at pH 7.5, 150 mM sodium chloride, 0.1% TWEEN®-20) with 1% BSA.

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Next, the antibodies are added to the membrane. The membrane is incubated overnight at 4 degrees C with gentle rocking in primary antibody solution consisting of the antibody, TBST, and 1% BSA. The next day, the membrane is washed three times, five minutes per wash, with TBST. The membrane is then incubated in the secondary antibody solution consisting of the antibody, TBST, and 1% BSA for one hour at ambient conditions with gentle rocking. After the incubation, the membrane is washed four times with TBST, ten minutes per wash.

Detection is accomplished by incubating the membrane with 10 to 30 mL of SuperSignal Solution for one minute at ambient conditions. After one minute, excess developing solution is removed, and the membrane is wrapped in plastic wrap. The membrane is exposed to X-ray film for twenty second, one minute, and two minute exposures (or longer if needed). The number and intensity of immunostained protein bands are compared to bands for the negative controlstimulated cells (basal level of phosphorylation) by visual comparison.

LP251(a) stimulates phosphorylation in the SK-N-MC cell line and activates the AKT pathway.

Example 6

25 <u>Cell Stimulation with Detection Utilizing Reporters</u>

Protein-induced cell responses are measured using

reporters. The SK-N-MC cell line (neuroblastoma; ATCC HTB
10) / NFKB reporter combination is used.

For the reporter used, positive controls are designed in the form of agonist cocktails. These cocktails include approximate maximal stimulatory doses of several ligands known to stimulate the regulated signal pathway. For the

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NFKB reporter, the NFKB / PkC pathway is stimulated by an agonist cocktail containing LPS and TNF-alpha as positive controls. Cell lines and reporters with no exogenous stimulus added are used as negative controls.

At time zero, the cells are transiently transfected with the reporter plasmids in tissue culture flasks using a standard optimized protocol for all cell lines (see Example 1). After 24 hours, the cells are trypsinized and seeded into 96-well poly-D-lysine coated assay plates at a rate of 20,000 cells per well in growth medium. After four to five hours, the medium is replaced with serum-free growth medium. At that time, stimulants for those reporters which required a 24-hour stimulation period are added. After 48 hours, stimulants for the reporters which required a five-hour stimulation period are added. Five hours later, all conditions are lysed using a lysis/luciferin cocktail, and the fluorescence of the samples is determined using a Micro Beta reader.

Each assay plate is plated to contain four positive control wells, sixteen negative control wells, and sixty-four test sample wells (two replicates of thirty-two test samples). The threshold value for a positive "hit" is a fluorescence signal equal to the mean plus two standard deviations of the negative control wells. Any test sample that, in both replicates, generates a signal above that threshold is defined as a "hit."

LP240 stimulates the NFKB pathway of the neuroblastoma cell line SK-N-MC.

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Example 7

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Cell Proliferation and Cytotoxicity Determination Utilizing Fluorescence Detection

This assay is designed to monitor gross changes in the number of cells remaining in culture after exposure to LP polypeptides for a period of three days. The following cells are used in this assay:

U373MG (astrocytoma cell line, ATCC HTB-22)
T1165 (plastocytoma cell line)
ECV304 endothelial cell line)

Prior to assay, cells are incubated in an appropriate assay medium to produce a sub-optimal growth rate, e.g., a 1:10 or 1:20 dilution of normal culture medium. Cells are grown in T-150 flasks, then harvested by trypsin digestion and replated at 40 to 50% confluence into poly-D-lysine-treated 96-well plates. Cells are only plated into the inner thirty-two wells to prevent edge artifacts due to medium evaporation; the outer wells are filled with buffer alone. Following incubation overnight to stabilize cell recovery, LP polypeptides are added to the appropriate wells. Each polypeptide is assayed in triplicate at two different concentrations, 1X and 0.1X dilution in assay medium. Two controls are also included on each assay plate: assay medium and normal growth medium.

After approximately 72 hours of exposure, the plates are processed to determine the number of viable cells. Plates are spun to increase the attachment of cells to the plate. The medium is then discarded, and 50 μ L of detection buffer is added to each well. The detection buffer consisted on MEM medium containing no phenol red (Gibco) with calcein AM (Molecular Probes) and PLURONIC® F-127

(Molecular Probes), each at a 1:2000 dilution. After incubating the plates in the dark at room temperature for thirty minutes, the fluorescence intensity of each well is measured using a Cytofluor 4000-plate reader (PerSeptive Biosystems). For a given cell type, the larger the fluorescence intensity, the greater the number of cells in the well. To determine the effects on cell growth from each plate, the intensity of each well containing cells stimulated with an LP polypeptide is subtracted from the intensity of the wells containing assay medium only 10 (controls). Thus, a positive number indicated stimulation of cell growth; a negative number indicated a reduction in growth. Additionally, confidence limits at 95 and 90% are calculated from the mean results. Results lying outside the 95% confidence limit are scored as "definite hits." Results 15 lying between the 95 and 90% confidence limits are scored as "maybes." The distinction between definite hits and maybes varied due to intraplate variability; thus, subjective scoring is used as a final determination for "hits."

LP251(a) stimulates the growth of T1165 cells and suppresses the growth of U373MG cells. LP240 stimulates the proliferation of the ECV304 endothelial cell line.

WE CLAIM:

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- Isolated nucleic acid comprising DNA having at least
 90% sequence identity to a polynucleotide selected from the
 group consisting of:
 - (a) a polynucleotide having a nucleotide sequence as shown in SEQ ID NO:1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, or 37;
 - (b) a polynucleotide encoding a polypeptide having the amino acid sequence as shown in SEQ ID NO:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, or 38;
 - (c) a polynucleotide encoding the mature form of a polypeptide having the amino acid sequence as shown in SEQ ID NO:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, or 38;
 - (d) a polynucleotide fragment of a polynucleotide asin (a), (b), or (c); and
 - (e) a polynucleotide having a nucleotide sequence which is complementary to the nucleotide sequence of a polynucleotide as in (a), (b),
 (c), or (d).
- 2. An isolated nucleic acid molecule encoding an polypeptide comprising DNA that hybridizes to the complement of the nucleic acid sequence that encodes LP105, LP061, LP224, LP240, LP239(a), LP243(a), LP243(b), LP253, LP218, LP251(a), LP252, LP239(b), LP223(a), LP255(a), LP244, LP186, LP251(b), LP255(b), LP223(b), or any fragment or variant thereof.

- 3. The isolated nucleic acid molecule of claim 2, wherein hybridization occurs under stringent hybridization and wash conditions.
- 5 4. A vector comprising the nucleic acid molecule of any of Claims 1 to 3.
- 5. The vector of Claim 4, wherein said nucleic acidmolecule is operably linked to control sequences recognized by10 a host cell transformed with the vector.
 - 6. A host cell comprising the vector of Claim 5.
- 7. A process for producing an LP polypeptide comprising culturing the host cell of Claim 6 under conditions suitable for expression of said LP polypeptide and recovering said LP polypeptide from the cell culture.
- 8. An isolated polypeptide comprising an amino acid sequence comprising about 90% sequence identity to a sequence of amino acid residues comprising LP105, LP061, LP224, LP240, LP239(a), LP243(a), LP243(b), LP253, LP218, LP251(a), LP252, LP239(b), LP223(a), LP255(a), LP244, LP186, LP251(b), LP255(b), or LP223(b) as shown in SEQ ID NO:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, or 38, respectively.
 - 9. An isolated polypeptide comprising a sequence of amino acid residues selected from the group consisting of:
 - (a) SEQ ID NO:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, or 38;

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- (b) fragments of (a) sufficient to provide a binding site for an LP polypeptide antibody; and
- (c) variants of (a) or (b).
- 5 10. An isolated polypeptide produced by the method of Claim 7.
 - 11. A chimeric molecule comprising an LP polypeptide fused to a heterologous amino acid sequence.
- 12. The chimeric molecule of Claim 11, wherein said heterologous amino acid sequence is an epitope tag sequence.
- 13. The chimeric molecule of Claim 12, wherein said heterologous amino acid sequence is an Fc region of an immunoglobulin.
 - 14. An antibody which specifically binds to an LP polypeptide.
 - 15. The antibody of Claim 14, where said antibody is a monoclonal antibody.
- 16. The antibody of Claim 15, wherein said antibody is selected from the group consisting of a humanized antibody and a human antibody.
- 17. A composition comprising a therapeutically effective amount of an active agent selected from the group consisting of:
 - (a) an LP polypeptide;
 - (b) an agonist to an LP polypeptide;

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- (c) an antagonist to an LP polypeptide;
- (d) an LP polypeptide antibody;
- (e) an anti-LP polypeptide-encoding mRNA specific ribozyme; and
- (f) a polynucleotide as in Claim 1, in combination with a pharmaceutically acceptable carrier.
- 18. A method of treating a mammal suffering from a disease, condition, or disorder associated with aberrant levels of an LP-polypeptide comprising administering a therapeutically effective amount of an LP polypeptide or LP polypeptide agonist.
- 19. A method of diagnosing a disease, condition, or
 disorder associated with aberrant levels of an LP polypeptide
 by: (1) culturing test cells or tissues expressing LP
 polypeptide; (2) administering a compound which can inhibit LP
 polypeptide modulated signaling; and (3) measuring the LP
 polypeptide-mediated phenotypic effects in the test cells or
 tissues.
 - 20. An article of manufacture comprising a container, label and therapeutically effective amount of the composition of Claim 17.
 - 21. Use of an LP polypeptide in the manufacture of a medicament for the treatment of a disease, condition, or disorder associated with aberrant levels of an LP polypeptide.

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His Tyr Ile Cys Lys Leu Ala Thr His Ala Ile His Leu Val Val Phe 150 145

His Glu Arg Ser Gly Asn Tyr Leu Trp His Gly His Leu Arg Leu Lys 160 165

Glu His Ile Asp Arg Lys Phe Val Pro Phe Gln Lys Leu Gln Phe Gly 185 180 175

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<221> mutation

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Thr Ser Phe Asp Tyr Glu Cys Thr Thr Thr Asp Phe Val Val His Gln 190 195 200

Thr Leu Pro Tyr Gln Ser Val Ser Val Asp Thr Phe Asn Ser Lys Asn 205 210 215

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Tyr Asp Glu Ser Trp Thr Lys Phe Val Lys Phe Gln Asp Ile Glu Val 285 290 295

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Thr	Asp 30	Phe	Ala	Phe	Arg	Leu 35	Tyr	Arg	Arg	Leu	Val 40	Leu	Glu	Thr	Pro
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Gly	Phe	Gln 95	His	Leu	Val	His	Ser 100	Leu	Thr	Val	Pro	Ser 105	Lys	Asp	Lev
Thr	Leu 110	Lys	Met	Gly	Ser	Ala 115	Leu	Phe	Val	Lys	Lys 120	Glu	Leu	Gln	Lev
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Gly Leu Asp Leu Leu Thr Ala Met Val Leu Val Asn His Ile Phe Phe 175 180 185

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Phe Val Leu Gln Met Asp Tyr Lys Gly Asp Ala Val Ala Phe Phe Val 240 245 250

Leu Pro Ser Lys Gly Lys Met Arg Gln Leu Glu Gln Ala Leu Ser Ala 255 260 265

Arg Thr Leu Arg Lys Trp Ser His Ser Leu Gln Lys Arg Trp Ile Glu 270 275 280

Val Phe Ile Pro Arg Phe Ser Ile Ser Ala Ser Tyr Asn Leu Glu Thr 285 290 295 300

Ile Leu Pro Lys Met Gly Ile Gln Asn Val Phe Asp Lys Asn Ala Asp 305 310 315

Phe Ser Gly Ile Ala Lys Arg Asp Ser Leu Gln Val Ser Lys Ala Thr 320 325 330

His Lys Ala Val Leu Asp Val Ser Glu Glu Gly Thr Glu Ala Thr Ala 335 340 345

Ala Thr Thr Thr Lys Phe Ile Val Arg Ser Lys Asp Gly Pro Ser Tyr 350 355 360

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gcc to	tc gcc aa ne Ala Aa	ac ctg sn Leu	ccg cg Pro Ar 40	g Leu	cgc Arg	gtc Val	ctg Leu	cgt Arg 45	ctc Leu	cgt Arg	ggc	aac Asn	242
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gtg Val	cgc Arg	Ġĵħ ĝĝc	gac Asp 325	gcg Ala	ctg Leu	cga Arg	aac Asņ	ctg Leu 330	ccg Pro	gac Asp	tcc Ser	gtg Val	ctg Leu 335	ttc Phe	gag Glu	1106
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				gac Asp												1346
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gcc gcg gcg ggc cag gga ggc gcg cgc aag ttc aac atg aag atg atc Ala Ala Ala Gly Gln Gly Gly Ala Arg Lys Phe Asn Met Lys Met Ile 515 520 525	1682
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. Asp Leu Ser Glu Asn Ala Ile Ala His Val Glu Pro Gly Ala Phe Ala

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- Asp Leu His Ser Leu Arg Arg Leu Glu Val Gly Asp Asn Asp Leu Val 85 90 95 100
- Phe Val Ser Arg Arg Ala Phe Ala Gly Leu Leu Ala Leu Glu Glu Leu 105 110 115
- Thr Leu Glu Arg Cys Asn Leu Thr Ala Leu Ser Gly Glu Ser Leu Gly 120 125 130
- His Leu Arg Ser Leu Gly Ala Leu Arg Leu Arg His Leu Ala Ile Ala 135 140 145
- Ser Leu Glu Asp Gln Asn Phe Arg Arg Leu Pro Gly Leu Leu His Leu 150 155 160
- Glu Ile Asp Asn Trp Pro Leu Leu Glu Glu Val Ala Ala Gly Ser Leu 165 170 175 180
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- Ala Val Pro Ala Ala Ala Leu Arg His Gln Ala His Leu Thr Cys Leu 200 205 210
- Asn Leu Ser His Asn Pro Ile Ser Thr Val Pro Arg Gly Ser Phe Arg 215 220 225
- Asp Leu Val Arg Leu Arg Glu Leu His Leu Ala Gly Ala Leu Leu Ala 230 235 240
- Val Val Glu Pro Gln Ala Phe Leu Gly Leu Arg Gln Ile Arg Leu Leu 245 250 255 260
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Ser Val Asn Thr Leu Glu Thr Leu Arg Val Asp Gly Asn Pro Leu Ala 280 285 290

Cys Asp Cys Arg Leu Leu Trp Ile Val Gln Arg Arg Lys Thr Leu Asn 295 300 305

Phe Asp Gly Arg Leu Pro Ala Cys Ala Thr Pro Ala Glu Val Arg Gly 310 315 320

Asp Ala Leu Arg Asn Leu Pro Asp Ser Val Leu Phe Glu Tyr Phe Val 325 330 335 340

Cys Arg Lys Pro Lys Ile Arg Glu Arg Arg Leu Gln Arg Val Thr Ala 345 350 355

Thr Ala Gly Glu Asp Val Arg Phe Leu Cys Arg Ala Glu Gly Glu Pro 360 365 370

Ala Pro Thr Val Ala Trp Val Thr Pro Gln His Arg Pro Val Thr Ala 375 380 385

Thr Ser Ala Gly Arg Ala Arg Val Leu Pro Gly Gly Thr Leu Glu Ile 390 395 400

Gln Asp Ala Arg Pro Gln Asp Ser Gly Thr Tyr Thr Cys Val Ala Ser 405 410 415 420

Asn Ala Gly Gly Asn Asp Thr Tyr Phe Ala Thr Leu Thr Val Arg Pro 425 430 435

Glu Pro Ala Ala Asn Arg Thr Pro Gly Glu Ala His Asn Glu Thr Leu 440 445 450

Ala Ala Leu Arg Ala Pro Leu Asp Leu Thr Thr Ile Leu Val Ser Thr 455 460 465

Ala Met Gly Cys Ile Thr Phe Leu Gly Val Val Leu Phe Cys Phe Val 470 475 480

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					gtc Val									441
					aag Lys									489
					cgc Arg									537
					gcc Ala 145									585
					aac Asn									633
					ggc gly									681
					aac Asn									729
					ccg Pro									777
					acc Thr 225									825
					gcg Ala									873
					ccc Pro									921
-	_	_	_	_	cgc Arg	-	-	_	-		_	_	_	969

					gcc Ala											1017
					ctg Leu 305											1065
					gag Glu											1113
					ctg Leu											1161
					ccg Pro											1209
					ctg Leu										gtg Val	1257
					atc Ile 385											1305
					gtc Val											1353
					tgg Trp									Thr		1401
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Glu Glu Leu Asp Leu Ser Glu Asn Ala Ile Ala His Val Glu Pro Gly
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Ala Phe Ala Asn Leu Pro Arg Leu Arg Val Leu Arg Leu Arg Gly Asn 75 80 85

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Thr Leu Leu Asp Leu Ser Glu Asn Lys Leu Val Ile Leu Leu Asp Tyr 105 110 115 120

Thr Phe Gln Asp Leu His Ser Leu Arg Arg Leu Glu Val Gly Asp Asn 125 130 135

Asp Leu Val Phe Val Ser Arg Arg Ala Phe Ala Gly Leu Leu Ala Leu 140 145 150

Glu Glu Leu Thr Leu Glu Arg Cys Asn Leu Thr Ala Leu Ser Gly Glu 155 160 165

Ser Leu Gly His Leu Arg Ser Leu Gly Ala Leu Arg Leu Arg His Leu 170 175 180

Ala Ile Ala Ser Leu Glu Asp Gln Asn Phe Arg Arg Leu Pro Gly Leu 185 190 195 200

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Gly Ser Leu Arg Gly Leu Asn Leu Thr Ser Leu Ser Val Thr His Thr 220 225 230

Asn Ile Thr Ala Val Pro Ala Ala Ala Leu Arg His Gln Ala His Leu 235 240 245 Thr Cys Leu Asn Leu Ser His Asn Pro Ile Ser Thr Val Pro Arg Gly 250 255 260

Ser Phe Arg Asp Leu Val Arg Leu Arg Glu Leu His Leu Ala Gly Ala 265 270 275 280

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Arg Leu Leu Asn Leu Ser Asn Asn Leu Leu Ser Thr Leu Glu Glu Ser 300 305 310

Thr Phe His Ser Val Asn Thr Leu Glu Thr Leu Arg Val Asp Gly Asn 315 320 325

Pro Leu Ala Cys Asp Cys Arg Leu Leu Trp Ile Val Gln Arg Arg Lys 330 335 340

Thr Leu Asn Phe Asp Gly Arg Leu Pro Ala Cys Ala Thr Pro Ala Glu 345 350 355 360

Val Arg Gly Asp Ala Leu Arg Asn Leu Pro Asp Ser Val Leu Phe Glu 365 370 375

Tyr Phe Val Cys Arg Lys Pro Lys Ile Arg Glu Arg Arg Leu Gln Arg 380 385 390

Val Thr Ala Thr Ala Gly Glu Asp Val Arg Phe Leu Cys Arg Ala Glu 395 400 405

Gly Glu Pro Ala Pro Thr Val Ala Trp Val Thr Pro Gln His Arg Pro 410 415 420

Val Thr Ala Thr Ser Ala Gly Arg Ala Arg Val Leu Pro Gly Gly Thr 425 430 435 440

Leu Glu Ile Gln Asp Ala Arg Pro Gln Asp Ser Gly Thr Tyr Thr Cys
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cag Gln	cat His	ggc Gly	tcg Ser 185	Asp	atg Met	cag Gln	gtg Val	Ser 190	Phe	gac Asp	cat His	gca Ala	ccg Pro 195	HIS	aac Asn		672
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catceteect gaaacgegae c atg etg tta agg gge gte etc etg geg ttg
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Gln Ala Leu Gln Leu Ala Gly Ala Leu Asp Leu Pro Ala Gly Ser Cys
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gcc ttt gaa gag agc act tgc ggc ttt gac tcc gtg ttg gcc tct ctg
Ala Phe Glu Glu Ser Thr Cys Gly Phe Asp Ser Val Leu Ala Ser Leu
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Pro Trp Ile Leu Asn Glu Glu'Gly His Tyr Ile Tyr Val Asp Thr Ser
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Phe Gly Lys Gln Gly Glu Lys Ala Val Leu Leu Ser Pro Asp Leu Gln
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atc Ile	tcc Ser	ccg Pro	ttg Leu 220	acc Thr	acg Thr	gcc Ala	ccc Pro	atg Met 225	Ala	ggc	tgc Cys	ctg Leu	tca Ser 230	Phe	tat Tyr	1071
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gat Asp	gtg Val 250	Ala	ggc Gly	ctt Leu	tac Tyr	gag Glu 255	Glu	ato Ile	tgg Trp	aaa Lys	gca Ala 260	Asp	agg Arg	r cca	o GJA r aaa	1167
aat Asn 265	Ala	gcc Ala	tgg Trp	Asn	ctt Leu 270	Ala	gag Glu	gtc Val	gag Glu	tto Phe 275	: Asr	gct Ala	cct Pro	tac Tyi	Pro 280	1215
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Ile Thr Phe Ly	g aag ccc atg s Lys Pro Met 445	Pro Thr Ly	ng gtg gtt ttc atg vs Val Val Phe Met 50	agc cta 1743 Ser Leu 455
tgc aaa agt tt Cys Lys Ser Pr 46	ne Trp Asp Cys	ggg ctt gt Gly Leu Va 465	ta gcc ctg gat gad al Ala Leu Asp Asp 470	TIE IIII
ata caa ttg gg Ile Gln Leu Gl 475	ga agc tgc tca Ly Ser Cys Ser	tct tca ga Ser Ser Gl 480	ag aaa ctt cca cct lu Lys Leu Pro Pro 485	cca cct 1839 Pro Pro
gga gag tgt ac Gly Glu Cys Tl 490	ct ttc gag caa nr Phe Glu Glr 495	ı Asp Glu Cy	yt aca ttt act cag ys Thr Phe Thr Glr 500	g gag aaa 1887 n Glu Lys

aga Arg 505	aac Asn	cgg Arg	agc Ser	agc Ser	tgg Trp 510	cac His	agg Arg	agg Arg	agg Arg	gga Gly 515	gaa Glu	act Thr	ccc Pro	act Thr	tcc Ser 520	1935
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tac Tyr	att Ile	gag Glu	gcc Ala 540	tcc Ser	cat His	atg Met	gtg Val	tat Tyr 545	gga Gly	caa Gln	aaa Lys	gca Ala	cgc Arg 550	ctc Leu	ttg Leu	2031
tcc Ser	agg Arg	cct Pro 555	ctg Leu	cga Arg	gga Gly	gtc Val	tct Ser 560	gga Gly	aaa Lys	cac His	tgc Cys	ttg Leu 565	acc Thr	ttt Phe	ttc Phe	2079
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aag Lys 585	gaa Glu	gaa Glu	gac Asp	agt Ser	gaa Glu 590	gag Glu	tcc Ser	ctc Leu	tta Leu	tgg Trp 595	Arg	aga Arg	aga Arg	ggt Gly	gaa Glu 600	2175 ·
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caa Gln	cac His	cag Gln	ata Ile 620	att Ile	ttt Phe	gaa Glu	gcc Ala	att Ile 625	Arg	gga Gly	gta Val	tca Ser	ata Ile 630	Arg	agt Ser	2271
gat Asp	att Ile	gcc Ala 635	Ile	gat Asp	gat Asp	gtt Val	aaa Lys 640	Phe	cag Gln	gca Ala	ı gga	Pro 645	Сув	: gga	ı gaa _{7.} Glu	2319
atg Met	gaa Glu 650	Asp	aca Thr	act Thr	caa Gln	caa Gln 655	Ser	tca Ser	gga Gly	tat Tyi	tet Ser 660	: GTI	g gad 1 As <u>r</u>	tta Le	a aat 1 Asn	2367
Glu 665	ı Ile	: G1:	ι Туг	•	ıgaaa											2419
aco	etcto	ttc	aato	aaaa	itg a	aaaa	aaag	jc aa	aatga	ata	c tg	gaca	gtct	taa	caattta	2479
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Lys Ala Val Leu Leu Ser Pro Asp Leu Gln Ala Glu Glu Trp Ser Cys 50 55 60

Leu Arg Leu Val Tyr Gln Ile Thr Thr Ser Ser Glu Ser Leu Ser Asp 65 70 75

Pro Ser Gln Leu Asn Leu Tyr Met Arg Phe Glu Asp Glu Ser Phe Asp 80 85 . 90

Arg Leu Leu Trp Ser Ala Lys Glu Pro Ser Asp Ser Trp Leu Ile Ala 95 100 105 110

Ser Leu Asp Leu Gln Asn Ser Ser Lys Lys Phe Lys Ile Leu Ile Glu 115 120 125

Gly Val Leu Gly Gln Gly Asn Thr Ala Ser Ile Ala Leu Phe Glu Ile 130 135 140

Lys Met Thr Thr Gly Tyr Cys Ile Glu Cys Asp Phe Glu Glu Asn His 145 150 155

Leu Cys Gly Phe Val Asn Arg Trp Asn Pro Asn Val Asn Trp Phe Val 160 165 170

Gly Gly Gly Ser Ile Arg Asn Val His Ser Ile Leu Pro Gln Asp His 175 180 185 190

Thr Phe Lys Ser Glu Leu Gly His Tyr Met Tyr Val Asp Ser Val Tyr 195 200 205

Val Lys His Phe Gln Glu Val Ala Gln Leu Ile Ser Pro Leu Thr Thr 210 215 220

Ala Pro Met Ala Gly Cys Leu Ser Phe Tyr Tyr Gln Ile Gln Gln Gly 225 230 235

Asn Asp Asn Val Phe Ser Leu Tyr Thr Arg Asp Val Ala Gly Leu Tyr 240 245 250

Glu Glu Ile Trp Lys Ala Asp Arg Pro Gly Asn Ala Ala Trp Asn Leu 255 260 265 270

Ala Glu Val Glu Phe Asn Ala Pro Tyr Pro Met Glu Val Ile Phe Glu 275 280 285

Val Ala Phe Asn Gly Pro Lys Gly Gly Tyr Val Ala Leu Asp Asp Ile 290 295 300

Ser Phe Ser Pro Val His Cys Gln Asn Gln Thr Glu Leu Leu Phe Ser 305 310 315

Ala Val Glu Ala Ser Cys Asn Phe Glu Gln Asp Leu Cys Asn Phe Tyr 320 325 330

Gln Asp Lys Glu Gly Pro Gly Trp Thr Arg Val Lys Val Lys Pro Asn 345 340 345

Met Tyr Arg Ala Gly Asp His Thr Thr Gly Leu Gly Tyr Tyr Leu Leu 355 360 365

Ala Asn Thr Lys Phe Thr Ser Gln Pro Gly Tyr Ile Gly Arg Leu Tyr 370 380

Gly Pro Ser Leu Pro Gly Asn Leu Gln Tyr Cys Leu Arg Phe His Tyr 385 390 395

Ala Ile Tyr Gly Phe Leu Lys Met Ser Asp Thr Leu Ala Val Tyr Ile 400 405 410

Phe Glu Glu Asn His Val Val Gln Glu Lys Ile Trp Ser Val Leu Glu 415 420 425 430

Ser Pro Arg Gly Val Trp Met Gln Ala Glu Ile Thr Phe Lys Lys Pro 435 440 445

Met Pro Thr Lys Val Val Phe Met Ser Leu Cys Lys Ser Phe Trp Asp 450 455 460

Cys Gly Leu Val Ala Leu Asp Asp Ile Thr Ile Gln Leu Gly Ser Cys 465 470 475

Ser Ser Ser Glu Lys Leu Pro Pro Pro Pro Gly Glu Cys Thr Phe Glu 480 485 490

Gln Asp Glu Cys Thr Phe Thr Gln Glu Lys Arg Asn Arg Ser Ser Trp 495 500 505 510

His Arg Arg Arg Gly Glu Thr Pro Thr Ser Tyr Thr Gly Pro Lys Gly 515 520 525

Asp His Thr Gly Val Gly Tyr Tyr Met Tyr Ile Glu Ala Ser His 530 540

Met Val Tyr Gly Gln Lys Ala Arg Leu Leu Ser Arg Pro Leu Arg Gly 545 555

Val Ser Gly Lys His Cys Leu Thr Phe Phe Tyr His Met Tyr Gly Gly 560 565 570

Gly Thr Gly Leu Leu Ser Val Tyr Leu Lys Lys Glu Glu Asp Ser Glu 575 580 585

Glu Ser Leu Leu Trp Arg Arg Arg Gly Glu Gln Ser Ile Ser Trp Leu 595 600 605

Arg Ala	Leu	610	GIU	TYT	ser	cys	615	ALY	GIII	1113	0211	620				
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atatttt	gtt	ccaa	a ato Med	t Al	a tc a Se	t ta r Ty	c ct r Le	t ta u Ty -1	r GI	ra gt .y Va	a ct al Le	c tt eu Ph	t go ne Al -1	.a va.	: ; L	171
ggc cto	c tgt ı Cys	gct Ala -5	cca Pro	atc Ile	tac Tyr	tgt Cys -1	gtg Val	tco Ser	ccg Pro	g gto Val	c aat L Asr 5	gco n Ala	c ccc a Pro	agt Ser		219
gca tac Ala Tyr 10	c ccc r Pro	cgc Arg	cct Pro	tcc Ser	Ser 15	aca Thr	aag Lys	g ago s Sei	c acc	c cct Pro 20	c gcc	c tca a Sei	a caq r Glı	g gtg n Val		267
tat tco Tyr Se: 25	c cto r Leu	aac 1 Asn	acc Thr	gac Asp 30	ttt	gco Ala	tto Phe	c cgo	c cta g Let 35	a tac	c cge	c agg	g cto	g gtt u Val 40		315

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Le	y gag u Glu	Thr	Pro	Ser 45	Gln	Asn	Ile	Phe	Phe 50	Ser	Pro	Val	Ser	Val 55	Ser	
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ga Gl	a gca u Ala	ı gaa ı Glu	gtc Val 140	ttt Phe	tct Ser	aca Thr	gat Asp	ttc Phe 145	tcc Ser	aac Asn	ccc Pro	tcc Ser	att Ile 150	gcc Ala	cag Gln	651
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ga G:	ag cte Lu Le	g aac u Asr 235	Cys	ttt Phe	gtg Val	ctg Leu	cag Gln 240	Met	gat Asp	tac Tyr	aag Lys	7.gga Gly 245	Ası	gco Ala	gtg a Val	939

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gcc Ala 265	ttg Leu	tca Ser	gcc Ala	aga Arg	aca Thr 270	ctg Leu	ata Ile	aag Lys	tgg Trp	agc Ser 275	cac His	tca Ser	ctc Leu	cag Gln	aaa Lys 280	1035
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			Lys			gtgg	gaa	atgg	cctg	tt a	actg	atgg	jc ac	attg	rctaa	1426
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Thr Asp Phe Ala Phe Arg Leu Tyr Arg Arg Leu Val Leu Glu Thr Pro 30 35 40

Ser Gln Asn Ile Phe Phe Ser Pro Val Ser Val Ser Thr Ser Leu Ala 45 50 55 60

Met Leu Ser Leu Gly Ala His Ser Val Thr Lys Thr Gln Ile Leu Gln 65 70 75

Gly Leu Gly Phe Asn Leu Thr His Thr Pro Glu Ser Ala Ile His Gln 80 85 90

Gly Phe Gln His Leu Val His Ser Leu Thr Val Pro Ser Lys Asp Leu 95 100 105 .

Thr Leu Lys Met Gly Ser Ala Leu Phe Val Lys Lys Glu Leu Gln Leu 110 115 120

Gln Ala Asn Phe Leu Gly Asn Val Lys Arg Leu Tyr Glu Ala Glu Val 125 130 135 140

Phe Ser Thr Asp Phe Ser Asn Pro Ser Ile Ala Gln Ala Arg Ile Asn 145 150 155

Ser His Val Lys Lys Lys Thr Gln Gly Lys Val Val Asp Ile Ile Gln 160 165 170

Gly Leu Asp Leu Leu Thr Ala Met Val Leu Val Asn His Ile Phe Phe 175 180 185

Lys Ala Lys Trp Glu Lys Pro Phe His Leu Glu Tyr Thr Arg Lys Asn 190 195 200

Phe Pro Phe Leu Val Gly Glu Gln Val Thr Val Gln Val Pro Met Met 205 210 215 220

His Gln Lys Glu Gln Phe Ala Phe Gly Val Asp Thr Glu Leu Asn Cys 225. 230 235

Phe Val Leu Gln Met Asp Tyr Lys Gly Asp Ala Val Ala Phe Phe Val 240 245 250

Leu Pro Ser Lys Gly Lys Met Arg Gln Leu Glu Gln Ala Leu Ser Ala 255 260 265

Arg Thr Leu Ile Lys Trp Ser His Ser Leu Gln Lys Arg Trp Ile Glu 270 275 280

Val Phe Ile Pro Arg Phe Ser Ile Ser Ala Ser Tyr Asn Leu Glu Thr 285 290 295 300

Ile Leu Pro Lys Met Gly Ile Gln Asn Ala Phe Asp Lys Asn Ala Asp 305 310 315

Phe Ser Gly Ile Ala Lys Arg Asp Ser Leu Gln Val Ser Lys Ala Thr 320 325 330

His Lys Ala Val Leu Asp Val Ser Glu Glu Gly Thr Glu Ala Thr Ala 335 340 345

Ala Thr Thr Thr Lys Phe Ile Val Arg Ser Lys Asp Gly Pro Ser Tyr 350 355 360

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ccc gg Pro Gl	ıt agg .y Arg	atg Met -20	tcc Ser	tgg Trp	ccc Pro	cac His	ggg Gly -15	gca Ala	ttg Leu	ctc Leu	ttc Phe	ctc Leu -10	tgg Trp	ctc Leu	219
ttc to	cc cca er Pro -5	ccc Pro	ctg Leu	G1A aaa	gcc Ala -1	ggt Gly 1	gga Gly	ggt Gly	gga Gly	gtg Val 5	gcc Ala	gtg Val	acg Thr	tct Ser	267
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tcc to Ser Cy	gc agc vs Ser	aac Asn	cag Gln 30	gcc Ala	agc Ser	cgg	gtg Val	atc Ile 35	tgc Cys	aca Thr	cgg Arg	aga Arg	gac Asp 40	ctg Leu	363
gcc ga	ag gtc Lu Val	cca Pro 45	gcc Ala	agc Ser	atc Ile	ccg Pro	gtc Val 50	aac Asn	acg Thr	cgg Arg	tac Tyr	ctg Leu 55	aac Asn	ctg Leu	411
caa ga Gln G	ag aac Lu Asn 60	ggc Gly	atc Ile	cag Gln	gtg Val	atc Ile 65	cgg Arg	acg Thr	gac Asp	acg Thr	ttc Phe 70	aag Lýs	cac His	ctg Leu	. 459
cgg ca Arg H	ac ctg is Leu 5	gag Glu	att Ile	ctg Leu	cag Gln 80	ctg Leu	agc Ser	aag Lys	aac Asn	ctg Leu 85	gtg Val	cgc Arg	aag Lys	atc Ile	507

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Cys Leu Ser Pro Arg Gly Pro Ser Ser Ala Thr Thr Gly Cys Leu Val 235 240 245

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Thr Val Trp Ser Ala His Ser Gly Pro Thr Arg Met Ala Thr Ala Ile 385 390

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Arg Gln Thr Gln Leu Arg Glu Leu Tyr Leu Gln His Asn Gln Leu Thr 170 175 180

Asp Ser Gly Leu Asp Ala Thr Thr Phe Ser Lys Leu His Ser Leu Glu 185 190 195

Tyr Leu Asp Leu Ser His Asn Gln Leu Thr Thr Val Pro Ala Gly Leu 200 . 205 210 215

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Arg Pro Leu Arg Gly Leu His Thr Leu His Leu Tyr Gly Asn Gly Leu 265 270 275

Asp Arg Val Pro Pro Ala Leu Pro Arg Arg Leu Arg Ala Leu Val Leu 280 285 290 295

Pro His Asn His Val Ala Ala Leu Gly Ala Arg Asp Leu Val Ala Thr 300 305 310

Pro Gly Leu Thr Glu Leu Asn Leu Ala Tyr Asn Arg Leu Ala Ser Ala 315 320 325

Arg Val His His Arg Ala Phe Arg Arg Leu Arg Ala Leu Arg Ser Leu 330 335 340

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Pro Glu Pro Leu Ala Gly Leu Asp Gln Leu Arg Glu Leu Ser Leu Ala 380 385 390	
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Pro Pro Thr Asp Leu Asn Val Thr Glu Gly Met Ala Ala Glu Leu Lys 335 340 345

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His Gly Pro Thr Arg Thr Val Glu Ile Ile Asn Val Glu Asp Glu Leu 575 580 585

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